

# Effects of wheat-flour biscuits fortified with iron and EDTA, alone and in combination, on blood lead concentration, iron status, and cognition in children: a double-blind randomized controlled trial<sup>1</sup>

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## ABSTRACT

**Background:** Lead is a common neurotoxicant and its absorption may be increased in iron deficiency (ID). Thus, iron fortification to prevent ID in populations is a promising lead mitigation strategy. Two common fortificants are ferrous sulfate (FeSO<sub>4</sub>) and ferric sodium EDTA (NaFeEDTA). EDTA can chelate iron and lead.

**Objectives:** Our study objective was to determine the effects of iron and EDTA, alone and in combination, on blood lead (BPb) concentration, iron status, and cognition.

**Design:** In this 2 × 2 factorial, double-blind placebo-controlled trial, 457 lead-exposed Moroccan children were stratified by school and grade and randomly assigned to consume biscuits (6 d/wk at school) containing 1) ~8 mg Fe as FeSO<sub>4</sub>, 2) ~8 mg Fe as NaFeEDTA that contained ~41 mg EDTA, 3) ~41 mg EDTA as sodium EDTA (Na<sub>2</sub>EDTA), or 4) placebo for 28 wk. The primary outcome was BPb concentration; secondary outcomes were iron status and cognitive outcomes from subtests of the Kaufman Assessment Battery for Children and the Hopkins Verbal Learning Test. These outcomes were measured at baseline and endpoint. All data were analyzed by intention-to-treat.

**Results:** The adjusted geometric mean BPb concentration at baseline was 4.3 μg/dL (95% CI: 4.2, 4.3 μg/dL), and at endpoint these values were 3.3 μg/dL (95% CI: 3.1, 3.5 μg/dL) for FeSO<sub>4</sub>, 2.9 μg/dL (95% CI: 2.7, 3.0 μg/dL) for NaFeEDTA, 3.3 μg/dL (95% CI: 3.1, 3.5 μg/dL) for EDTA, and 3.7 μg/dL (95% CI: 3.5, 3.9 μg/dL) for placebo. We found an effect of iron (*P* = 0.009) and EDTA (*P* = 0.012) for reduced BPb concentrations at endpoint, but no iron × EDTA interaction. Iron fortification improved iron status, but there were no positive effects of iron or EDTA on cognitive test scores.

**Conclusions:** Food fortification with iron and EDTA additively reduces BPb concentrations. Our findings suggest that NaFeEDTA should be the iron fortificant of choice in lead-exposed populations. This trial was registered at [clinicaltrials.gov](http://clinicaltrials.gov) as NCT01573013. *Am J Clin Nutr* 2016;104:1318–26.

**Keywords:** NaFeEDTA, cognition, fortification, iron, lead

## INTRODUCTION

Lead exposure and iron deficiency anemia (IDA)<sup>7</sup> are major global health problems that are concentrated in young children and

may irreversibly impair neurodevelopment (1, 2). Lead is a well-known neurotoxicant, and low-level lead exposure remains a public health problem not only in developing but also in developed countries (3, 4). Even low-level exposure is associated with a decrease in intelligence quotient (IQ), and the steepest decline in IQ is predicted by blood lead (BPb) concentration increments in the lower BPb range of 2.0–10 μg/dL (1, 2, 5). Recent estimations suggest that BPb concentrations as low as 0.1–1.0 μg/dL are associated with a decrease in IQ (3). As a result, the WHO has withdrawn its provisional tolerable upper intake limit for lead (6).

Lead exposure in children occurs largely through oral ingestion (2, 7). Increasing dietary iron may benefit lead-exposed populations because the absorption of lead may be increased in iron deficiency (ID), although not all studies agree (8, 9). There are likely multiple absorptive pathways for oral lead (10), but iron and lead share a common intestinal transporter—divalent metal transporter 1 (DMT1)—which is upregulated during ID (11). In addition, because DMT1 has a higher affinity for iron than lead, the presence of luminal iron may competitively inhibit this pathway of lead absorption (12, 13). A longitudinal study reported a 4- to 5-fold increased risk of subsequent lead poisoning in iron-deficient children (14). However, in an intervention trial in Mexican children, iron supplements did not reduce BPb concentrations (15). The CDC recommends an iron-rich diet for

