Dried Blood Spot Thyroglobulin as a Biomarker of Iodine Status in Pregnant Women

Sara Stinca,¹ Maria Andersson,¹ Sandra Weibel,¹ Isabelle Herter-Aeberli,¹ Ralph Fingerhut,² Sueppong Gowachirapant,³ Sonja Y. Hess,⁴ Nidhi Jaiswal,⁵ Tomislav Jukić,⁶ Zvonko Kusic,⁶ Ngoako Solomon Mabapa,⁷ Ashwini Kumar Nepal,⁸ Teofilo O. L. San Luis,⁹ Jia Qing Zhen,¹⁰ and Michael Bruce Zimmermann¹

¹Human Nutrition Laboratory, Institute of Food Nutrition and Health, ETH Zurich, Zurich 8092, Switzerland; ²Swiss Newborn Screening Laboratory, Children's Research Center (CRC), University Children's Hospital of Zurich, Zurich 8032, Switzerland; ³Institute of Nutrition, Mahidol University, Nakhon Pathom 73170, Thailand; ⁴Department of Nutrition, University of California, Davis, Davis, California 95616; ⁵St. John's Research Institute, St. John's National Academy of Health Sciences, Bangalore 560034, India; ⁶Department of Oncology and Nuclear Medicine, University of Zagreb School of Medicine, Sisters of Charity University Hospital Centre, Zagreb 10,000, Croatia; ⁷Department of Nutrition, University of Venda, Thohoyandou 0950, South Africa; ⁸Department of Biochemistry, B.P. Koirala Institute of Health Sciences, Ghopa, Dharan 56700, Nepal; ⁹Iodine Global Network (IGN), Manila 1102, Philippines; and ¹⁰Shanxi Institute for Endemic Disease Prevention and Treatment, LinFen 041000, China

Context: Thyroglobulin (Tg) could be a sensitive biomarker of iodine nutrition in pregnant women (PW). A dried blood spot (DBS) assay would simplify collection and transport in field studies.

Objectives: Our aims were to (1) establish and test a reference range for DBS-Tg in PW; (2) determine whether co-measurement of Tg antibodies (Abs) is necessary to define population iodine status.

Design, Setting, and Participants: Standardized cross-sectional studies of 3870 PW from 11 countries. For the DBS-Tg reference range, we included TgAb-negative PW (n = 599) from 3 countries with sufficient iodine intake.

Main Outcome Measures: We measured the urinary iodine concentration and DBS thyroidstimulating hormone, total thyroxin, Tg, and TgAb.

Results: In the reference population, the median DBS-Tg was 9.2 μ g/L (95% confidence interval, 8.7 to 9.8 μ g/L) and was not significantly different among trimesters. The reference range was 0.3 to 43.5 μ g/L. Over a range of iodine intake, the Tg concentrations were U-shaped. Within countries, the median DBS-Tg and the presence of elevated DBS-Tg did not differ significantly between all PW and PW who were TgAb-negative.

Conclusions: A median DBS-Tg of ~10 μ g/L with <3% of values ≥44 μ g/L indicated population iodine sufficiency. Concurrent measurement of TgAb did not appear necessary to assess the population iodine status. (*J Clin Endocrinol Metab* 102: 23–32, 2017)

Thyroglobulin (Tg) is a thyroid-specific protein and a storage and synthesis site for thyroid hormones (1). During iodine deficiency, an increase in thyroid size and/ or activity results in an increase in the blood Tg concentration (2, 3). In children and adults, Tg might be a

Copyright © 2017 by the Endocrine Society Received 28 July 2016. Accepted 6 October 2016. First Published Online 12 October 2016 sensitive biomarker of iodine nutrition and responds quickly to changes in iodine intake (2–6). Tg can be measured on dried blood spots (DBSs), which could simplify collection, storage, and transport in field studies (5). DBS-Tg has been recommended by the World Health

ISSN Print 0021-972X ISSN Online 1945-7197 Printed in USA

Abbreviations: Ab, antibody; CI, confidence interval; CV, coefficient of variation; DBS, dried whole-blood spot; IQR, interquartile range; PW, pregnant woman; Tg, thyro-globulin; TgAb, autoantibody against Tg; TSH, thyroid-stimulating hormone; TT4, total thyroxin; UIC, urinary iodine concentration; WHO, World Health Organization.

Organization (WHO) for monitoring of iodine status in school-age children (6). An international reference range has been established (4), and it might also be a sensitive biomarker of iodine excess in children (7).

DBS-Tg could also be a promising biomarker to assess iodine status in pregnant women (PW), in particular, because iodine requirements and the urinary iodine concentration (UIC) reference range for pregnancy remain uncertain. However, Tg data from PW with varying iodine intake are scarce and no international reference range is available. The available data suggest that Tg might be modestly elevated during pregnancy in both iodine-deficient and iodine-sufficient women (8, 9) owing to increased thyroid activity. Tg antibodies (TgAb) can confound the individual assessment of Tg in clinical monitoring of thyroid disorders. However, no data are available on whether co-measurement of TgAb, with Tg, is necessary in PW to define the population iodine status.

Therefore, our aims were (1) to establish, in iodinesufficient euthyroid TgAb-negative PW, an international reference range for DBS-Tg that could be used to monitor iodine nutrition; (2) to test the DBS-Tg reference range and assess thyroid function in PW over the range of iodine intake currently defined by the WHO/United Nations Children's Fund/International Council for Control of Iodine Deficiency Disorders as insufficient (median UIC, <150 µg/L), adequate (median UIC, 150 to 249 µg/L); and more-than-adequate (median UIC, 250 to 499 µg/L); and (3) to determine whether co-measurement of DBS-Tg antibodies (TgAbs) and DBS-Tg is necessary in population studies of iodine status in PW.

Subjects and Methods

Subjects

We included PW (n = 3870) in all 3 trimesters living in 11 countries: 2 in Europe (Croatia and Switzerland), 1 in North Africa and the Eastern Mediterranean (Morocco), 3 in Sub-Saharan Africa (Niger, South Africa, and Tanzania), 3 in Asia (India, Nepal, and China), and 2 in Southeast Asia (Thailand and the Philippines). We selected these countries to include PW with varying iodine status and provide regional and ethnic representation. All studies were local or regional studies, except for in Switzerland, where the study was nationally representative. The inclusion criteria were (1) healthy women aged 18 to 44 years; (2) singleton pregnancy; (3) nonsmoker; (4) no major chronic diseases; (5) no history of thyroid disease; and (6) no chronic use of medications. To establish the DBS-Tg reference range, we first included all women from countries in which the median UIC of the pregnant population was 150 to 299 μ g/L. We next included individual PW from this group who were euthyroid, TgAb-negative and not taking iodinecontaining supplements.

