

Alpha-synuclein in the nucleus accumbens induces changes in cocaine behaviour in rats

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Abstract

The mesolimbic dopaminergic system is widely recognized to be critical to the neurobiology of cocaine reward and addiction. The neuronal protein, α -synuclein, is an important regulator in dopaminergic transmission. It interacts with the dopamine transporter, and regulates dopaminergic content, neurotransmission and synaptic strength of dopaminergic neurons. Alpha-synuclein levels are elevated in midbrain dopamine neurons of chronic cocaine abusers, and its expression is increased in psychostimulant-treated animals [M.S. Brenz-Verca *et al.* (2003) *J. Neurosci.*, 18, 1923–1938]. This suggests a role for α -synuclein in psychostimulant-induced behavioural effects. To investigate this hypothesis, we tested the effect of stimulation and silencing of α -synuclein expression in the nucleus accumbens (NAcc) on two cocaine-induced behavioural effects in the rat. For this purpose, animals were administered with lentiviruses driving α -synuclein overexpression under the control of a doxycycline regulatable promoter and/or with three lentiviruses expressing target-specific siRNAs, aimed at silencing α -synuclein mRNA expression. Animals were then tested for cocaine-induced locomotion (15 mg/kg i.p.) or cocaine-induced intravenous self-administration (SA; 0.7 mg/kg, 1 h/day). Overexpression of α -synuclein in the NAcc induced a 45% increase in locomotor activity and a 1.9-fold increase of cocaine SA, which could be abolished when the same animal was fed doxycycline. Furthermore, local inhibition of α -synuclein in the NAcc resulted in significant hypolocomotion activity and a decrease in SA. Our results demonstrate that α -synuclein is able to modulate cocaine-induced behavioural effects. This suggests that targeting α -synuclein function could provide new therapeutic strategies to treat cocaine abuse, for which there is no available treatment.

Introduction

Drug addiction is a chronic illness that remains a major public health problem. Among drugs of abuse, cocaine is one of the most addictive substances in use today. It binds to the dopamine transporter (DAT) and blocks the presynaptic uptake of dopamine (DA), mediating the behavioural and reinforcing effects that contribute to its abuse liability. Analysis of the DAT-binding sites shows an upregulation in *post mortem* brains of cocaine addicts and in acutely abstinent cocaine-dependent individuals (Lee *et al.*, 2001; Mash *et al.*, 2002). Therefore, long-term cocaine abuse leads to neuroadaptive changes in DA homeostasis and in the mesolimbic dopaminergic system, the critical pathway mediating addiction to psychostimulant and reinforcement.

The synaptic protein, α -synuclein, is an important regulator in dopaminergic transmission (Abeliovich *et al.*, 2000; Mosharov *et al.*, 2006). Synucleins are small, soluble proteins expressed primarily in neural tissue and in certain tumours. The family includes three known proteins: α -synuclein, β -synuclein and γ -synuclein (George, 2001; Li *et al.*, 2002). α -Synuclein plays a role on synaptic vesicular transport and synaptic plasticity (Bennett *et al.*, 1999; Murphy *et al.*, 2000), and has a fundamental function in the genesis of neurodegenerative disorders (Cooper *et al.*, 2006). It is a major component of Lewy body in sporadic forms of Parkinson's disease, and its overexpression is implicated in the aetiology of Parkinson's and Alzheimer's diseases (Lotharius & Brundin, 2002). Two mutant forms (A53T and A30P) are

linked to the familiar form of Parkinson's disease (Polymeropoulos *et al.*, 1997; Kruger *et al.*, 1998; George, 2001; Goedert, 2001). Furthermore, it is highly expressed in dopaminergic neurons (Abeliovich *et al.*, 2000; Perez & Hastings, 2004; Yavich *et al.*, 2004) and is involved in DA homeostasis, downregulating the activity of tyrosine hydroxylase and other DA synthesis genes, such as GTP cyclohydroxylase (Sidhu *et al.*, 2004a). α -Synuclein also binds to the DAT and serotonin transporters, and affects their activities and their recruitment to the plasma membrane (Wersinger & Sidhu, 2003, 2005; Wersinger *et al.*, 2003, 2006). The shuttling of DAT to and away from the plasma membrane is modified by the action of α -synuclein, thereby changing the efficiency of DA uptake at the nerve terminals (Sidhu *et al.*, 2004b).

We have described α - and γ -synuclein overexpression in the mesolimbic dopaminergic pathway upon psychostimulants (Brenz-Verca *et al.*, 2003). Furthermore, an overexpression of α -synuclein in DA neurons is observed in cocaine abusers (Mash *et al.*, 2003) or following methamphetamine administration (Maucelli *et al.*, 2006). α -Synuclein protein was increased in striatal synaptosomes from cocaine users compared with age-matched drug-free controls (Yujing *et al.*, 2005). On the other hand, an increase in expression of α -synuclein mRNA in alcohol-dependent patients within the withdrawal state has been observed and has been associated with an increase in alcohol craving (Weiss *et al.*, 1993; Liang *et al.*, 2003; Bonsch *et al.*, 2004, 2005). Interestingly, α -synuclein and DA metabolism play convergent roles in nigrostriatal toxicity (Fornai *et al.*, 2006), and α -synuclein levels are elevated in midbrain DA neurons of chronic cocaine abusers (Mash *et al.*, 2003) and are

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associated with methamphetamine psychosis and dependence (Kobayashi *et al.*, 2004), whereas α -synuclein levels are significantly increased in the amygdala and striatum/accumbens (but not in the mesencephalon) of morphine-withdrawn mice (Ziolkowska *et al.*, 2005). Taken together all these findings suggest that α -synuclein plays a key role in addiction, regulating the DA-signalling pathway. Co-ordinate transcriptional regulation of DA synthesis genes by α -synuclein has been described (Baptista *et al.*, 2003).

The aim of the present study was to investigate the role of α -synuclein in the rewarding effects of chronic cocaine administration. For this purpose, a lentivirus driving the rat α -synuclein overexpression under the control of a tetracycline regulatable promoter was constructed. Furthermore, three lentiviruses expressing α -synuclein-targeted siRNAs, aimed at silencing α -synuclein mRNA expression, were also constructed. These viruses were tested *in vitro* and served for *in vivo* gene transfer, by means of stereotaxic injections into the nucleus accumbens (NAcc) of rat brains. The impact of α -synuclein expression on chronic cocaine administration was assessed, using locomotor activity and self-administration (SA) paradigms. Overexpression of α -synuclein in the NAcc induced a 45% increase in locomotor activity and a 1.9-fold increase of cocaine SA, which could be abolished when the same animal was fed doxycycline. Furthermore, local inhibition of α -synuclein in the NAcc – efficiently achieved *in vivo* using lentiviruses expressing α -synuclein-specific siRNAs – resulted in significant hypolocomotion activity and a decrease in SA.

Materials and methods

Ex vivo methods

Lentivirus constructions

Construction of Lenti- α -Synuclein (Lenti- α Syn). The α -synuclein DNA (GenBank accession no. AF007758) was polymerase chain reaction (PCR) amplified using the pGEMT-rat-Alpha-Synuclein as template and the following primers: 5'-CGC **GGA TCC GCG ATG GAT GTG TTC ATG AAA-3'** and 5'-CCG **CTC GAG CGG TTA ATG ATG ATG ATG ATG GGC TTC AGG CTC ATA GTC T-3'**. The forward primer contains a *BamH-I* restriction site followed by the 5' rat α -synuclein DNA-specific sequence; the reverse primer contains the 3' rat α -synuclein DNA-specific sequence, the 6-Histidine epitope sequence (in bold), a stop codon and a *Xho-I* restriction site. The PCR product was digested with *BamH-I* and *Xho-I*, and cloned into *BamH-I/Xho-I* restriction sites into pTK431 (Bahi *et al.*, 2005a) that expresses the gene of interest under control of a Tet-Off promoter. The green fluorescent protein (GFP) expression vector pTK433 (Bahi *et al.*, 2004a,b) was used as a control vector.

Construction of Lenti- α -synuclein-siRNAs (Lenti- α Syn-siRNA). To silence α -synuclein expression *in vitro* and *in vivo*, three targets were designed, according to the α -synuclein mRNA sequence. The following targets within the α -synuclein sequence were selected, based on Hannon's design criterion (<http://katahdin.cshl.org:9331/RNAi/html/rnai.html>): first target: bp 36–52; second target: bp 305–320; third target: bp 406–423. To each oligo, a *Xho-I* restriction site was added at 3', and a U6-3'-specific 10mer at 5'. Using the pSilencer 1.0-U6 (Ambion, UK) as a template and a U6 promoter-specific forward primer containing *BamH-I* restriction site (5'-GCG **GAT CCC GCT CTA GAA CTA GTG C-3'**), each siRNA target was added to the mouse U6 promoter by PCR, using the following PCR program: 120 s at 94 °C (initial denaturation) followed by 35 cycles (45 s at 94 °C, 45 s at 64 °C and 45 s at 72 °C) in 4% dimethyl sulphoxide (Sigma,

Switzerland). The PCR product was digested with *BamH-I* and *Xho-I*, cloned into similar sites into pTK431, and sequenced to verify the integrity of each construct.

Lentivirus production

The vector plasmids (either pTK431- α -Synuclein, pTK433-GFP or pTK431-U6- α -Syn-siRNAs), together with the packaging construct plasmid p-NRF and the envelope plasmid pMDG-VSV-G were co-transfected into human embryonic kidney (HEK)293T cells to produce the viral particles (Bahi *et al.*, 2004a,b). The viral titres were determined by p24 antigen measurements (KPL, USA). For the *in vivo* experiments, the different viral stocks were matched for viral particle content and used at 0.2 mg/mL of p24.