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<sup>7</sup> Abbreviations used: AGP, α<sub>1</sub>-acid glycoprotein; BPb, blood lead; CRP, C-reactive protein; DMT1, divalent metal transporter 1, FeSO<sub>4</sub>, ferrous sulfate; HVL, Hopkins Verbal Learning Test; ID, iron deficiency; IDA, iron deficiency anemia; IQ, intelligence quotient; KABC-II, Kaufman Assessment Battery for Children, Second Edition; Na<sub>2</sub>EDTA, sodium EDTA; NaFeEDTA, ferric sodium EDTA; SF, serum ferritin; TfR, transferrin receptor; ZnPP, zinc protoporphyrin.

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lead-exposed children but has emphasized the need for randomized trials (16).

Sodium iron EDTA (NaFeEDTA) is a recommended iron fortificant for foods (6). EDTA is a metal chelator that binds both iron and lead (17). Given intravenously, it is an effective BPb chelator (18). Orally administered EDTA can chelate lead in the gut lumen and reduce its absorption (9, 19). Although <5% of oral EDTA is absorbed (20), small amounts of absorbed EDTA could possibly chelate BPb and increase its clearance. The affinity of EDTA for metals is pH-dependent: binding to ferric iron is favored at a low gastric pH, but in the alkaline duodenum the iron is released and in the distal gut, at near neutral pH, the affinity of EDTA is higher for other metals, including lead (17). Therefore, our study aimed to determine the effect of iron and EDTA, alone and in combination, on BPb concentration, iron status, and cognition in lead-exposed Moroccan children. We hypothesized that 1) both iron and EDTA would reduce BPb concentrations and 2) iron and EDTA would have an additive effect to reduce BPb concentrations.

## METHODS

### Study design and participants

We conducted this randomized  $2 \times 2$  factorial, double-blind placebo-controlled trial from September 2011 to July 2012 at 2 schools near Marrakesh, Morocco (clinicaltrials.gov; identification NCT01573013). We screened 479 children for eligibility. Our inclusion criteria were as follows: 1) preschool-aged third-through sixth-graders, who were 2) apparently healthy, 3) had hemoglobin concentrations  $>7$  g/dL, and 4) did not use iron supplements. We invited 457 children who met these criteria to join the study between September 2011 and January 2012. We obtained written informed consent from parents and verbal consent from the children. The Ministry of Health in Rabat, Morocco, and the ethical committee of ETH Zurich, Switzerland, approved the study protocol. The study protocol complied with the Declara-

tion of Helsinki and local laws and regulations. A data safety board monitored the study.

### Randomization and masking

We randomly assigned the eligible children, stratified by school and grade, by using computer-generated randomization sequences to 4 groups who received biscuits containing one of the following fortificants: 1)  $\sim 8$  mg Fe as ferrous sulfate ( $\text{FeSO}_4$ ), 2)  $\sim 8$  mg Fe as NaFeEDTA containing  $\sim 41$  mg EDTA, 3)  $\sim 41$  mg EDTA as sodium EDTA ( $\text{Na}_2\text{EDTA}$ ), or 4) placebo (**Figure 1**).

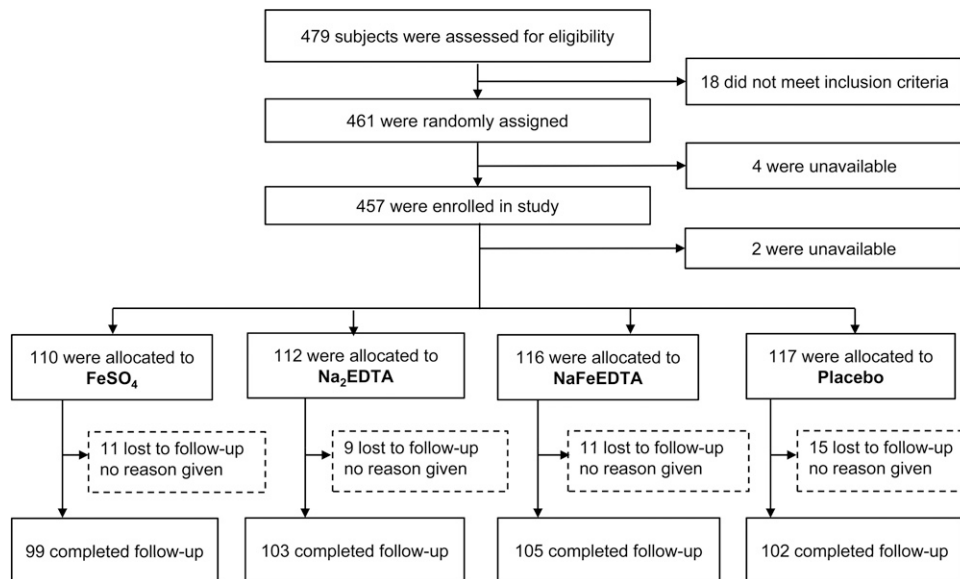
All of the biscuits (mainly composed of wheat flour, sugar, palm oil, and almonds) were identical in recipe, with the exception of the fortificant. We obtained the fortificants from Paul Lohmann GmbH, and Hug AG produced the biscuits. The iron content of the biscuits was measured by using atomic absorption spectrometry, and EDTA content was calculated from measured iron content in the fortified biscuit, taking into account the amount of native iron in the control (nonfortified) biscuit. Subjects, investigators, and sponsors were masked to group assignment.