With the relative precision for the 97.5 percentile for DBS-Tg specified at 3% to 5% of the total length of the 95% reference

range and the estimated standard of DBS-Tg at 2.3 μ g/L (based on data from iodine-sufficient Swedish PW (9)), we estimated a sample size of ~540 PW would be required to establish the DBS-Tg reference range in PW with sufficient iodine intake (10). The ethical committee at each local institution involved in the study approved the study protocol. All the subjects provided informed written consent, and we collected the data from 2008 to 2016.

Study design

We recruited PW through clinics providing routine prenatal care. The participants completed a brief questionnaire on general health status, including year of birth, gestational week (from the date of the last menstrual period), trimester of pregnancy, singleton pregnancy, smoking, history of thyroid disease, major chronic disease, and use of medication or iodine supplements. A spot urine sample was collected and stored at -20° C until analysis. We spotted whole blood from a finger prick onto filter paper cards (Whatman 903; GE Healthcare, Little Chalfont, UK). After spotting, the DBS cards were dried at room temperature for 24 hours and then stored in sealed low-density bags at -20° C until analysis.

Laboratory analyses

We measured UICs using a modification of the Sandell-Kolthoff method (11) and laboratory-specific urine control material. All laboratories in the present study were certified by the Program to Ensure the Quality of Urinary Iodine Procedures (US Centers for Disease Control and Prevention, Atlanta, GA) and participated successfully in its quarterly external validation. We used the WHO/United Nations Children's Fund/International Council for Control of Iodine Deficiency Disorders criteria based on the median UIC to classify iodine nutrition in the population of PW at each site: insufficient, median UIC <150 μ g/L; adequate, median UIC 150 to 249 μ g/L; more-than-adequate, median UIC 250 to 499 μ g/L (6).

We measured DBS-Tg using a recently developed enzymelinked immunosorbent assay (12). At a DBS-Tg concentration of 27.2 \pm 4.8 µg/L and 59.9 \pm 14.7 µg/L, the interassay coefficient of variation (CV; n = 100) was 17.8% and 24.6%, respectively. We measured the serum TgAb concentration in Indian and Thai women using the Immulite 2000 TgAb kit (Immulite 2000, Siemens, Munich, Germany). The manufacturer's specified reference range is nondetectable to 40 U/mL. We measured DBS-TgAb concentrations using a serum enzyme-linked immunosorbent assay (TgAb enzyme-linked immunosorbent assay, version 2; RSR, Cardiff, UK) adapted in our laboratory for DBS. The intra-assay CV was 7% at 150 ± 50 U/mL and 7.8% at 520 ± 150 U/mL, respectively. The interassay CV was 13.8% at 150 ± 50 U/mL and 9.9% at 520 ± 150 U/mL. The manufacturer cutoff for TgAb positivity is ≥ 65 U/mL.

Using the GSP Neonatal hTSH kit (PerkinElmer Life Sciences, Turku, Finland), we measured DBS thyroid-stimulating hormone (TSH) and, using the GSP Neonatal T4 kit (PerkinElmer Life Sciences), DBS-total thyroxin (TT4) with the help of an automated time-resolved fluoroimmunoassay (13) at the Swiss Newborn Screening Laboratory, University Children's Hospital (Zurich, Switzerland). The TSH interassay CV was 11.2% at 15.2 \pm 1.7 mIU/L and 10.8% at 62.7 \pm 6.8 mIU/L. The TT4 interassay CV was 13.3% at 42.9 \pm 5.7 nmol/L,

12.3% at 100.7 \pm 12.4 nmol/L, and 11.9% at 166.4 \pm 19.8 nmol/L. The normal reference values for DBS-TT4 in nonpregnant adults with this assay are 65 to 165 nmol/L. For DBS-TT4 in PW in the first trimester, we used the reference range of 65 to 165 nmol/L. For the second and third trimesters, we multiplied the nonpregnant adult reference range by 1.5 and used the resulting range of 97.5 to 247.5 nmol/L (14). The normal reference values for DBS-TSH in nonpregnant adults with this assay are 0.1 to 3.7 mIU/L. For DBS-TSH in PW in the second and third trimesters, we used this reference range. Using the trimester-specific serum TSH reference ranges suggested by the American Thyroid Association (15) (*i.e.*, a serum TSH ~18% lower in the first than in second and third trimesters), we lowered the upper limit of our TSH assay to 3.0 mIU/L in the first trimester.

Statistical analyses

Data and statistical analyses were done with Excel 2011 (Microsoft Corp., Redmond, WA) and IBM SPSS Statistics for Mac, version 23 (IBM Corp., Armonk, NY). Outliers were investigated using the Tukey test, but no outliers were removed. Non-normally distributed data were log-transformed for analysis. For parameters with values between 0 and 1 (TSH and Tg), a constant of 1 was added to the values before transformation and was subtracted when back-transformed. Normally distributed data are presented as the mean ± standard deviation or geometrical mean [95% confidence interval (CI)] for log data. Non-normally distributed data are presented as the median [interquartile range (IQR)]. The nonparametric 95% CIs around the median and percentiles (2.5 and 97.5) were obtained using the bootstrap technique (n = 1000). To establish the DBS-Tg reference range, the 2.5 and 97.5 ranked percentiles were calculated, in accordance with the International Federation of Clinical Chemistry and Clinical and Laboratory Standards Institute (formerly, National Committee for Clinical Laboratory Standards) guidelines (16–18), using the following formulas: 0.025 \times (n + 1) and $0.975 \times (n + 1).$

We selected Indian (n = 398), South African (n = 384), and Nepalese (n = 159) PW as the reference populations because these sites had a median UIC of 150 to 299 μ g/L (6). PW were excluded if they did not have UIC data available (n = 36) or all thyroid parameters available (n = 247) or were TgAb-positive (n = 59). The final sample for the reference range was 599 PW. We used the Kruskal-Wallis analysis of variance or the Mann-Whitney *U* test to compare differences in UIC and thyroid hormone levels among the trimesters and between sites, and the Dunn-Bonferroni test for *post hoc* comparisons. To evaluate the associations between variables, Spearman's correlations were calculated.