In vitro α -synuclein assays

The efficiency of the Lenti- α Syn was tested *in vitro* by infection of HEK293T cells. 1×10^5 HEK293T cells were plated. The next day, lentiviruses stocks were mixed with 4 μ g/mL Polybrene (Sigma, Switzerland) with or without 30 ng/mL doxycycline (Sigma), incubated for 30 min at room temperature, added to the cells and incubated at 37 °C. After 24 h the medium was replaced with normal growth medium (with or without 30 ng/mL doxycycline) and cells were left for a further 48 h. Cells were then collected and used for total RNA isolation (for real-time PCR) or for total protein isolation (Western blot). Infections with the different lentiviruses were done as follows: (a) doxycycline regulation: cells were co-infected using 0, 2, 4 or 8 μ L of Lenti- α Syn stock, and one part of the cells was incubated in the presence of 30 ng/mL doxycycline and the other part without doxycycline; (b) *in vitro* silencing of α -synuclein: cells were infected with 4 μ L of Lenti- α Syn stock, either alone or together with 2 μ L of Lenti- α Syn-siRNA T1, 2 μ L of Lenti- α Syn-siRNA T2, 2 μ L of Lenti- α Syn-siRNA T3, or with 2 or 4 μ L of all three targets together (mix of Lenti- α Syn-siRNA T1, Lenti- α Syn-siRNA T2 and Lenti- α Syn-siRNA T3).

Quantitative real-time PCR and quantification of α -synuclein transcripts

For quantitative real-time PCR, primer sets for rat α -synuclein, glyceralaldehyde-3-phosphate-dehydrogenase (GAPDH) or β -actin were designed to amplify 100- to 200-bp products, using PRIMER3 software: (http://frodo.wi.mit.edu/cgi-bin/primer3/primer3_http://www.cgi). The following specific primer pairs were used: α -synuclein: 5'-GCG TCC TCT ATG TAG CTT CCA-3' and 3'-AGC AGC AAT GTT CCC AGC; GAPDH: 5'-ATG ACT CTA CCC ACG GCA AG-3' and 5'-CAT ACT CAG CAC CAG CAT CAC-3'; β -actin: 5'-AGC CAT GTA CGT AGC CAT CC-3' and 5'-CTC TCA GCT GTG GTG GTG AA-3'. GAPDH and β -actin were used as endogenous controls for normalization. No difference in expression was observed between these two genes in all quantifications.

Total RNA was extracted from the HEK293T cells (for *in vitro* quantification) and from brains of cocaine-treated animals (for *in vivo* quantification) using TRIzol Reagent (Invitrogen), including an RNase-free DNase step. RNA was quantified by spectrophotometry, and its integrity verified by agarose gel electrophoresis as visualized with ethidium bromide staining. First-strand cDNA was generated from 2 μ g total RNA and Oligo(dT₁₂₋₁₈)-primer with the M-MLV reverse transcription kit (Invitrogen) in a total volume of 20 μ L. The reaction product was used for quantitative real-time PCR using the real-time PCR iCycler (Bio-Rad, Switzerland). cDNA preparation (2 μ L), 0.5 μ M of forward and reverse primers, and 10 μ L of IQ SYBR Green Supermix (Biorad) in a total volume of 20 μ L were

applied, and PCR was performed as follows: 3 min at 95 °C (initial denaturation); 20 °C/s temperature transition rate up to 95 °C for 45 s, 62 °C for 45 s, repeated 40 times (amplification). The PCR reaction was evaluated by melting curve analysis and by checking the PCR products on 2% agarose gel.

The PCR cycle number at which each assay target reached the threshold detection line was determined ('threshold cycles', C_t value). The C_t of each gene was normalized against that of GAPDH or β -actin. To determine the linearity and detection limit of the assay, cDNA samples were amplified for successive 10-fold dilutions in a series of real-time PCRs, using a duplicate assay on each dilution, so that the correlation coefficient could be calculated from the standard curve of C_t values. Comparisons were made between cocaine and saline groups, and significance was calculated using two-tailed Student's *t*-test, and the level of statistical significance was set at $P < 0.05$. Data were expressed as means \pm SEM. The ΔC_t for each candidate was calculated as: $\Delta C_t = [C_t (\text{candidate}) - C_t (\text{GAPDH or } \beta\text{-actin})]$.

The relative abundance of each target in each protocol can be calculated as the ratio between treated and untreated samples (Bahi & Dreyer, 2004; Bahi *et al.*, 2004b).

Western blotting

Rat NAcc (150–200 mg) or transfected HEK293T cells were homogenized in buffer (in mM: Tris, 50, pH 7.5; NaCl, 120; CaCl₂, 1.5; MgCl₂, 5; KCl, 5; EDTA, 5) with a protease inhibitor mixture (Sigma, St Louis, MO, USA; 1 mL/20 g of tissue). Homogenates were solubilized with 1% digitonin, followed by the addition of secondary solubilization buffer [in mM: Tris, 50, pH 7.6; NaCl, 150; 1% Nonidet-P40; 0.5% sodium deoxycholate; EDTA, 2; sodium orthovanadate, 1; phenylmethylsulphonyl fluoride (PMSF), 1; 1% Triton X-100], and centrifuged at 10 000 *g* at 4 °C for 15 min. Solubilized extracts were subjected to sodium dodecyl sulphate–polyacrylamide gel electrophoresis (SDS–PAGE). Blots were blocked with 5% non-fat dried milk dissolved in Tris-Buffered Saline Tween-20 (TBST) (in mM: Tris, 10; NaCl, 150; 0.1% Tween-20) for 1 h at room temperature, and incubated with primary antibodies for either 6-His (1 : 2000, ABCam, ab5000) or α -synuclein (1 : 1000, ABCam, ab1904) for 10–12 h at 4 °C. Blots were washed three times with TBST and incubated for 1 h with the peroxidase-conjugated secondary antibody solution. Proteins were then visualized using enhanced chemiluminescence (Millipore). Membranes were washed for 30 min in TBST and placed in stripping solution (glycine-HCl 25 mM, pH 2.0, 1% SDS) for 30 min and used as described above for β -actin detection. (1 : 4000, Sigma). Signals were detected with a Fluorchem tm 8900 apparatus (Alpha Innotech), and the TIFF pictures were quantified with the multianalyst software (Biorad).

Immunohistochemistry

For immunohistochemistry assays, rats were decapitated. Brains were quickly removed from the skull, immediately frozen in isopentane (at –30 °C for 3 min) and kept at –25 °C. Coronal sections were cut at 14 μ m in a cryostat and placed on gelatinized glass slides, air-dried at room temperature for 20 min and kept at –25 °C until further processing.

Brain sections were fixed for 10 min in cold acetone and washed three times in 1 \times phosphate-buffered saline (PBS). Non-binding sites were blocked by incubating slices for 1 h in 1 \times PBS containing 1% bovine serum albumin, 1% Triton X-100 and 3% normal goat serum. Sections were then incubated overnight at 4 °C with primary antibodies specifically against either α -synuclein (mouse anti- α -synuclein antibody, ABCam ab1904, 1 : 5000), 6-His

(mouse anti-histidine antibody, MCA1396, Serotec, 1 : 5000), NeuN (mouse anti-neuronal-nuclei, MAB 377, Chemicon, 1 : 5000) or synaptophysin (mouse anti-synaptophysin, Chemicon, MAB329, 1 : 1000). Antibodies were diluted in 1 \times PBS containing 0.1% Triton X-100 and 1% normal goat serum. Thereafter, slices were washed three times in 1 \times PBS, and incubated for 2 h in the secondary antibody (Texas red-conjugated goat anti-mouse immunoglobulin G, 1 : 500) or (FITC-conjugated goat anti-mouse IgG, 1 : 500) in 1 \times PBS containing 0.1% Triton X-100. The sections were coverslipped with a medium containing glycerol in PBS (AF1 mounting solution, Citifluor).

Negative controls included omission or substitution of primary antibodies.

Fluorescence microscopy. To observe the stained sections, a multi-fluorescence microscope (Axioptan 2 imaging; Zeiss) was used with a 40 \times or 100 \times objective. Fluorophores (FITC and Texas red) used were detected with the appropriate detecting systems (HAL 100). FITC was excited at 495 nm and was detected through a light path ranging 510–550 nm. Texas red was excited at 570 nm and was detected through a light path ranging 600–660 nm. Stained sections were visualized, photographed using a multichannel camera (Axio-cam, Zeiss) combined with acquisition software (Axiovision system 3.1) and recorded on CD.

In vivo methods

Animal handling and behavioural analysis

Animals used in this experiment were male Wistar rats weighing 250–280 g. All animal experiments were carried out in accordance with the guidelines and regulations for Animal Experimentation, BAG, Bern, Switzerland. The animals were housed four per cage, in clear plastic cages with wire grid lids. Access to food and water was unrestricted. The animals were kept in the animal facility maintained on a 12 h light : dark cycle (lights off at 07.00 h). At the end of each *in vivo* experiment, the animals were decapitated, and brains removed for analysis.

For doxycycline treatment, animals were maintained with 0.02% doxycycline (Sigma) and 5% sucrose supplemented in water. During the gene overexpression period, animals were maintained only with 5% sucrose.

Stereotaxic surgery

Stereotaxic surgery and injection of the lentiviral vector were performed according to previous publications (Bahi *et al.*, 2004a,b). Rats were anesthetized with ketamine-xylazine (10 mg/kg and 0.1 mg/kg) administered i.p. Animals were bilaterally injected into the NAcc with 2 μ L of concentrated lentiviral stock (0.2 mg/mL of p24, corresponding to 8×10^9 IU/mL). The injections were performed bilaterally at the following coordinates, as calculated from bregma and the dura mera: anterior +1.4; lateral \pm 1.2; ventral –6.8 (Paxinos & Watson, 1998).

Intravenous surgery

Wistar rats (250–280 g) were individually housed under a constant 12 h light : dark cycle (lights off at 07.00 h) with free access to food and water. Under anaesthesia, a SILASTIC catheter (15 μ L dead volume) was inserted in the external jugular vein. Rats were anaesthetized with a cocktail mixture of ketamine (10 mg/kg) and xylazine HCl (0.1 mg/kg) administered i.p. in a volume of 1 mL/kg. The surgical procedure consisted of making an incision between the breastbone and the clavicle to expose the jugular vein.