### Procedures and outcomes

The intervention lasted 28 wk, and we administered 2 or 3 biscuits/d for 6 d/wk, depending on the child's body weight, to stay within the acceptable daily intake of EDTA ( $1.9 \text{ mg EDTA} \cdot \text{kg body weight}^{-1} \cdot \text{d}^{-1}$ ) (4, 21). Trained fieldworkers directly supervised biscuit consumption and recorded compliance daily.

The primary outcome measure of this study was BPb concentration at 28 wk of the intervention. Secondary outcome measures were iron status indicators and cognitive test scores at 28 wk of the intervention. Other outcome variables included C-reactive protein (CRP),  $\alpha_1$ -acid glycoprotein (AGP), and the prevalence of anemia, ID, and IDA, as well as the prevalence of elevated BPb concentrations.

At baseline, endpoint, and at 2 intermediate time points on the basis of a sparse random serial sampling protocol (22) (only



**FIGURE 1** Trial profile.  $\text{FeSO}_4$ , ferrous sulfate;  $\text{Na}_2\text{EDTA}$ , sodium EDTA; NaFeEDTA, ferric sodium EDTA.

baseline and endpoint results are presented), we measured height and weight and collected a 10-mL venous blood sample into EDTA-coated trace element-free tubes (Becton Dickinson) for the analysis of BPb, iron status indicators, and inflammatory markers. We analyzed hemoglobin and mean corpuscular volume by using a Sysmex XT-2000i Automated Hematology Analyzer and the corresponding standard material (L1–L3: QC-21810810 to QC-21810812). BPb was measured in baseline and endpoint whole-blood samples at the Karolinska Institute, Sweden, by using inductively coupled plasma mass spectrometry (Agilent 7700X; Agilent Technologies), as previously described (23). Analysis of Seronorm blood reference samples [reference values (acceptance range): 1.02  $\mu\text{g Pb/dL}$  (0.60–1.44  $\mu\text{g Pb/dL}$ ) and 31.0  $\mu\text{g Pb/dL}$  (18.6–43.4  $\mu\text{g Pb/dL}$ )] showed  $0.97 \pm 0.04 \mu\text{g Pb/dL}$  and  $32.1 \pm 0.76 \mu\text{g Pb/dL}$ . The limit of detection (3 times the SD of the element concentration in the calibration blank) was 0.002  $\mu\text{g Pb/dL}$ , and no samples had a BPb concentration below this level. Elevated BPb concentrations were defined as BPb  $>3 \mu\text{g/dL}$  and  $>5 \mu\text{g/dL}$  to provide categorical variables of low-level lead exposure and  $>10 \mu\text{g/dL}$  based on older reference values (24). Zinc protoporphyrin (ZnPP) was analyzed on washed erythrocytes by using a hematofluorometer as described previously (25); values  $\geq 70 \mu\text{mol/heme}$  may indicate iron-deficient erythropoiesis, but ZnPP can also be elevated by increased body lead (26). We measured serum concentrations of ferritin [serum ferritin (SF)], transferrin receptor (TfR), CRP, and AGP by using ELISA (27). We used SF  $<12 \mu\text{g/L}$  for children  $<5$  y of age, SF  $<15 \mu\text{g/L}$  for children  $\geq 5$  y (28), or TfR  $>8.3 \text{ mg/L}$  to define ID. To adjust for the elevating effect of inflammation on SF, we used correction factors suggested by Thurnham et al. (29, 30) to adjust SF concentrations by the participant's inflammation status defined as follows: no inflammation (CRP  $\leq 5 \text{ mg/L}$ , AGP  $\leq 1 \text{ g/L}$ ), incubation (CRP  $>5 \text{ mg/L}$ , AGP  $\leq 1 \text{ g/L}$ ), early convalescence (CRP  $>5 \text{ mg/L}$ , AGP  $>1 \text{ g/L}$ ), and late convalescence (CRP  $\leq 5 \text{ mg/L}$ , AGP  $>1 \text{ g/L}$ ). We defined anemia by using age-specific hemoglobin cutoffs of  $<11.0 \text{ g/dL}$ ,  $<11.5 \text{ g/dL}$ , and  $<12.0 \text{ g/dL}$  for children aged  $<5$  y, 5–11 y, and 12 y, respectively (31). We used WHO Anthro (version 3.2.2, 2011) and WHO Anthro Plus (version 1.0.3, 2010) to calculate anthropometric scores (32).