For the remaining country data sets, we excluded 886 women because they did not meet the inclusion criteria (n = 208) and/or had no data available for UIC (n = 326) or for all thyroid parameters (n = 352). Thus, the final data set consisted of 2984 PW, and we measured TgAbs on 2015 of these PW. We used the Kruskal-Wallis analysis of variance and Mann-Whitney *U* test within countries and the generalized linear mixed effect model, with the trimester as a fixed factor and the country as a random factor for the pooled data sets (all sites) to compare the differences in UIC and thyroid hormone levels, followed by Bonferroni *post hoc* comparisons. We used independent *t* tests (with log-transformed data), the Mann-Whitney *U* test, and the

generalized linear mixed effect model to compare the median DBS-Tg between the TgAb-positive and TgAb-negative groups. Spearman's correlations were calculated, and multiple linear regression analysis was performed if ≥ 2 factors correlated with DBS-Tg or TgAbs. We used χ^2 tests and the binary logistic generalized linear mixed effect model followed by Bonferroni correction to test for differences in prevalence among trimesters and countries. For the countries for which we had data on gestational age, we performed binary logistic regression analysis for TgAb prevalence, with gestational age as the fixed factor and country as the random factor. Differences with P < 0.05 were considered statistically significant. We constructed scatterplots using individual values of PW (n = 2984) of DBS-Tg vs UIC, TSH, or TT4 and added Loess smoothed line calculations (with 70% of points to fit) to describe the best fit. Bubble charts showing DBS-Tg vs UIC, TSH, or TT4 were clustered using the sample size of each country, with a second-order polynomial trend line added to show the best fit.

Results

Reference range for DBS-Tg in pregnancy

Among the women in India, South Africa, and Nepal included in the reference population (n = 599), the median age was 23.2 years (IQR, 21.2 to 26.8), weight was 51.2 kg (IQR, 46.6 to 58.8), height was 154.6 cm (IQR, 150.4 to 158.4), and body mass index was 21.6 kg/m² (IQR, 19.8 to 24.2). The overall median UIC was 193.8 µg/L (IQR, 100.9 to 335.9). The pooled median DBS-Tg was 9.2 µg/L (IQR, 5.3 to 14.9), and the 95% CI around the median was 8.7 to 9.8 µg/L. The 2.5 and 97.5 DBS-Tg percentiles were 0.3 µg/L (95% CI, 0.2 to 0.6) and 43.5 µg/L (95% CI, 33.5 to 53.7). No statistically significant difference was found in DBS-Tg among trimesters (P = 0.245; Fig. 1). The median DBS-Tg was 9.7 μ g/L (IQR, 7.2 to 16.0) in the first trimester (n = 128), 9.4 μ g/L (IQR, 5.2 to 14.7) in the second trimester (n = 260), and 8.0 µg/L (IQR, 4.7 to 14.3) in the third trimester (n = 211). For Nepal and South Africa, we did not have data on the week of gestation, only trimester data; thus, we could not determine whether a relationship existed between gestational week and DBS-Tg in our reference population. The 2.5 and 97.5 DBS-Tg percentiles by trimester were 1.8 μ g/L (95% CI, 0.2 to 3.1) and 47.4 µg/L (95% CI, 28.9 to 70.3) in the first trimester, 0.3 µg/L (95% CI, 0.2 to 0.7) and 43.9 µg/L (95% CI, 29.8 to 56.3) in the second trimester, and 0.2 µg/L (95% CI, 0.1 to 0.5) and 42.4 µg/L (95% CI, 32.9 to 63.4) in the third trimester, respectively.

The prevalence of TgAb-positive PW was 9% (n = 658). The pooled median DBS-Tg in TgAb-positive women (n = 59) was 8.9 μ g/L (IQR, 4.8 to 13.6) and was not significantly different statistically than that in TgAb-negative women (n = 599; 9.2 μ g/L, IQR, 5.3 to 14.9 μ g/L; *P* = 0.543). The differences in the median



Figure 1. Box plot (median, interquartile range) of DBS-Tg derived from 599 euthyroid, TgAb-negative, pregnant women during the 3 trimesters of pregnancy (P = 0.245).

DBS-Tg in the TgAb-positive and TgAb-negative PW across trimesters also was not statistically significant: 5.5 μg/L (IQR, 2.4 to 9.6) vs 9.7 μg/L (IQR, 7.2 to 16.0) in the first trimester (P = 0.325); 11.6 µg/L (IQR, 6.8 to 18.4) vs 9.4 µg/L (IQR, 5.2 to 14.7) in the second trimester (P = 0.476) and 7.9 µg/L (IQR, 5.0 to 10.8) vs 8.0 μ g/L (IQR, 4.7 to 14.3) in the third trimester (P = 0.967). The pooled median DBS-TSH and DBS-TT4 were 0.8 mIU/L (IQR, 0.6 to 1.2) and 107.7 nmol/L (IQR, 85.2 to 137.0), respectively. The median DBS-TSH was greater in the first trimester (median, 1.1 mIU/L; IQR, 0.9 to 1.4) than in the second (median, 0.8 mIU/L; IQR, 0.6 to 1.2) and third (median, 0.8 mIU/L; IQR, 0.6 to 1.0) trimesters. The difference was statistically significant for both (P <0.01). The median DBS-TT4 did not differ among the trimesters (P = 0.088). The pooled data showed no overall relevant correlation between DBS-Tg and UIC (r_s = 0.042; P = not statistically significant) or between DBS-Tg and DBS-TSH ($r_s = 0.033$; P = not statistically significant). The only statistically significant positive overall correlation was found between DBS-Tg and DBS-TT4 $(r_{\rm s} = 0.126; P = 0.002).$

Assessment of iodine status in pregnancy using DBS-Tg and reference range

By country, Moroccan women had the lowest median UIC (31.5 μ g/L, IQR, 16.7 to 57.6) and Tanzanian women the highest (429.3 μ g/L, IQR, 270.1 to 614.9; Table 1). By country, Filipino women had the lowest median DBS-Tg (median, 6.4 μ g/L; IQR, 3.2 to 10.9), and Moroccan women had the highest median DBS-Tg (median, 62.4 μ g/L; IQR, 35.3 to 93.7; Table 1). The median age, gestational week, weight, height, and body mass index of the women are listed in Supplemental

Table 1. Figure 2 shows the median DBS-Tg plotted against the median UIC by country. The frequency of elevated DBS-Tg values was significantly greater in iodine-deficient and more-than-adequate countries than in iodine-sufficient countries (P < 0.05; Table 1).