The catheter was inserted into the jugular vein and secured with sutures. The catheter was then passed under the skin and fixed in the mid-scapular region. For the first 3 days after surgery, the catheters were flushed daily with 0.1 mL of saline mixed with heparin (100 U/mL) and gentamicin (1 mg/kg). Thereafter, the catheters were flushed with a saline–heparin mixture after each session. Standard operant chambers (Coulbourn Instruments, Allentown, PA, USA) housed in ventilated sound-attenuating cubicles (Coulbourn Instruments) with fans to mask outside noise were utilized for the SA. One response lever with three ‘cue’ lights above was located on one wall of each chamber. Close to the lever was a recessed food receptacle into which food pellets (45 mg each; Bio-Serv, Frenchtown, NJ, USA) could be dispensed from a pellet dispenser. A house light was also located within the chamber. These chambers were equipped with syringe pump systems, which consisted of an infusion pump (Razel model A-99; Stamford, CT, USA) with a 10-mL plastic syringe connected by a single-channel 22-gauge swivel (Instech Laboratories; Plymouth Meeting, PA, USA) with Teflon tubing. The tubing could be connected to the animal’s catheter system in order to deliver cocaine solution. Graphic State Notation (Coulbourn Instruments) was used to program experimental parameters (e.g. schedules of reinforcements, time periods, etc.), and tabulate and store session data.

Procedures

Experiment 1: cocaine-induced locomotion. Four groups of animals ($n = 6$) were prepared: one group was stereotaxically injected with 2 μ L of Lenti-GFP (controls); one group was stereotaxically injected with 2 μ L of Lenti- α Syn; a third group was injected with 2 μ L of a mix of all three Lenti- α Syn-siRNA; and the fourth group was injected with 2 μ L Lenti- α Syn together with 2 mL of a mix of the three Lenti- α Syn-siRNA. Seven–12 days after stereotaxic surgery, locomotor activity was monitored in daily sessions over 3×5 days in 43.2×43.2 cm MED-OFA-RS cages (MED Associates, USA) during the dark cycle, according to previously published procedures (Bahi *et al.*, 2004a,b). Animals were daily injected cocaine i.p. and were fed without doxycycline for 5 days (Session A), then fed doxycycline for 7 days (Session B), and finally fed without doxycycline for 7 days (Session C). Locomotor activity was recorded on Days 1–5, 8–12 and 15–19; over the 2-day periods between sessions – necessary for full doxycycline-mediated switch of ectopic α -synuclein expression – no measurements were performed (Table 1). Prior to each locomotor activity measurements, the animal was injected with 0.9% saline, and placed into the activity-monitoring cage for a 15 min baseline. After this 15-min period, the session automatically paused for a few seconds

and, during this interval, each subject received cocaine-HCl (Sigma, 15 mg/kg, i.p.) and was then placed back into the locomotor activity-monitoring cage for a further 60 min. Statistical evaluation of behavioural analysis was performed according to previous publications (see Bahi *et al.*, 2004a,b). At the end of the third session (Session C) half of the animals were killed. The other half were switched back on doxycycline for 48 h then killed. All animals were killed by decapitation. During this switch period, animals were still daily administered cocaine and, as for the other group, were killed 24 h after the last cocaine injection. Brains were used either for immunocytochemistry or quantitative real-time PCR.

Two additional groups ($n = 8$) of naïve animals, one fed doxycycline for 5 days, were also killed and their brains dissected. These groups received no cocaine and no lentivirus treatment, and their brain part served as controls for evaluation of cocaine-induced expression of α -synuclein in other groups in quantitative real-time PCR measurements.

Experiment 2: cocaine-induced intravenous SA. Prior to surgery, rats were food-restricted to 85% of free feeding body weight and trained to lever-press for food reinforcement using a fixed ratio (FR)1 schedule of reinforcement during 60-min sessions. Training continued until the animal earned the constant number of reinforcers for two consecutive days. After this training period and a sufficient time to regain weight lost during food restriction, the stereotaxic and the intravenous catheter surgeries were performed at the same time. Four different groups of animals, stereotaxically injected like in Experiment 1, were prepared ($n = 9$). After 7 days of recovery, rats were tested for cocaine SA. They were placed into the operant chambers, connected to the catheter/cannula system, and allowed to SA cocaine during daily 1-h sessions. Testing started 2 h after lights off. One depression of the active lever (FR1) resulted in an intravenous infusion of 20 μ L of a solution containing 0.7 mg/kg of cocaine. The cocaine infusion was delivered over a 2-s period followed by a 40-s time out. Active lever presses that were emitted during the infusion time or during the time out period were tabulated but did not result in any further cocaine delivery. During the infusion, the house light was turned off, and the cue lights over the active lever were illuminated. During the time out period the house light was turned on and the cue lights were switched off. The criterion for acquisition of cocaine SA was defined by a stable number of self-infusions over at least three consecutive sessions ($\pm 20\%$). When cocaine showed control over behaviour under the FR1 schedule of reinforcement, the FR requirement was raised to FR2, FR3 and then to FR5, the schedule used for the remainder of the experiment.

TABLE 1. Summary of experimental protocol for each group of animals in Experiment 1

	Sessions																					
	A					Switch		B					Switch		C					End or switch		
Days 1–22:	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
Doxycycline (+/–)																						
Group 1	–	–	–	–	–	+	+	+	+	+	+	+	–	–	–	–	–	–	–	–	– End ($n = 4$)	
Group 2	–	–	–	–	–	+	+	+	+	+	+	+	–	–	–	–	–	–	–	–	+	+ End ($n = 4$)
Group 3	–	–	–	–	–	+	+	+	+	+	+	+	–	–	–	–	–	–	–	–	– End ($n = 4$)	
Group 4	–	–	–	–	–	+	+	+	+	+	+	+	–	–	–	–	–	–	–	–	+	+ End ($n = 4$)
Measurement	+	+	+	+	+	–	–	+	+	+	+	+	–	–	+	+	+	+	+	–	–	–

See the Materials and methods section for details. At the end of the experiment the animals were killed and the brains used either for immunocytochemistry or quantitative real-time PCR.

As in Experiment 1, half of the animals were killed by decapitation 24 h after the last cocaine SA session, while the remaining ($n = 5$) were fed doxycycline for 48 h before being killed. During this switch period, animals were still daily tested and, as for the other group, were killed 24 h after the last cocaine SA session. Brains were used either for immunocytochemistry, Western blot or quantitative real-time PCR analysis.

In vivo assays of α -synuclein

The efficiency of the lentivirus-mediated α -synuclein expression *in vivo* was assessed by stereotaxic injection, into the NAcc, of Lenti- α Syn stock alone in a first group of animals, or Lenti- α Syn together with Lenti- α Syn-siRNAs mix in a second group of animals. To suppress endogenous α -synuclein expression, a third group of animals were infected only with 2 μ L of Lenti- α Syn-siRNAs mix. Finally, a control group of animals was injected in the same brain area with Lenti-GFP. After surgery, animals were left for recovery for 7 days. At the end of the behavioural analysis (locomotor activity or SA), some of the animals were killed by decapitation 24 h after the last cocaine injection, while the remaining animals were fed doxycycline for 48 h before being killed. During this switch period, animals were still daily administered cocaine and, as for the other group, were killed 24 h after the last cocaine injection. Brains were either homogenized for protein or quantitative real-time PCR analysis, or dissected out for immunohistochemistry analysis. For molecular analysis, brains were dissected out and the NAcc, striatum and ventral tegmental area (VTA) regions were isolated. Tissues were used for RNA extraction using TRIzol Reagent (Invitrogen) or for protein extraction.

Statistical analyses

Two-way ANOVA was used to compare both lentiviral and drug treatment in the presence or absence of doxycycline, using GraphPad PRISM (V3.0, GraphPad, San Diego, California, USA). For locomotor activity two-way ANOVA was used, with time as the within-subject factors and lentiviral treatment as the between-subject factors followed by Bonferroni *post hoc* tests. For SA and real-time PCR analysis, two-way ANOVAs were used with drug treatment (water vs doxycycline) as the within-subject factors and lentiviral treatment as the between-subject factors followed by Bonferroni *post hoc* tests.

Results

Lentivirus-mediated α -synuclein expression in HEK293T cells

α -Synuclein was cloned into a tetracycline-regulatable lentivirus system. The lentivirus expressing α -synuclein (Lenti- α Syn) was used for *in vitro* assays (see Materials and methods). HEK293T cells were infected with different amounts of virus (Fig. 1A), and α -synuclein mRNA was measured by real-time PCR. As shown, Lenti- α Syn induced α -synuclein gene and protein expression in a dose (titre)-dependent manner. Using 2, 4 and 8 μ L of viral stock, α -synuclein mRNA increased by 4.7-, 8.8- and 12.6-fold ($F_{6,28} = 32.81$, $P < 0.001$), respectively. This induction is completely reversible in the presence of doxycycline, which blocks lentivirus-mediated α -synuclein overexpression, both at the mRNA (Fig. 1A) and the protein levels (Fig. 4). For real-time PCR, α -synuclein transcripts levels were normalized against β -actin used as endogenous control. GAPDH expression, an endogenous control, is unaffected by lentivirus-mediated gene transfer (Fig. 1A and B)