At baseline and endpoint, we performed cognitive tests, which have been previously validated in children of this age (25), in the local Arabic or Berber dialect. We administered 4 subtests from the Kaufman Assessment Battery for Children, Second Edition (KABC-II): the Atlantis (working memory) and Atlantis delayed (long-term memory and retrieval) tests from the learning scale, the hand movement test from the sequential processing scale (short-term memory, manual activity), and the Triangles test from the simultaneous processing scale (visuospatial cognition). We also administered the Hopkins Verbal Learning Test (HVLT) (33, 34). The testing procedures are described in detail elsewhere (25, 35–37).

### Statistical analysis

The sample size was estimated on the basis of the predicted improvement in BPb concentrations after iron fortification (38). The sample size calculations indicated that 85 subjects/group would be needed to detect a mean  $\pm$  SD difference in BPb of  $4.1 \pm 8.2 \mu\text{g/dL}$  over the 28 wk (38), with an  $\alpha$  of 0.05 and

a  $\beta$  of 0.20. Anticipating a drop-out rate of 10%, we planned to enroll  $\geq 95$  children/group.

For statistical analysis, we used IBM SPSS Statistics (version 22.0.0). We checked data for normal distribution (by using Q-Q plots and Shapiro-Wilk tests) and log-transformed skewed data before analysis. Parametric data are presented as means  $\pm$  SDs and nonparametric data as covariate-adjusted geometric means with 95% CIs. The latter were calculated from the estimated marginal means obtained by running separate 1-factor (treatment group as the factor) ANCOVA on all outcome variables (baseline and endpoint), including respective baseline values (only for endpoint variables), age, sex, site, and grade as covariates.

Data analysis was performed according to intention-to-treat without the removal of outliers and by using multiple imputations to treat dropouts (Figure 1) and missing data ( $<23\%$ ,  $\leq 20\%$ , and  $<15\%$  missing data for BPb, iron status, and cognitive measures, respectively; e.g., due to refusal to draw blood, assay fails, or inadequate blood volumes to perform analysis), under the assumption that data were missing at random. Five imputations were produced by using an iterative Markov chain Monte Carlo method and a linear regression model that included the treatment groups (iron and EDTA), a treatment-interaction term (iron  $\times$  EDTA), the respective baseline or endpoint variable, grade, sex, age, and site as independent predictors. For skewed variables, multiple imputations were produced on transformed data.

For each continuous outcome variable, we determined the estimated intervention effects ( $\beta$  values and 95% CIs) of iron (iron compared with no iron) and EDTA (EDTA compared with no EDTA), as well as their interaction (iron  $\times$  EDTA), by performing a 2-factor ANCOVA on the endpoint variable (dependent variable) with the use of the respective baseline values, age, sex, site, and grade as covariates. Data were log-transformed for the ANCOVA analysis, and the intervention effects ( $\beta$  values and 95% CIs) are reported on the log scale. In the presence of a significant interaction on the endpoint variable, between-group comparisons were performed by using 1-factor ANCOVA with Bonferroni adjustments and controlled for respective baseline values, age, sex, site, and grade. To test whether the difference in iron dose provided to children above and below 30 kg body weight affected any of the outcomes, we performed separate 3-factor ANCOVA to determine whether there were any interactions of body weight ( $<30$  kg compared with  $\geq 30$  kg body weight) with iron and/or EDTA treatment.