Effect of DBS-TgAb on DBS-Tg

DBS-TgAb was measured in 2015 pregnant women. For the remaining 969 women, we did not have an adequate sample left on the DBS cards to measure TgAb after the analyses for the other thyroid parameters. The overall prevalence of TgAb-positive PW was 18.5% (n = 2015; Table 1). The overall percentage of TgAb-positive PW was not significantly different among trimesters: 15.2% in the first, 19.5% in the second, and 18.7% in the third trimester (P = 0.266; Table 1). We found no statistically significant difference in TgAb prevalence among gestational weeks (P = 0.274). The percentage of TgAbpositive PW was significantly greater in iodine-deficient countries (26.7%; n = 237) than in iodine-sufficient countries (12%; n = 135; P < 0.01). The overall median DBS-Tg concentration in TgAb-negative PW (n = 1643) was 16.8 µg/L (IQR, 8.7 to 30.8) and in TgAbpositive PW (n = 372) was 24.5 μ g/L (IQR, 12.9 to 40.7) (P = 0.252; Supplemental Fig. 1). No statistically significant association was seen between DBS-Tg and the titers of DBS-TgAb (P = 0.391). Within the countries, comparing TgAb-negative and TgAb-positive PW, the median DBS-Tg was significantly different statistically only in Thailand (P = 0.039; Supplemental Table 2). The overall median DBS-Tg (18.3 µg/L) in all PW (including TgAb-positive women) was not substantially greater than when TgAb-positive women were excluded (16.8 μ g/L; P = 0.683). Also, within all countries, we found no statistically significant difference comparing the median DBS-Tg in all PW to that in the TgAb-negative PW (P >0.05 for all countries; Fig. 3). Although the percentage of elevated Tg values in TgAb-negative PW was 13.0% (n = 213) and was significantly lower statistically than in TgAb-positive women (22.8%; n = 85; P < 0.001), the percentage of elevated DBS-Tg values in all PW (14.8%, n = 298) was not significantly different statistically from the percentage when the TgAb-positive women were excluded (13%, n = 213; *P* = 0.113).

Relationships between DBS-Tg and other thyroid function test results

The overall median DBS-TSH was 0.7 mIU/L (IQR, 0.5 to 1.0), and the overall median DBS-TT4 was 121.0 nmol/L (IQR, 90.0 to 158.7; Supplemental Table 3). No statistically significant difference was found in the pooled median UIC among trimesters. We found a statistically significant difference among trimesters for the pooled

Variables	Trimester of Pregnancy				
	First	Second	Third	All	
Morocco Pregnant women (n) UIC (µg/L) Tg (µg/L) Elevated Tg (>43.5 µg/L) TgAb-positive	51 35.7 ^a (21.3–60.0) 54.4 ^a (30.4–79.1) 62.7 (32) ^a 22.7 (5) ^a	118 28.9 ^a (14.5–57.4) 63.3 ^a (34.1–93.8) 66.9 (79) ^a 30.3 (20) ^a	76 30.8 ^a (16.6–52.8) 72.6 ^a (38.0–110.8) 68.4 (52) ^a 18.0 (9) ^a	245 31.5 (16.7–57.6) 62.4 (35.2–93.7) 66.5 (163) ^b 24.6 (34) ^c	
Pregnant women (n)	13	178	254	445	
UIC (µg/L)	81.3 ^a (43.6–156.8)	75.7 ^a (41.9–118.6)	63.9 ^a (31.4–109.5)	69.5 (37.6–115.9)	
Tg (µg/L)	26.5 ^a (22.2–35.8)	36.5 ^a (25.3–51.1)	33.5 ^a (23.1–48.7)	34.0 (23.7–49.3)	
Elevated Tg (>43.5 µg/L)	7.7 (1) ^a	35.4 (63) ^a	31.5 (80) ^a	32.4 (144) ^d	
TgAb-positive	0 (0)	51.7 (46) ^a	48.9 (69) ^a	49.4 (115) ^e	
Pregnant women (n)	96	98	98	292	
UIC (μg/L)	155.5a (117.9–203.4)	79.8 ^f (59.4–88.6)	74.7 ^{f.g} (68.4–85.9)	86.6 (71.6–119.5)	
Tg (μg/L)	8.5 ^a (5.9–12.6)	4.3 ^f (2.0–6.8)	6.6 ^{f.g} (3.1–12.4)	6.4 (3.2–10.9)	
Elevated Tg (>43.5 μg/L)	0 ^a	0 ^a	2.0 (2) ^a	0.7 (2) ^c	
TgAb-positive	NA	NA	NA	NA	
Pregnant women (n)	58	58	47	163	
UIC (μg/L)	145.0 ^a (90.3–223.0)	125.5 ^a (52.0–196.3)	132.0° (68.0–240.0)	135.0 (72.0–208.0)	
Tg (μg/L)	15.1 ^a (9.6–21.1)	13.0 ^a (8.2–20.9)	11.5° (7.7–19.2)	13.0 (8.5–20.9)	
Elevated Tg (>43.5 μg/L)	3.4 (2) ^a	1.7 (1) ^a	2.1 (1)°	2.5 (4) ^c	
TgAb-positive	NA	NA	NA	NA	
Pregnant women (n)	66	96	91	256	
UIC (μg/L)	137.3 ^a (70.7–309.8)	131.8 ^a (65.6–336.1)	158.1 ^a (57.6–311.7)	143.2 (67.2–323.8)	
Tg (μg/L)	26.8 ^a (17.0–37.8)	21.8 ^a (14.2–36.3)	24.2 ^a (15.5–38.0)	24.6 (15.8–37.1)	
Elevated Tg (>43.5 μg/L)	15.2 (10) ^a	14.6 (14) ^a	15.4 (14) ^a	15.0 (38) ^h	
TgAb-positive	24.2 (16) ^a	25.3 (24) ^a	20.9 (19) ^a	23.4 (59) ^c	
Pregnant women (n) UIC (μg/L) Tg (μg/L) Elevated Tg (>43.5 μg/L) TgAb-positive	NA NA NA NA	115 145.2 ^a (86.9–230.8) 8.5 ^a (4.9–13.8) 0.9 (1) ^a 15.9 (18) ^a	153 152.5 ^a (108.9–216.5) 8.5 ^a (5.7–13.7) 3.3 (5) ^a 7.3 (11) ^f	268 148.7 (96.2–220.9) 8.5 (5.4–13.7) 2.2 (6) ^c 11.0 (29) ^{b,i}	
Pregnant women (n)	7	95	105	207	
UIC (μg/L)	146.2 ^a (101.6–385.7)	197.5 ^a (107.2–291.2)	157.5 ^a (79.6–314.8)	174.0 (95.3–297.6)	
Tg (μg/L)	10.6 ^a (5.2–43.5)	7.3 ^a (4.1–14.7)	8.3 ^a (4.5–16.9)	8.0 (4.1–15.1)	
Elevated Tg (>43.5 μg/L)	14.3 (1) ^a	3.2 (3) ^a	3.8 (4) ^a	3.9 (8) ^c	
TgAb-positive	0 (0) ^a	4.3 (4) ^a	1.0 (1) ^a	2.5 (5) ^d	
N	113	124	102	339	
UIC (μ g/L)	204.2 ^a (117.7–346.9)	180.1 ^a (110.8–303.8)	157.9 ^a (87.5–275.3)	180.7 (102.8–307.1)	
Tg (μ g/L)	29.9 ^a (22.5–42.4)	24.3 ^f (17.9–33.7)	22.5 ^{f.g} (15.8–31.9)	25.9 (18.7–35.8)	
Elevated Tg (> 43.5 μ g/L)	23.9 (27) ^a	15.3 (19) ^{a,f}	6.9 (7) ^f	15.6 (53) ^h	
TgAb-positive	19.0 (16) ^a	21.0 (21) ^a	19.6 (18) ^a	19.9 (55) ^{c,h,i}	
N	133	111	63	307	
UIC (μ g/L)	159.2 ^a (94.7–313.8)	207.5 ^a (97.3–322.8)	145.0 ^a (70.8–307.1)	184.4 (92.1–315.3)	
Tg (μ g/L)	9.4 ^a (6.6–15.0)	8.8 ^a (4.6–12.9)	6.9 ^a (3.6–14.2)	9.0 (5.2–13.7)	
Elevated Tg (> 43.5 μ g/L)	1.5 (2) ^a	0.9 (1) ^a	0 ^a	1.0 (3) ^c	
TgAb-positive	7.6 (10) ^a	2.7 (3) ^a	3.3 (2) ^a	5.0 (15) ^{b,d}	
Pregnant women (n) UIC (μg/L) Tg (μg/L) Elevated Tg (>43.5 μg/L) TgAb-positive	NA NA NA NA	85 310.3 ^a (165.5–427.0) 12.7 ^a (8.6–18.6) 3.5 (3) ^a 24.7 (21) ^a	71 274.3 ^a (151.5–397.0) 8.8 ^f (7.0–12.8) 1.4 (1) ^a 25.4 (18) ^a	156 290.1 (161.5–403.8) 10.7 (7.4–15.1) 2.6 (4) ^c 25.0 (39) ^c (<i>Continued</i>)	