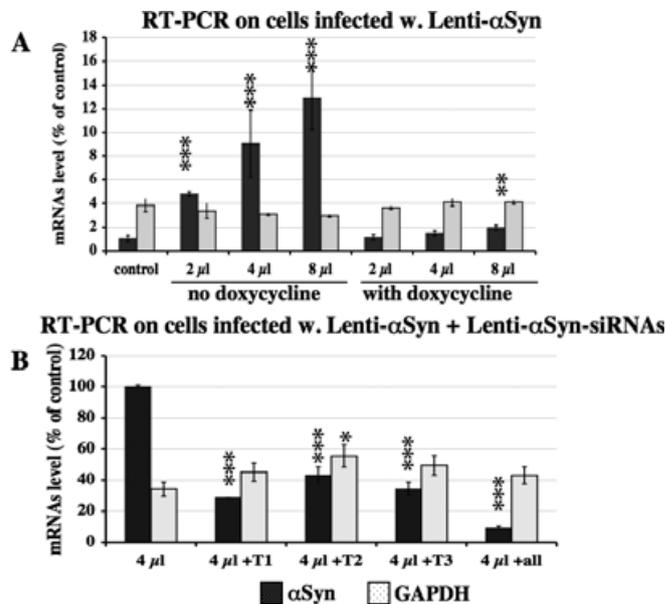


FIG. 1. Quantification of *in vitro* expression of α -synuclein mRNA in HEK293T cells using quantitative real-time polymerase chain reaction (PCR). (A) Dose-response effects of Lenti- α Syn. HEK293T cells were infected with 0, 2, 4 or 8 μ L Lenti- α Syn in culture medium in the presence or absence of doxycycline (30 ng/mL), as indicated. After 96 h, culture medium was removed and cells were used for total RNA extraction, cDNA preparation and quantitative real-time PCR using specific oligos. Results were normalized against GAPDH used as endogenous control. Normalized transcript levels were from cells infected without doxycycline. Control: non-infected HEK293 cells. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.005$ vs non-infected cells. (B) Silencing efficiency of Lenti- α Syn-siRNAs. HEK293T cells were infected with 4 μ L Lenti- α Syn with or without other viruses, as indicated. After 96 h, culture medium was removed and cells were used for total RNA extraction, cDNA preparation and quantitative real-time PCR using specific sets of primers. Results were normalized vs GAPDH as endogenous control. ** $P < 0.05$ and *** $P < 0.005$ vs cells infected with Lenti- α Syn.

siRNAs-expressing lentiviruses and α -synuclein silencing *in vitro*

In order to knockdown α -synuclein expression, lentivirus-based RNA interference was used. Three different siRNAs were designed, cloned into the lentiviral transfer plasmid (Lenti- α Syn-siRNAs T1, T2 and T3) and their silencing efficiency was assessed *in vitro* in Lenti- α Syn-infected HEK293T cells (Fig. 1B). Target specificity was controlled by co-infections of HEK293T cells with Lenti-GFP. Ninety-six hours after infection, cells were harvested, total RNA was extracted and transcripts levels were measured by means of quantitative real-time PCR using specific primers for each candidate. All normalizations were performed against β -actin (no significant difference was observed when GAPDH instead of β -actin was used for normalization, $P > 0.05$). Cells co-infected with Lenti- α Syn were set-up to 100% of α -synuclein expression level (Fig. 1B; '4 μ L'). When Lenti-GFP was also added during co-infection no significant change of α -synuclein mRNA levels was observed, yielding $\sim 98\%$ of α -synuclein transcripts, vs 100% in controls (data not shown). In contrast, co-infection with Lenti- α Syn-siRNAs T1, T2 and T3 resulted in drastic α -synuclein downregulation, yielding 29.1%, 43.1% and 34.2% residual α -synuclein expression levels, respectively. (Fig. 1B; $F_{4,20} = 22.70$, $P < 0.01$). Co-infection in the presence of all three Lenti- α Syn-siRNAs together resulted in 90.8% α -synuclein mRNA knockdown (Fig. 1B; '4 μ L + all', $P < 0.01$). Under all conditions, the expression levels of other genes (e.g. GAPDH; Fig. 1B) were not

affected by Lenti- α Syn-siRNAs, indicating that the targets are highly α -synuclein specific.

Western blot analysis, using either anti- α -synuclein antibody or anti-6His antibody, showed that protein expression highly correlates with transcripts levels (Fig. 2). Quantification from 6-His-based Western blots shows that co-infection of Lenti- α Syn with T1, T2 or T3 resulted in 12%, 22% and 3% residual α -synuclein, respectively, whereas co-infection of Lenti- α Syn with all three Lenti- α Syn-siRNAs together resulted in <4% residual α -synuclein (Fig. 2; $F_{6,28} = 23.07$, $P < 0.001$).

In vivo cocaine-mediated α -synuclein expression and silencing

Four groups of animals ($n = 8$), stereotaxically injected with different combinations of lentiviruses, were prepared and behaviour was measured in three 5-day sessions (locomotor activity or SA, see Materials and methods, and Table 1). Animals received daily cocaine injections. During the first 5-day session ('Session A') animals were fed without doxycycline during five consecutive days, and then switched to the doxycycline regimen for 7 days, but the behavioural analysis was performed only 2 days after the switch, for five consecutive days ('Session B'). Finally, animals were switched again without doxycycline and measured 2 days later for five consecutive days ('Session C'). At the end of the behavioural analyses, half of the animals from each group were killed, the second half of the animals was fed doxycycline for 48 h (turning off α -synuclein overexpression

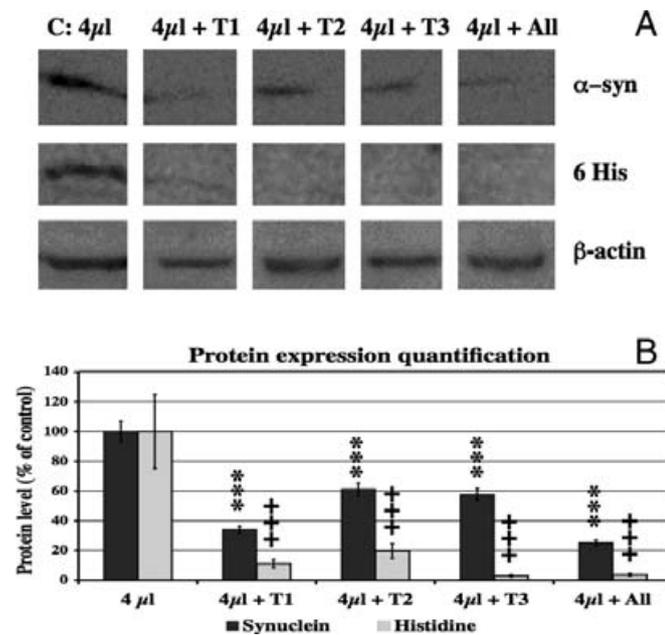


FIG. 2. Expression of α -synuclein in the HEK293T cells. Cells were transfected as in Fig. 1. Proteins were extracted as described in Materials and methods, to compare α -synuclein expression in different lentivirus infected cells (control C in left-most column). (A) Immunoblots of α -synuclein expression. Western blots from gels loaded with 15 μ g protein/lane were analysed for α -synuclein or 6-histidine expression, and re-probed with anti- β -actin antibody to compare α -synuclein expression in lentivirus-infected cells. (B) Semi-quantitative analysis of Western blot signals. α -Synuclein signals were estimated using multianalyst software and normalized against GAPDH signal. *** $P < 0.001$ α -synuclein protein quantification vs cells infected with 4 μ L Lenti- α Syn; +++ $P < 0.001$ 6-histidine quantification vs cells infected with 4 μ L Lenti- α Syn, two-way ANOVA, Bonferroni *post hoc* tests.

in the NAcc), and then killed. After killing, brains were removed, NAcc tissues were dissected out and used either for total RNA extraction or Western blot analysis to test lentivirus-mediated and cocaine-induced α -synuclein expression *in vivo*.

mRNA expression in vivo

After reverse transcription, mRNA expression levels were measured by means of quantitative real-time PCR. Data have been normalized with respect to the control groups that consisted of naïve animals not receiving cocaine but fed either with or without doxycycline. These two groups served to assess cocaine-mediated induction of α -synuclein. Data are shown in Fig. 3.

When animals have been injected with Lenti-GFP, α -synuclein expression was ~ 3.1 compared with saline-injected control (Fig. 3; $F_{4,20} = 103.03$, $P < 0.01$), reflecting cocaine-mediated induction of endogenous α -synuclein. Animals of this group switched to doxycycline displayed ~ 2.5 -fold cocaine-induced increase in α -synuclein expression, compared with saline-injected control animals (Fig. 3; $P > 0.05$, $F_{4,20} = 103.03$), a ratio not significantly different from the corresponding group fed without doxycycline. The ratios of other targets, e.g. β -actin mRNA, remained unchanged over saline-treated control animals in both cases.

When animals were injected with only Lenti- α Syn, strong induction of α -synuclein mRNA expression was observed in the absence of doxycycline, with a ratio of ~ 12.3 ($F_{4,20} = 103.03$, $P < 0.001$) compared with saline-treated controls (Fig. 3). This expression level dropped 3.3-fold with doxycycline, up to a ratio of 3.7 (compared with naïve controls), equivalent to the ratio observed for GFP-treated animals ($F_{4,20} = 103.03$, $P < 0.01$). β -Actin mRNA expression was not affected, and ratios of ~ 1.5 and ~ 1.3 , respectively (no doxycycline), and ~ 1.6 and ~ 2.8 , respectively (with doxycycline), were found for cocaine- and saline-treated animals under these conditions

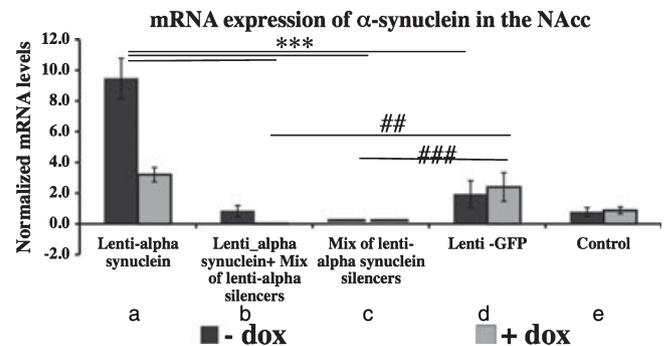


FIG. 3. *In vivo* overexpression and silencing of α -synuclein mRNA in the nucleus accumbens (NAcc) after infection with Lenti- α Syn and/or Lenti- α Syn-siRNAs: quantification of transcripts by means of quantitative real-time PCR. Animals ($n = 8$) were bilaterally injected into the NAcc with either: (a) Lenti- α Syn alone; (b) Lenti- α Syn combined with a mix of all Lenti- α Syn-siRNAs; (c) a mix of Lenti- α Syn-siRNAs; or (d) Lenti-green fluorescent protein (GFP); and (e) untreated animals (as a control). At the end of the behavioural analysis (locomotor activity), part of the groups used were killed by decapitation 24 h after the last injection; the NAccs were dissected out and used for total RNA isolation, cDNA preparation and quantitative real-time PCR using specific oligos. Results were normalized against GAPDH. ***Represents points significantly different from Lenti- α Syn-infected rats (** $P < 0.001$). ## and ### Represent points significantly different from Lenti-GFP-injected rats. (## $P < 0.01$; ### $P < 0.001$) Two-way ANOVA, Bonferroni *post hoc* tests.