Table 1. UIC and DBS-Tg in Pregnant Women Stratified by Country and Trimester

Variables	Trimester of Pregnancy				
	First	Second	Third	All	
Tanzania					
Pregnant women (n)	16	159	131	306	
UIC (µq/L)	474.0 ^a (409.7–586.3)	408.7 ^a (267.6–610.8)	439.4 ^a (254.8–629.6)	429.3 (270.1–614.9)	
Ta (µa/L)	20.9 ^a (15.7–36.0)	24.9 ^a (17.2–35.6)	28.9 ^a (19.8–40.3)	25.8 (18.2–38.4)	
Elevated Tg (>43.5 μ g/L)	18.8 (3) ^a	15.1 (24) ^a	18.3 (24) ^a	16.7 (51) ^h	
TgAb-positive	$20.0(2)^{a}$	9.1 (9) ^a	$12.0(10)^{a}$	10.9 (21) ^{6,h,i}	
All sites					
Pregnant women (n)	553	1237	1191	2984	
UIC (µa/L)	152.2 ^a (81.2–257.2)	133.1 ^a (65.6–288.7)	119.6 ^a (63.5–255.7)	132.4 (68.3–269.4)	
Ta (µa/L)	17.2 ^a (9.2–31.1)	18.7 ^a (8.9–33.4)	19.4 ^f (8.7–34.2)	18.8 (8.9–33.3)	
Elevated Tg (>43.5 μ g/L)	14.1 (78) ^a	$16.8(208)^{a}$	$16.0(190)^{a}$	16.0 (476)	
TgAb-positive	15.2 (49) ^a	19.5 (166) ^a	18.7 (157) ^a	18.5 (372)	

Table 1. Continued

Data presented as median (IQR) or % (n).

Abbreviation: NA, not available.

^{*a-i*}Statistically significant differences for data in the same row or column with different superscript letters [P < 0.05; Kruskal-Wallis test within countries with Dunn-Bonferroni test for *post hoc* comparisons; generalized mixed effect model for pooled data sets (all sites)] with trimester as fixed factor and country as random factor, with Bonferroni test for *post hoc* comparisons; χ^2 test followed by *z*-test for proportions].

median DBS-TSH (P = 0.045), median DBS-TT4 (P <0.01), and median DBS-Tg (P = 0.018). Although we did not have data on the gestational week for all women, in those women for whom we had gestational age data (n = 2437), the gestational week was a statistically significant predictor of DBS-Tg (P < 0.001). The prevalence of thyroid dysfunction, stratified by trimester and site, is presented in Supplemental Table 4. The pooled DBS-Tg value correlated statistically significantly and positively with the DBS-TT4 value ($r_s = 0.464$, P < 0.01) and the DBS-TSH value ($r_s = 0.247, P < 0.01$) and negatively with the UIC ($r_s = -0.164$, P < 0.01; Supplemental Table 5). The overall regression of TT4, TSH, and UIC on DBS-Tg was statistically significant ($R^2 = 0.221$; P < 0.001). The standardized coefficient was $\beta = 0.400$ for TT4, $\beta = 0.070$ for TSH, and $\beta = -0.159$ for UIC. The multiple regression for TT4, TSH, and UIC on DBS-Tg was statistically significant in the second and third trimesters (P < 0.001): $R^2 = 0.236, \beta = 0.395$ for TT4, $\beta = 0.100$ for TSH, and $\beta = -0.189$ for UIC in the second trimester; and R² = 0.255, $\beta = 0.406$ for TT4, $\beta = 0.118$ for TSH, and $\beta = -0.133$ for UIC in the third trimester. In the first trimester, linear regression of DBS-Tg on TT4 and TSH was conducted. The regression of TT4 and TSH on DBS-Tg was statistically significant (P < 0.001): $R^2 = 0.201$, $\beta = 0.405$ for TT4 and β = 0.088 for TSH. Pooled TgAb correlated significantly and positively with DBS-TSH ($r_s = 0.207, P < 0.01$), DBS-TT4 ($r_s = 0.421, P < 0.01$), DBS-Tg ($r_s = 0.383, P < 0.01$), and country ($r_s = 0.233$, P < 0.01) and negatively with UIC $(r_s = -0.96, P < 0.01)$. The overall regression of TgAb on TT4, Tg, and TSH was statistically significant ($R^2 = 0.304$, β = 0.421 for TT4, β = 0.153 for Tg, and β = 0.123 for TSH; P < 0.01).