($F_{4,20} = 103.03$, $P < 0.01$), similar to those observed in the Lenti-GFP-treated group.

When animals have been injected with Lenti- α Syn together with a mix of Lenti- α Syn-siRNAs, drastic inhibition of α -synuclein expression is found in the absence of doxycycline, resulting in a ratio of ~ 1.1 ($F_{4,20} = 103.03$, $P < 0.01$) compared with naïve controls, i.e. at least 11-fold inhibition (Fig. 3). With doxycycline, complete inhibition of α -synuclein was found, with ratios ~ 0.01 compared with naïve controls ($F_{4,20} = 103.03$, $P < 0.001$). Indeed, doxycycline blocks exogenous, lentivirus-mediated α -synuclein overexpression (but not endogenously expressed α -synuclein), while silencers also block endogenous α -synuclein. Under these conditions, the other targets (β -actin, etc.) were not modified at all, indicating that Lenti- α Syn-siRNAs had no effects on their expression levels.

Finally, when animals have been injected with the Lenti- α Syn-siRNAs only (without Lenti- α Syn), the α -synuclein mRNA expression level dropped up to ~ 0.007 ($P < 0.001$) and even to ~ 0.003 with doxycycline ($F_{4,20} = 103.03$, $P < 0.001$) – corresponding to full silencing of endogenous α -synuclein mRNA expression – whereas β -actin was not modified. This value, compared with that observed in the Lenti-GFP-treated group, indicates that a $> 99\%$ knockdown of the endogenous α -synuclein has been achieved. As described previously (Bahi *et al.*, 2004a,b, 2005a,b), the use of a mix of three lentivirus silencers most often warrants optimal gene silencing, as requested for observation of solid behavioural changes.

Protein expression in vivo

Protein expression was assessed from the NAcc of treated animals after behavioural analysis (see Table 1). After killings, brains were dissected out, and the NAccs were used for Western blot analysis (Fig. 4).

Cocaine induces an approximate 1.8-fold increase in α -synuclein expression in naïve (control) animals not injected with any lentivirus (Fig. 4), in agreement with previous publications (Brenz-Verca *et al.*, 2003). In the absence of doxycycline, Lenti- α Syn-treated animals display a 7.3-fold higher expression of ectopic α -synuclein after chronic cocaine treatment (Fig. 4; 100%, $P < 0.001$, $F_{3,16} = 239.68$), but this expression level is reduced by 70% in the presence of doxycycline. Animals treated with both Lenti- α Syn and Lenti- α Syn-siRNAs display 22% and 6% expression in the presence or absence of doxycycline ($P < 0.001$, $F_{3,16} = 239.68$), compared with the first group (animals treated with Lenti- α Syn only, in the absence of doxycycline). The remaining α -synuclein expression reflects cocaine-mediated endogenous expression of α -synuclein, which is almost fully knocked down by Lenti- α Syn-siRNAs, yielding 4–5% expression only compared with the Lenti- α Syn group and 18% of the control group only (Fig. 4; $F_{3,16} = 239.68$, $P < 0.001$).

Immunohistochemistry was performed on brains from animals treated under similar conditions (Fig. 5). Animals treated with Lenti- α Syn display a very strong overexpression of α -synuclein in the NAcc (Fig. 5; left panels); under these conditions doxycycline induced a very large ($> 90\%$) suppression of α -synuclein overexpression in the NAcc. Animals treated with Lenti- α Syn combined with Lenti- α Syn-siRNA displayed a strong ($> 98\%$) suppression of α -synuclein, although some α -synuclein could still be observed (Fig. 5; left-middle panels) that was almost fully abolished when under doxycycline. In contrast, animals treated only with Lenti- α Syn-siRNA, silencing endogenously expressed α -synuclein, display $> 99\%$ silencing of endogenous cocaine-mediated α -synuclein expression. Cocaine-mediated α -synuclein expression could be assessed from the control GFP-

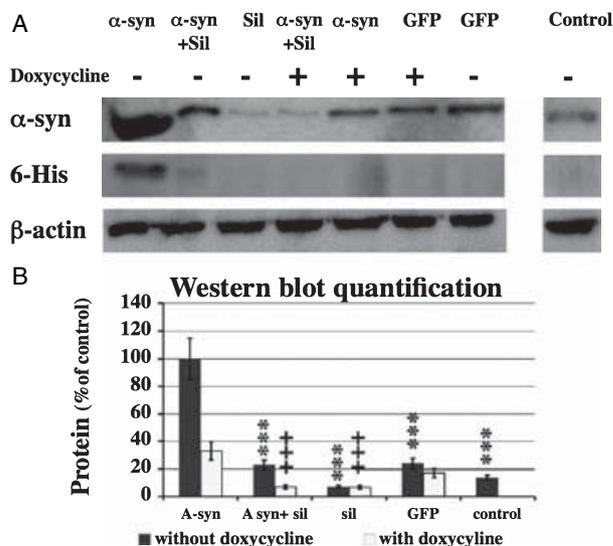


FIG. 4. Expression of α -synuclein in the NAcc from lentivirus-injected rats. Animals were prepared as in Fig. 3. Proteins were extracted from the NAcc at the end of each 5-day sessions of locomotor activity, as described in Materials and methods. (A) Immunoblot of α -synuclein. Western blots, from gels loaded with 15 μ g protein/lane, were analysed for α -synuclein or 6-histidine expression, and reprobated with anti- β -actin antibody to compare α -synuclein expression in different lentivirus-injected rats. (B) Semi-quantitative analysis of Western blot signals. α -Synuclein signals were estimated using multianalyst software and normalized against the GAPDH signal. *** $P < 0.001$ α -synuclein protein quantification vs data from α -synuclein-injected rats without doxycycline, +++ $P < 0.001$ α -synuclein protein quantification vs data from α -synuclein-injected rats with doxycycline Two-way ANOVA, Bonferroni *post hoc* tests.

treated group and was not affected by doxycycline treatment (Fig. 5; right panels).

Furthermore, the subcellular localization of α -synuclein has been determined via double-immunolabelling. NeuN was used as a specific marker for neuronal nuclei, and synaptophysin was used to detect presynaptic vesicles (Supplementary material, Fig. S1). As shown in Fig. 5, α -synuclein was consistently co-expressed with synaptophysin, indicating that the protein is found in presynaptic vesicles. Furthermore, double-immunolabelling of α -synuclein and NeuN does not show α -synuclein co-localization in the nuclear part of the neurons; under these conditions α -synuclein appears rather expressed in a neuritic manner. Taken together these results demonstrate that lentiviral overexpression of α -synuclein results in high protein levels that could be abolished by doxycycline treatment. Furthermore, lentiviral-mediated overexpression of α -synuclein leads to increased protein levels both in neurites and presynaptic vesicles of neuronal cells (supplementary Fig. S2).

Locomotor activity changes induced upon α -synuclein expression

Previous work clearly established that GFP overexpression in the NAcc has no significant effect on locomotor activity, compared with naïve animals (Bahi *et al.*, 2004a,b). For this reason, Lenti-GFP-treated animals have been used as a control in the behavioural test (Fig. 6). Animals infected with Lenti-GFP displayed a significant induction of locomotor activity after cocaine administration, with a peak at ~ 4100 mm/min at 30 min (i.e. 10 min after drug administration; Fig. 6A; upper row, right panel, 'Session A (no