Figure 4A and Supplemental Figs. 2A and 3A show the plots of DBS-Tg, DBS-TSH, and DBS-TT4 against UIC, including the Loess smoothed line depicting the best fit. To show the influence of iodine intake on thyroid function, we plotted the median UIC against the median DBS-Tg, DBS-TSH, and DBS-TT4 in the bubble plots (clustered by country; Fig. 4B and Supplemental Figs. 2B and 3B).

Discussion

The proposed Tg reference ranges in iodine-sufficient areas for adults and children are generally in the range of 4 to 40 μ g/L (4, 19). In the present study, we have proposed a reference range for DBS-Tg of 0.3 to $43.5 \,\mu g/L$, slightly wider, but otherwise similar, to the Tg reference range proposed for other population groups (4, 19). These data suggest that a population of PW with a prevalence of <3% of DBS-Tg \geq 44 µg/L could be categorized as iodine sufficient. In addition, from the pooled median DBS-Tg in the reference population of 9.2 µg/L and a 95% CI of 8.7 to 9.8 µg/L, a target median DBS-Tg of $\sim 10 \,\mu$ g/L can be used to categorize iodine sufficiency in a population of PW, a value comparable to those proposed for nonpregnant adult populations (2, 20). Previous studies of iodine-deficient PW (defined by a UIC $<150 \mu g/L$) have reported a median serum Tg of $\geq 10 \ \mu g/L \ (21-23)$ across trimesters. Similarly, previous studies of PW in iodine-deficient areas in which the UIC was not measured also support this median Tg value (24-26). In our reference population, the DBS-Tg concentrations did not differ significantly among the trimesters; thus, we have recommended a single reference range for all 3 trimesters.



Figure 2. Box plot (median, interquartile range) of DBS-Tg vs median UIC stratified by country. Countries were ranked from lowest median UIC (Morocco, 31.5 μ g/L) to highest median UIC (Tanzania, 429.3 μ g/L). The top dotted line represents the 97.5 percentile upper reference limit. The middle, solid line represents the median (50th percentile). The bottom dotted line represents the 2.5 percentile lower reference limit.

In a range from severely deficient (Morocco) to morethan-adequate iodine intake (Tanzania), the median DBS-Tg showed a shallow U-shaped curve (Fig. 4). An increasing severity of iodine deficiency was associated with higher DBS-Tg and greater prevalence of elevated DBS-Tg. Greater Tg concentrations in iodine-deficient PW likely reflect an increase in thyroid activity and/or size (2, 3) to meet the increased maternal-fetal requirement for thyroid hormone despite the lack of iodine (8, 27, 28). The mechanism of the increase in Tg at high iodine intakes is uncertain but might involve a failure of the thyroid to escape from the Wolff-Chaikoff effect, resulting in inhibition of thyroid peroxidase and reduced Tg proteolysis (29). Our data suggest that Tg could be a sensitive functional indicator of both iodine deficiency and excess in PW and could be used in conjunction with UIC, the recommended exposure biomarker (6).

The prevalence of TgAb positivity among PW has been reported to vary from 2.3% to 20% (30, 31). In our data, the overall prevalence was 18.5%. Although thyroid autoimmunity typically wanes during the course of gestation (32–34), we did not find a consistent pattern of a lower prevalence of TgAbs in later gestation. The overall percentage of TgAb-positive PW was not significantly different among trimesters. In some countries (India, China, Switzerland, Thailand), a decline was seen in TgAb prevalence during gestation; however, in other countries (Nepal, South Africa, Morocco, Niger), an increase was seen. The prevalence of TgAbs was significantly greater in the women from iodine-deficient countries compared with those from iodine-sufficient countries. This is in contrast to studies of nonpregnant women, in whom the frequency of thyroid autoimmunity tends to be higher in iodine-sufficient than in iodine-deficient populations (35).

The cost of measuring TgAbs in large field studies to assess iodine status can only be justified if their concurrent measurement improves the estimates of DBS-Tg. Our data suggest concomitant TgAb measurement is unnecessary. Within countries, comparing all PW with those who were TgAb-negative (*i.e.*, excluding TgAbpositive women), no statistically significant difference was seen in the median DBS-Tg or in the prevalence of elevated DBS-Tg values.

In our study, among PW in the countries with sufficient or more-than-adequate iodine intake, no statistically significant correlation was found between Tg and UIC or



Figure 3. Box plot (median, interquartile range) of DBS-Tg for TgAb-positive and TgAb-negative (gray) vs TgAb-negative only (white) stratified by country.

TSH or TT4, in agreement with previous studies of PW (8, 36–39). This might result by the iodine-deficient thyroid upregulating thyroid hormone synthesis and maintaining euthyroidism through mechanisms independent of TSH (40, 41). In contrast, in the iodine-deficient countries, Tg substantially and positively correlated with TSH in Morocco and the Philippines, but not in Niger. In contrast, Tg substantially and negatively correlated with UIC only in Morocco. Previous studies of correlations between Tg and UIC in iodine-deficient PW found a negative correlation (8, 21) or no statistically significant correlation (38, 42). The lack of correlation between Tg and UIC might result from confounding by the variable amounts of iodine stored in the thyroid in iodinedeficient pregnant women that can contribute to thyroid hormone synthesis and/or the wide interday variability in DBS-UIC (43).

The strengths of the present study include the international study population, the large sample size with wide variations in iodine status, the use of standardized methods for the UIC and thyroid function tests, and the co-measurement of TgAbs. A limitation of our study was that we used a single spot UIC, expressed as μ g/L, to categorize iodine status, in accordance with the WHO recommendations (6). We tried to at least partially overcome this limitation by assessing the median DBS-Tg in populations of PW living in areas with varying iodine status and relating this to the median UIC. However, as discussed, this could have limited our ability to find correlations between UIC and Tg, except in the countries with the lowest iodine intake, where the day-to-day UIC might have been less variable. Also, we did not measure creatinine in the urine samples and express the iodine concentration per unit of creatinine, which might have improved our individual classification of iodine status (44). The use of DBS simplifies the collection and transport of samples, which can lower field costs compared with the use of serum assays. However, if the DBSs are collected poorly or incompletely dried, that could increase the variation. Another caveat is that large variations exist between different serum assays for Tg, limiting direct comparisons between our findings and studies using different Tg assays. We are uncertain why in 2 of the countries in our study with successful iodine programs, China and Switzerland, the median DBS-Tg was $>10 \mu g/L$. Despite these limitations, our findings



Figure 4. (A) Scatterplot (using individual values of 2984 pregnant women) of DBS-Tg vs UIC, with Loess smoothed line added to show best fit. Data are presented on a log scale for both UIC and DBS-Tg. (B) Bubble chart (clustered by country) of median DBS-Tg vs median UIC with a second-order polynomial trend line. The size of the bubbles reflects the sample size for each country.

suggest that the use of DBS-Tg in pregnancy, together with UIC, to define iodine status could be a promising approach. If Tg proves to be a sensitive biomarker of deficient and excess iodine intake in pregnancy, its future use might allow refinement of the current range of median UIC to define adequate iodine status in PW.