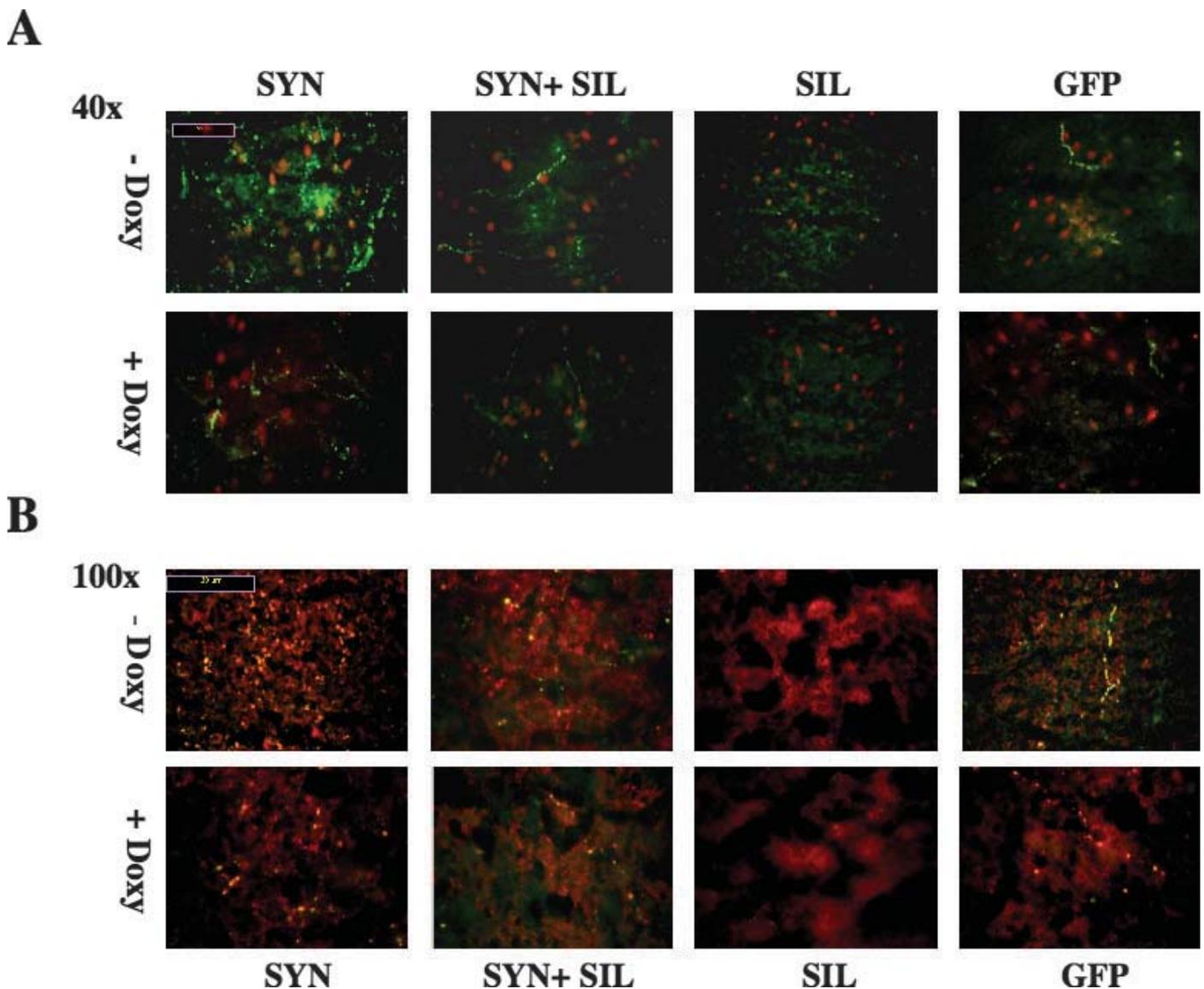


FIG. 5. Immunohistochemistry of α -synuclein expression in the NAcc. Immunohistochemistry was performed after *in vivo* delivery of either: (a) Lenti- α Syn alone (2 μ L); (b) Lenti- α Syn (2 μ L) combined with Lenti- α Syn-siRNAs (2 μ L); (c) Lenti- α Syn-siRNAs (2 μ L); or (d) Lenti-green fluorescent protein (GFP; 2 μ L). Lentiviruses were injected into the NAcc according to the coordinates described in the Materials and methods. Animals used for behavioural analysis were fed either 5% sucrose or 5% sucrose/0.02% doxycycline (see text for details). Animals were killed at the end of behavioural analyses; brains were dissected out and processed for immunohistochemistry (see Materials and methods). Upper panels: expression of α -synuclein from animals fed 5% sucrose; lower panels: expression of α -synuclein from animals under doxycycline treatment. (A) Sections were double-labelled using antibodies against α -synuclein (green) and NeuN (red), and visualized by fluorescence microscopy at 40 \times magnitude. Lentiviral overexpression of α -synuclein shows an increase of α -synuclein (green) that is not co-localized with the nuclei neurons labelling (red), but seems more likely expressed in neurites. (B) Sections were double-labelled using antibodies against α -synuclein (green) and synaptophysin (red), and visualized by fluorescence microscopy at 100 \times magnitude. Yellow colour shows a large proportion of synaptic terminals co-expressing α -synuclein and synaptophysin.

dox')). During Session B (i.e. animals fed doxycycline), the distance travelled after cocaine injection was not significantly altered, with a peak at \sim 5000 mm/min at 30 min (Fig. 6A; middle row, right panel, 'Session B (dox)'). Upon doxycycline removal (Session C), enabling for GFP re-expression, the locomotor activity was similar to the other sessions, with a peak at \sim 4200 mm/min at 30 min (Fig. 6A; lower row, right panel, 'Session C (no dox)').

Animals infected with Lenti- α Syn displayed a significant increase (68%) in locomotor activity after cocaine administration, compared with Lenti-GFP-treated animals, with a peak at \sim 6800 mm/min observed 10 min after drug administration (Fig. 6A; upper row, left panel, 'Session A (no dox)', $F_{3,224} = 236.1$, $P < 0.001$). After 5 days

the same animals were fed doxycycline in the drinking water, inducing a downregulation of exogenous lentivirus-expressed α -synuclein in the NAcc. Two days after the doxycycline switch, behaviour upon chronic cocaine administration was further monitored for five consecutive days (Session B). The distance travelled after cocaine injection displayed a peak at \sim 5100 mm/min at 30 min (Fig. 6A; middle row, left panel, 'Session B (dox)'), i.e. almost identical to levels observed with Lenti-GFP-treated animals ($P > 0.05$, $F_{3,224} = 688.36$). After 5 days monitoring under this regimen, doxycycline was removed (Session C), enabling re-expression of lentivirus-mediated α -synuclein in the NAcc). Behaviour, measured 2 days later for five consecutive days under these conditions, expressed a peak of locomotor activity at

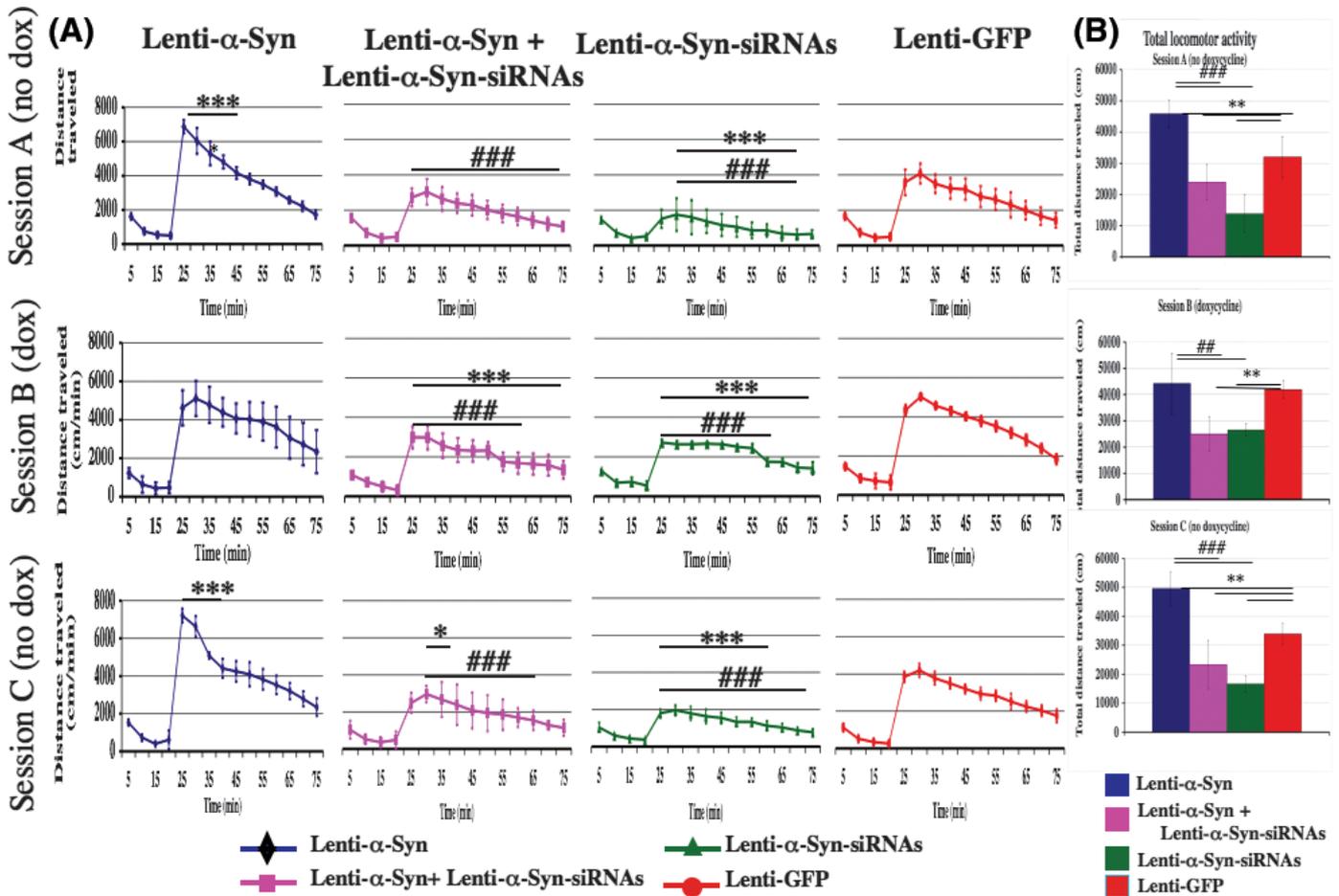


Fig. 6. (A and B) *In vivo* effects of α -synuclein overexpression or silencing in the NAcc on cocaine-induced locomotor activity. Values represent means \pm SEM of 5 days ($n = 8$). One week after surgery, chronic cocaine delivery was started and locomotor activity was monitored. At each daily session, animals received saline injections before the habituation period (15 min) followed by cocaine delivery (i.p. 15 mg/kg), and the locomotor activity was monitored immediately after drug injection. Overexpression of α -synuclein in the NAcc induced a significant increase on cocaine-induced locomotor activity that could be abolished by doxycycline treatment. Silencing of α -synuclein lead to a decrease of cocaine-induced locomotor activity. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$, vs data from Lenti-green fluorescent protein (GFP)-infected rats; ## $P < 0.01$ and ### $P < 0.001$ vs data from Lenti- α syn-injected rats, two-way ANOVA, Bonferroni *post hoc* tests.

~ 7300 mm/min at 30 min (Fig. 6; lower row, left panel, 'Session C (no dox)', $F_{3,224} = 544.33$, $P < 0.001$). The difference between peaks in Sessions A and C was not significant ($P > 0.2$).

Another group, co-injected Lenti- α Syn and Lenti- α Syn-siRNAs, was submitted to the same series of regimen and cocaine treatment. In Session A, locomotor activity was significantly lower than in the previous group under the same conditions, with a peak at ~ 3100 mm/min at 30 min (Fig. 6A; upper row, central panel, 'Session A (no dox)', $P < 0.001$ as compared with α -synuclein-treated group, $F_{3,224} = 236.12$). In Session B, locomotor activity slightly decreased, with a peak of ~ 2900 mm/min at 30 min (Fig. 6A; middle row, central panel, 'Session B (dox)'); $P < 0.001$ as compared with α -synuclein-treated group; $F_{3,224} = 688.36$). Under these conditions, downregulation of exogenous, lentivirus-mediated α -synuclein expression is achieved, together with knockdown of endogenous (cocaine-induced) expression. The observed residual behavioural activity is about 30% lower than in corresponding GFP-treated animals, as in the absence of α -synuclein-specific siRNAs, synuclein would be strongly induced by cocaine under these conditions. In Session C, when doxycycline has been removed from water on the same animals, enabling full expression of the ectopic α -synuclein in the NAcc, the locomotor activity drops to the basal

level, reaching levels comparable to the first session (~ 3000 mm/min 15 min after cocaine delivery; Fig. 6A; lower row, central panel, 'Session C (no dox)', $P < 0.001$ as compared with α -synuclein-treated group, $F_{3,224} = 544.33$).

The last group of animals, infected only with Lenti- α Syn-siRNAs to assess effects of endogenous α -synuclein, displayed very low locomotor activity ~ 1700 mm/min at 30 min in the absence of doxycycline (Fig. 6A; upper row, right panel, 'Session A (no dox)', $P < 0.001$, $F_{3,224} = 236.12$ as compared with α -synuclein-treated group). In Session B, i.e. when switched to the doxycycline regimen, no significant change was observed, with a peak of ~ 2600 mm/min at 30 min (Fig. 6A; middle row, right panel 'Session B (dox)', $P < 0.001$ as compared with α -synuclein-treated group, $F_{3,224} = 688.36$). Upon doxycycline removal, 5 days later, locomotor activity peaked at ~ 2000 mm/min (Fig. 6A; lower row, right panel, 'Session C (no dox)', $P < 0.001$ as compared with α -synuclein-treated group, $F_{3,224} = 544.33$). The observed difference between these three sessions within this group of animals was not significant ($P > 0.2$). These observed behavioural effects were highly correlated with changes of α -synuclein, as verified at protein levels by means of Western blot analysis (Fig. 4) and immunohistochemistry (Fig. 5).

Drug-induced behavioural sensitization was observed during Session A for all groups, over the five initial days (Fig. 7). Sensitization is highest in the Lenti-GFP-treated animals, which display a 63% increase in locomotor activity over this session, vs about 15% increase in α -synuclein-overexpressing animals ($P < 0.001$, $F_{2,48} = 10.52$). Silencing α -synuclein raises this effect, to values close to GFP-treated animals. During subsequent Sessions B and C, further sensitization could not be observed significantly ($P > 0.05$, $F_{6,48} = 5.24$).

Changes in SA upon α -synuclein expression

Three groups of animals ($n = 8$) were used to test the effects of α -synuclein expression in the NAcc on the cocaine SA paradigm (Fig. 7). One group was injected with the doxycycline regulatable Lenti- α Syn, a second group was co-injected with Lenti- α Syn and Lenti- α Syn-siRNAs, and a third group was injected with the doxycycline regulatable Lenti-GFP and served as a control group. Animals were trained for SA, then operated, as described in the Materials and methods. Data are presented in Fig. 8.

GFP-injected animals demonstrated a strong SA behaviour under a FR5 schedule of reinforcement with a mean of 15.5 infusions per session. After 5 days, the same animals were fed with doxycycline in the drinking water. During two consecutive days, animals could SA cocaine, but behaviour was not measured in order to achieve a full doxycycline switch of α -synuclein expression before measurements. Thereafter, SA was further monitored for five consecutive days under doxycycline (Session B). Under these conditions the number of infusions was unchanged, with a mean at 15.2 per session.

The second group, injected with Lenti- α Syn and overexpressing of α -synuclein in the NAcc, displayed a strong increase in SA of cocaine during Session A, with a mean of 30 infusions per session, i.e. a 1.9-fold increase over the control GFP group ($F_{2,180} = 745.64$, $P < 0.001$). After 5 days, animals were fed doxycycline (Session B), suppressing α -synuclein overexpression, and SA was measured 2 days later for five consecutive days. Under these conditions, the number of cocaine self-infusions dropped to 18.2 per session ($P > 0.05$).

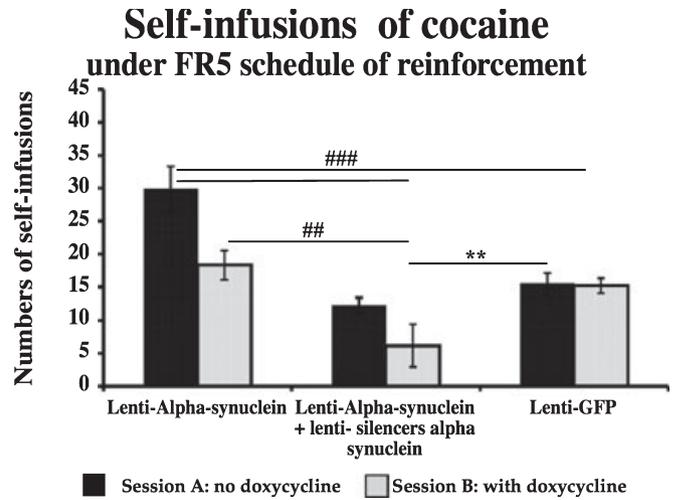


FIG. 8. Effects of α -synuclein on cocaine SA under fixed ratio (FR)5 schedule of reinforcement. Each histogram represents the means of dose-response for five consecutive days ($n = 8$). SA was performed for five consecutive days in the presence of doxycycline in the drinking water (Session A); then doxycycline was removed (Session B). Overexpression of α -synuclein in the NAcc results in a significant increase in the number of cocaine SA. This change could be abolished by doxycycline treatment. $**P < 0.01$ vs data from Lenti-GFP-infected rats; $##P < 0.01$ and $###P < 0.001$ vs data from α -synuclein-injected rats, two-way ANOVA, Bonferroni *post hoc* tests. Results show an increase in numbers of cocaine self-infusions in animals injected with Lenti- α Syn as compared with the Lenti-green fluorescent protein (GFP)-injected group ($###P < 0.001$) and with the Lenti- α Syn + Lenti- α Syn-siRNAs-injected group. Therefore, after doxycycline treatment, animals injected with Lenti- α Syn still display a significant increase in numbers of cocaine self-infusions as compared with Lenti- α Syn + Lenti- α Syn-siRNAs rats ($##P < 0.01$).

The third group of animals – co-injected with Lenti- α Syn and Lenti- α Syn-siRNAs – displayed a mean of 12.2 infusions per session in Session A, not significantly different from the control group ($P > 0.05$). Doxycycline treatment switches off Lenti- α Syn but not

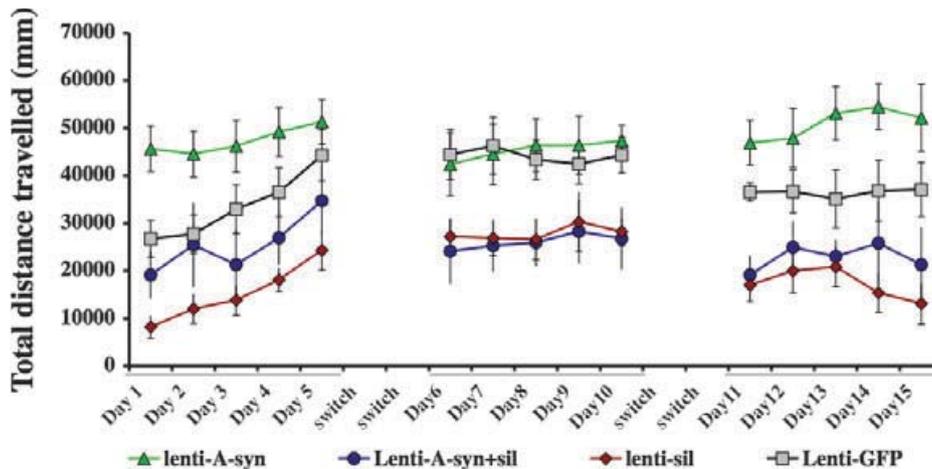


FIG. 7. Drug-induced behavioural sensitization. Animals used for behavioural analysis were fed either 5% sucrose or 5% sucrose/0.02% doxycycline (see text for details). Values represent means \pm SEM of peak locomotor activity from one group of animals ($n = 8$) each day. One week after surgery, chronic cocaine delivery was started and locomotor activity was monitored (see Fig. 6). After five consecutive days of drug delivery, animals were fed doxycycline and measured 2 days after the switch for another period of five consecutive days. Then the regimen was switched back (no doxycycline) and, 2 days after the second switch, was measured for 5 days.

Lenti- α Syn-siRNAs, resulting both in a suppression of lentivirus-mediated α -synuclein overexpression as well as in silencing endogenous α -synuclein expression. Under these conditions (Session B), animals displayed a very high decrease in cocaine SA, with a mean of 6.2 infusions per session. This represents a 40.8% ($F_{2,180} = 745.64$, $P < 0.01$) and 34.1% ($F_{2,180} = 745.64$, $P < 0.01$) decrease in the number of cocaine self-infusion, compared with the GFP control group and the α -synuclein-injected group, respectively, under the same conditions.