Acknowledgments

We thank Vincent Assey, Susanne Dold, Jessica Farebrother, Césaire Ouédraogo, Joshua Saltiban, Ryan Wessells, and Rebecca Young, the field workers who participated in the sample collection, and the participating pregnant women.

Address all correspondence and requests for reprints to: Sara Stinca, PhD, Human Nutrition Laboratory, Institute of Food Nutrition and Health, ETH Zurich, Schmelzbergstrasse 7, LFV D 27.2, Zurich 8092, Switzerland. E-mail: sara.stinca@hest.ethz.ch.

This work was funded by the Swiss State Secretariat for Education, Research and Innovation (Bern, Switzerland), Global Alliance for Improved Nutrition, the United Nations Children's Fund, and the Human Nutrition Laboratory, Institute of Food, Nutrition and Health, ETH Zurich (Zurich, Switzerland). Administrative support was provided by the European Union Horizon 2020 project (EUThyroid 15.0146).

Disclosure Summary: The authors have nothing to disclose.

References

- Okosieme OE, Lazarus JH. Thyroglobulin: a thyroid autoantigen and marker of DTC. Available at: http://www.cli-online.com/ fileadmin/pdf/pdf_general/thyroglobulin-a-thyroid-autoantigen-andmarker-of-dtc.pdf. Accessed 25 August, 2015.
- Vejbjerg P, Knudsen N, Perrild H, Laurberg P, Carlé A, Pedersen IB, Rasmussen LB, Ovesen L, Jørgensen T. Thyroglobulin as a marker of iodine nutrition status in the general population. *Eur J Endocrinol.* 2009;161(3):475–481.
- Knudsen N, Bülow I, Jørgensen T, Perrild H, Ovesen L, Laurberg P. Serum Tg—a sensitive marker of thyroid abnormalities and iodine deficiency in epidemiological studies. J Clin Endocrinol Metab. 2001;86(8):3599–3603.
- 4. Zimmermann MB, de Benoist B, Corigliano S, Jooste PL, Molinari L, Moosa K, Pretell EA, Al-Dallal ZS, Wei Y, Zu-Pei C, Torresani T. Assessment of iodine status using dried blood spot thyroglobulin: development of reference material and establishment of an international reference range in iodine-sufficient children. *J Clin Endocrinol Metab.* 2006;91(12):4881–4887.
- Zimmermann MB, Moretti D, Chaouki N, Torresani T. Development of a dried whole-blood spot thyroglobulin assay and its evaluation as an indicator of thyroid status in goitrous children receiving iodized salt. Am J Clin Nutr. 2003;77(6):1453–1458.
- 6. World Health Organization, United Nations Children's Fund, International Council for Control of Iodine Deficiency Disorders. Assessment of Iodine Deficiency Disorders and Monitoring Their Elimination: A Guide for Programme Managers. 3rd ed. Geneva, Switzerland: World Health Organization; 2007.
- Zimmermann MB, Aeberli I, Andersson M, Assey V, Yorg JAJ, Jooste P, Jukić T, Kartono D, Kusić Z, Pretell E, San Luis TOL Jr, Untoro J, Timmer A. Thyroglobulin is a sensitive measure of both deficient and excess iodine intakes in children and indicates no adverse effects on thyroid function in the UIC range of 100-299 μg/L: a UNICEF/ICCIDD study group report. *J Clin Endocrinol Metab.* 2013;98(3):1271–1280.
- Eltom A, Elnagar B, Elbagir M, Gebre-Medhin M. Thyroglobulin in serum as an indicator of iodine status during pregnancy. *Scand J Clin Lab Invest*. 2000;60(1):1–7.
- Soldin OP, Tractenberg RE, Hollowell JG, Jonklaas J, Janicic N, Soldin SJ. Trimester-specific changes in maternal thyroid hormone, thyrotropin, and thyroglobulin concentrations during gestation: trends and associations across trimesters in iodine sufficiency. *Thyroid.* 2004;14(12):1084–1090.
- Armitage P, Berry G, Matthews JN. Statistical Methods in Medical Research. Oxford, UK: Blackwell Scientific Publications; 2002:397–399.
- Pino S, Fang SL, Braverman LE. Ammonium persulfate: a safe alternative oxidizing reagent for measuring urinary iodine. *Clin Chem.* 1996;42(2):239–243.
- Stinca S, Andersson M, Erhardt J, Zimmermann MB. Development and validation of a new low-cost enzyme-linked immunoassay for serum and dried blood spot thyroglobulin. *Thyroid*. 2015;25(12): 1297–1305.
- Fingerhut R, Torresani T. Evaluation of the genetic screening processor (GSP (TM)) for newborn screening. *Anal Methods*. 2013; 5(18):4769–4776.
- De Groot L, Abalovich M, Alexander EK, Amino N, Barbour L, Cobin RH, Eastman CJ, Lazarus JH, Luton D, Mandel SJ, Mestman J, Rovet J, Sullivan S. Management of thyroid dysfunction during pregnancy and postpartum: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab. 2012;97(8):2543–2565.
- 15. Stagnaro-Green A, Abalovich M, Alexander E, Azizi F, Mestman J, Negro R, Nixon A, Pearce EN, Soldin OP, Sullivan S, Wiersinga W; American Thyroid Association Taskforce on Thyroid Disease During Pregnancy and Postpartum. Guidelines of the American Thyroid Association for the diagnosis and management of thyroid disease during pregnancy and postpartum. *Thyroid*. 2011;21(10): 1081–1125.