Discussion

In the present study we aimed at establishing a causal relationship between alterations in α -synuclein levels and cocaine-induced behaviours. To this end, we have developed inducible lentiviral tools that very efficiently enable to locally alter α -synuclein levels, and studied the consequences of varying α -synuclein levels on cocaine-induced locomotion and SA. In previous studies we showed that cocaine administration induces a strong increase of α - and γ -synucleins in the NAcc (Brenz-Verca *et al.*, 2003). This was corroborated in the present study: upon cocaine administration, α -synuclein mRNA in GFP-treated animals is induced 3.1-fold in the NAcc, compared with naïve, saline control animals. There is correlative evidence in humans that cocaine abusers overexpress α -synuclein in dopaminergic neurons, mainly in the VTA and substantia nigra (Mash *et al.*, 2003).

In this study we present evidence that an increase in α -synuclein expression strongly correlates with behavioural changes observed under different drug administration paradigms. Our data clearly show a differential effect of overexpression and siRNA conditions with locomotion correlating with expression level. Local overexpression of α -synuclein in the NAcc rapidly induces increased locomotor response to cocaine and increased SA of the drug, both effects that can be fully reversed in the presence of doxycycline. Furthermore, selective knockdown of α -synuclein in the NAcc also strongly affects both locomotor activity and SA. Behavioural effects strongly correlate mRNA and protein expression of α -synuclein in this brain area. This suggests that α -synuclein expression in the NAcc plays an important role in cocaine-mediated behaviour. Behavioural changes have been compared with GFP-treated animals, as previous studies established that GFP does not affect behaviour in the paradigms under investigation (Bahi *et al.*, 2004a,b). In addition, saline-treated animals were used as control, showing that cocaine indeed has some effects on α -synuclein mRNA levels in the NAcc. The increased SA of cocaine suggests that enhanced expression of α -synuclein makes the animals less sensitive to cocaine. The reinforcing properties of cocaine appear to have decreased, as suggested by the increased number of self-infusion, thus α -synuclein overexpression may induce reduced cocaine reinforcement. The animals are compensating for the decreased effect by taking more drug. Therefore, increased α -synuclein expression in the NAcc may result in enhanced locomotion induced by cocaine, but decreased reinforcing properties of the drug. On the other hand, our studies have so far been limited to single dose injection, and no dose-effect responses have been performed in the present study. Thus, it is also possible that α -synuclein expression in the NAcc induces a rightward shift in the dose-effect curve. Shifts in dose-injection curves may reflect reward allostasis, not sensitization (Ahmed & Koob, 2004), and escalated drug intake may often be suggestive of a negative reinforcement due to allostatic decrease in reward function (Ahmed & Koob, 1999, 2005). Further studies at different doses may clarify this.

Interestingly, changes in α -synuclein expression result in apparent cocaine-induced sensitization in the first sessions (Session A), but no more in Sessions B and C. However, the absence of sensitization in synuclein knockdown animals could be interpreted as the results of a ceiling effect. The level of cocaine-induced locomotion may then be so high already on the first exposure that there are no further possibilities for a further increase. As a matter of fact, cocaine-induced sensitization cannot be reversed on the very same animal fed doxycycline, then upon doxycycline removal. On the other hand, behavioural sensitization can arise from neuroadaptations in multiple brain nuclei (Vanderschuren & Kalivas, 2000), and our data could rather reflect the differential involvement of the accumbens vs other brain regions in the expression or induction of cocaine sensitization. Indeed, cortical and allocortical brain regions such as the prefrontal cortex appear to be more critical for cocaine sensitization.

It has been established that cocaine abuse elevates α -synuclein and DAT levels in the human striatum (Qin *et al.*, 2005). In addition, α -synuclein interactions with the DAT mediate the reuptake of DA released into the synaptic cleft by DA-producing neurons (Perez & Hastings, 2004; Vekrellis *et al.*, 2004; Wersinger & Sidhu, 2005). This interaction regulates dopaminergic content, neurotransmission and synaptic strength at dopaminergic terminals (Zhou *et al.*, 2002). Studies in α -synuclein knockout mice revealed that it might inhibit refilling and reduce the storage capacity of presynaptic DA-containing vesicles, thus serving as a negative regulator of dopaminergic neurotransmission (Abeliovich *et al.*, 2000; Yavich *et al.*, 2004). α -Synuclein exerts modulatory effects on DAT function, as well as on tyrosine hydroxylase activity, the rate-limiting enzyme in DA biosynthesis (Clayton & George, 1999; Perez *et al.*, 2002), and on vesicular storage of DA (Ueda *et al.*, 1993; Akopian & Wood, 1995). This would imply that a primary normative function of α -synuclein in DA-producing neurons might be the regulation and maintenance of DA homeostasis (Neystat *et al.*, 1999; Lotharius *et al.*, 2002). The high concentration of synucleins in presynaptic terminals suggested an essential role for synucleins in synapse formation, neurotransmitter release or synaptic plasticity (Clayton & George, 1998). As a matter of fact, our data may strongly support this view, as changes in its expression in the NAcc strongly affects reward, a process essentially related to DA release in this brain area. In our study, viral injection was performed into the NAcc. Lentiviruses being replication incompetent and self-inactivating, they will not spread away from the injected area.

Because of its high levels and its role in dopaminergic pathways, it has been suggested that α -synuclein could be potentially involved in the effects exerted by psychostimulants on the dopaminergic reward pathway. α -Synuclein mRNA regulation by chronic morphine administration in the mouse amygdala, a target region of dopaminergic projection, plays an essential role in reinstatement of opiate-reinforced behaviours (Sutcliffe *et al.*, 2000; Fuchs & See, 2002; Shalev *et al.*, 2002). On the other hand, mice withdrawn from chronic morphine treatment display downregulation of α -synuclein mRNA in the NAcc and related brain areas, e.g. the basolateral amygdala, the dorsal striatum and the VTA (Ziolkowska *et al.*, 2005), in agreement with the observation reported in our present study for chronic cocaine. The changes were the most pronounced after longer periods of withdrawal, and were significantly increased in the striatum/accumbens for as long as 2 weeks after treatment cessation (Ziolkowska *et al.*, 2005).

Synucleins are not essential for basic synaptic functions (e.g. neurotransmitter release, synaptic vesicle numbers and pools, and synaptic plasticity), but are redundantly required for maintaining normal DA levels in the nigrostriatal system (Chandra *et al.*, 2004). No major phenotype was observed in α -synuclein knockout mice (Abeliovich *et al.*, 2000). A confounding problem in studying

α -synuclein is the presence of β - and γ -synucleins, highly homologous isoforms (Ueda *et al.*, 1993; Akopian & Wood, 1995; George *et al.*, 1995; Ji *et al.*, 1997). The α - and β -synucleins are widely co-localized in presynaptic nerve terminals throughout the brain, but absent from peripheral tissues (Maroteaux *et al.*, 1988). In contrast, γ -synuclein is abundant in specialized neurons, such as dorsal root ganglia (Akopian & Wood, 1995; Murphy *et al.*, 2000), and in non-neuronal tissues or certain tumour cells (Ji *et al.*, 1997; Buchman *et al.*, 1998; Surguchov *et al.*, 1999). The high degree of co-expression of α - and β -synuclein indicates the potential for functional redundancy, which may have obscured an essential role of α -synuclein in α -synuclein knockout mice. The tools we have developed circumvent these difficulties, and enable for very local gene transfer and very local changes of gene expression. Because α -synuclein is described as a widely presynaptic protein, the cocaine-mediated changes in mRNA and protein levels that we report in this study are likely to take place in the same populations of projection neurons. Axonal localization of α -synuclein was confirmed by immunofluorescent labelling. Other studies, however, described further increased mRNA expression in response to, e.g. morphine in distinct populations as well, whose somata are in different brain areas (Ziolkowska *et al.*, 2005).

From our studies, a role for α -synuclein in addiction to cocaine may clearly be postulated. However, addiction refers to a specific pattern of compulsive drug-seeking and drug-taking behaviour, one that predominates over most other activities in life (Robinson, 2004; Vanderschuren & Everitt, 2004). Deroche-Gamonet clearly identified addiction as the result from the interaction of two variables (Deroche-Gamonet *et al.*, 2004): (i) the degree of exposure to drugs (because addiction-like behaviour appears only after extended access to cocaine); and (ii) the degree of vulnerability in the exposed individual (because, despite a similar drug intake in all subjects, addiction-like behaviour appears only in a few). It is the interaction between a long exposure to drug and a vulnerable phenotype, not one or the other factor in itself, that seems to determine the development of addiction. The vulnerable phenotype greatly depends on local drug-mediated expression changes of a very large array of genes, as addiction is largely a multigenetic brain disorder. A sound characterization of the role of the different genes regulated upon addiction, as exemplified with the highly specific tools developed in this study for α -synuclein, is therefore essential for understanding the molecular mechanisms involved in this process. Our results demonstrate that α -synuclein is able to modulate cocaine-induced behavioural effects. This suggests that targeting α -synuclein function could provide new therapeutic strategies to treat cocaine abuse for which there is no available treatment.

Supplementary material

The following supplementary material may be found on <http://www.blackwell-synergy.com>

Fig. S1. Immunohistochemistry of α -synuclein and NeuN expression in the Nacc.

Fig. S2. Immunohistochemistry of α -synuclein expression in the Nacc.

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Abbreviations

DA, dopamine; DAT, dopamine transporter; FR, fixed ratio; GAPDH, glyceraldehyde-3-phosphate-dehydrogenase; GFP, green fluorescent protein; HEK, human embryonic kidney; NAcc, nucleus accumbens; PBS, phosphate-buffered saline; PCR, polymerase chain reaction; SA, self-administration; TBST, Tris-buffered Saline Tween-20; VTA, ventral tegmental area.

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