- Linnet K. Nonparametric estimation of reference intervals by simple and bootstrap-based procedures. *Clin Chem.* 2000;46(6 Pt 1): 867–869.
- Solberg HE. International Federation of Clinical Chemistry (IFCC), Scientific Committee, Clinical Section, Expert Panel on Theory of Reference Values, and International Committee for Standardization in Haematology (ICSH), Standing Committee on Reference Values. Approved recommendation (1986) on the theory of reference values. Part 5. Statistical treatment of collected reference values. Determination of reference limits. J Clin Chem Clin Biochem. 1987;25(5):645–656.
- Clinical and Laboratory Standards Institute. Defining, Establishing and Verifying Reference Intervals in the Cinical Laboratory. Approved Guidelines. EP28-A3C. 3rd ed. Wayne, PA: Clinical and Laboratory Standards Institute; 2010:1–72.
- National Academy of Clinical Biochemistry. Laboratory medicine practice guidelines. In: Demers LM, Spencer CA, eds: Laboratory Support for the Diagnosis and Monitoring of Thyroid Disease. Washington, DC: American Association for Clinical Chemistry; 2003:1–125.
- 20. World Health Organization, United Nations Children's Fund, International Council for Control of Iodine Deficiency Disorders. Indicators for Assessing Iodine Deficiency Disorders and Their Control Through Salt Iodization. Geneva, Switzerland: World Health Organization; 1994.
- Moreno-Reyes R, Glinoer D, Van Oyen H, Vandevijvere S. High prevalence of thyroid disorders in pregnant women in a mildly iodine-deficient country: a population-based study. J Clin Endocrinol Metab. 2013;98(9):3694–3701.
- 22. Koukkou E, Kravaritis S, Mamali I, Markantes GG, Michalaki M, Adonakis GG, Georgopoulos NA, Markou KB. No increase in renal iodine excretion during pregnancy: a telling comparison between pregnant women and their spouses. *Hormones (Athens)*. 2014;13(3):375–381.
- Ma ZF, Skeaff SA. Thyroglobulin as a biomarker of iodine deficiency: a review. *Thyroid*. 2014;24(8):1195–1209.
- Kung AWC, Lao TT, Chau MT, Tam SCF, Low LCK. Goitrogenesis during pregnancy and neonatal hypothyroxinaemia in a borderline iodine sufficient area. *Clin Endocrinol (Oxf)*. 2000; 53(6):725–731.
- 25. Delshad H, Touhidi M, Abdollahi Z, Hedayati M, Salehi F, Azizi F. Inadequate iodine nutrition of pregnant women in an area of iodine sufficiency. *J Endocrinol Invest*. 2016;**39**(7):755–762.
- Domínguez I, Reviriego S, Rojo-Martínez G, Valdés MJ, Carrasco R, Coronas I, López-Ojeda J, Pacheco M, Garriga MJ, García-Fuentes E, González Romero S, C-Soriguer Escofet FJ. Iodine deficiency and thyroid function in healthy pregnant women [in Spanish]. *Med Clin (Barc)*. 2004;122(12):449–453.
- Glinoer D, de Nayer P, Bourdoux P, Lemone M, Robyn C, van Steirteghem A, Kinthaert J, Lejeune B. Regulation of maternal thyroid during pregnancy. J Clin Endocrinol Metab. 1990;71(2):276–287.
- Glinoer D. The regulation of thyroid function in pregnancy: pathways of endocrine adaptation from physiology to pathology. *Endocr Rev.* 1997;18(3):404–433.
- 29. Leung AM, Braverman LE. Consequences of excess iodine. *Nat Rev Endocrinol.* 2014;10(3):136–142.

- Marwaha RK, Chopra S, Gopalakrishnan S, Sharma B, Kanwar RS, Sastry A, Singh S. Establishment of reference range for thyroid hormones in normal pregnant Indian women. *BJOG*. 2008;115(5):602–606.
- Hollowell JG, Staehling NW, Flanders WD, Hannon WH, Gunter EW, Spencer CA, Braverman LE. Serum TSH, T(4), and thyroid antibodies in the United States population (1988 to 1994): National Health and Nutrition Examination Survey (NHANES III). J Clin Endocrinol Metab. 2002;87(2):489–499.
- Glinoer D, Riahi M, Grün JP, Kinthaert J. Risk of subclinical hypothyroidism in pregnant women with asymptomatic autoimmune thyroid disorders. J Clin Endocrinol Metab. 1994;79(1):197–204.
- Kuijpens JL, Pop VJ, Vader HL, Drexhage HA, Wiersinga WM. Prediction of postpartum thyroid dysfunction: can it be improved? *Eur J Endocrinol.* 1998;139(1):36–43.
- 34. Smyth PP, Wijeyaratne CN, Kaluarachi WN, Smith DF, Premawardhana LD, Parkes AB, Jayasinghe A, de Silva DG, Lazarus JH. Sequential studies on thyroid antibodies during pregnancy. *Thyroid*. 2005;15(5):474–477.
- Zimmermann MB, Boelaert K. Iodine deficiency and thyroid disorders. *Lancet Diabetes Endocrinol.* 2015;3(4):286–295.
- 36. Amouzegar A, Khazan M, Hedayati M, Azizi F. An assessment of the iodine status and the correlation between iodine nutrition and thyroid function during pregnancy in an iodine sufficient area. *Eur J Clin Nutr.* 2014;68(3):397–400.
- 37. Fuse Y, Ohashi T, Yamaguchi S, Yamaguchi M, Shishiba Y, Irie M. Iodine status of pregnant and postpartum Japanese women: effect of iodine intake on maternal and neonatal thyroid function in an iodine-sufficient area. J Clin Endocrinol Metab. 2011;96(12): 3846–3854.
- Brucker-Davis F, Ferrari P, Gal J, Berthier F, Fenichel P, Hieronimus S. Iodine status has no impact on thyroid function in early healthy pregnancy. *J Thyroid Res.* 2012;2012:168764.
- 39. Raverot V, Bournaud C, Sassolas G, Orgiazzi J, Claustrat F, Gaucherand P, Mellier G, Claustrat B, Borson-Chazot F, Zimmermann M. Pregnant French women living in the Lyon area are iodine deficient and have elevated serum thyroglobulin concentrations. *Thyroid*. 2012;22(5):522–528.
- Mitchell ML, Klein RZ, Sargent JD, Meter RA, Haddow JE, Waisbren SE, Faix JD. Iodine sufficiency and measurements of thyroid function in maternal hypothyroidism. *Clin Endocrinol* (Oxf). 2003;58(5):612–616.
- Morreale de Escobar G, Obregón MJ, Escobar del Rey F. Is neuropsychological development related to maternal hypothyroidism or to maternal hypothyroxinemia? *J Clin Endocrinol Metab.* 2000; 85(11):3975–3987.
- 42. Brough L, Jin Y, Shukri NH, Wharemate ZR, Weber JL, Coad J. Iodine intake and status during pregnancy and lactation before and after government initiatives to improve iodine status, in Palmerston North, New Zealand: a pilot study. *Matern Child Nutr.* 2015; 11(4):646–655.
- Zimmermann MB, Andersson M. Assessment of iodine nutrition in populations: past, present, and future. Nutr Rev. 2012;70(10):553–570.
- 44. Vejbjerg P, Knudsen N, Perrild H, Laurberg P, Andersen S, Rasmussen LB, Ovesen L, Jørgensen T. Estimation of iodine intake from various urinary iodine measurements in population studies. *Thyroid.* 2009;**19**(11):1281–1286.