For each dichotomous outcome variable, we determined ORs and 95% CIs for each factor (iron and EDTA), as well as their interactions, by using 2-factor binary logistic regression analysis on the endpoint variable, with respective baseline values, sex, age, grade, and site controlled for.  $P < 0.05$  was considered significant.

## RESULTS

### Study participants

Between 18 September 2011 and 15 January 2012, a total of 457 participants were enrolled in the trial (Figure 1). No serious adverse events were reported during the trial.

The mean  $\pm$  SD iron content of both the NaFeEDTA and FeSO<sub>4</sub> biscuits was  $3.6 \pm 0.5 \text{ mg}$ . The NaFeEDTA and Na<sub>2</sub>EDTA biscuits both contained  $18.6 \pm 0.5 \text{ mg EDTA}$ . Children with

a body weight <30 kg ( $n = 390$ ) consumed 2 biscuits/d, which provided  $7.2 \pm 0.9$  mg Fe and  $37.1 \pm 0.9$  mg EDTA; children  $\geq 30$  kg ( $n = 77$ ) consumed 3 biscuits/d, which provided  $10.8 \pm 1.4$  mg Fe and  $55.7 \pm 1.4$  mg EDTA. The difference in iron dose provided to children with a body weight <30 kg and  $\geq 30$  kg did not affect any of the outcome variables, because we found no significant interactions of body weight (<30 kg compared with  $\geq 30$  kg) with iron and/or EDTA (data not shown). Overall, children in the 2 iron groups consumed  $7.8 \pm 1.0$  mg Fe/intervention day, and children in the 2 EDTA groups consumed  $40.1 \pm 1.0$  mg EDTA/d. Compliance was high (82.2%) and did not significantly differ between groups.

Overall baseline prevalence of anemia (defined by hemoglobin) was 21%, and overall prevalences of ID defined by SF, TfR, or ZnPP were 32%, 7%, or 34%, respectively (data not shown). Baseline characteristics of the study participants in each of the 4 groups are shown in **Table 1**.

BPb concentrations at baseline and after 28 wk of intervention are shown in **Table 2**. At baseline, the overall adjusted geometric mean BPb concentration was  $4.3 \mu\text{g/dL}$  (95% CI: 4.2,  $4.3 \mu\text{g/dL}$ ), and 2%, 29%, and 83% of children had BPb concentrations >10, >5, and >3  $\mu\text{g/dL}$ , respectively. We found a significant effect of iron ( $P = 0.009$ ) and EDTA ( $P = 0.012$ ) for reduced BPb concentrations at endpoint, but no iron  $\times$  EDTA interaction ( $P = 0.606$ ). Furthermore, children who received iron had significantly lower odds (0.48; 95% CI: 0.25, 0.90) of having BPb concentrations >3  $\mu\text{g/dL}$  than did those who did not receive iron ( $P = 0.023$ ).

Red blood cell indexes and indicators of iron and inflammatory status at baseline and after 28 wk of the intervention are shown in **Table 3**. We found significant effects of iron fortification on all red blood cell indexes and iron status indicators (all  $P < 0.05$ ). Furthermore, children who received iron had significantly lower odds of being iron-deficient (on the basis of TfR or SF), anemic, or iron-deficient anemic at endpoint (iron-deficient:  $P < 0.001$ ; anemic:  $P = 0.002$ ; iron-deficient anemic:  $P = 0.003$ ). We also found a significant effect of EDTA fortification for higher AGP ( $P = 0.019$ ) concentrations at endpoint. The children who re-

ceived EDTA-fortified biscuits had a 2.5-fold higher odds of having elevated AGP concentrations at endpoint.

The KABC-II subtest and HVLTL scores at baseline and endpoint are shown in **Table 4**. We found no significant effects of iron or EDTA fortification and no iron  $\times$  EDTA interactions on any of the KABC-II or HVLTL test scores.

## DISCUSSION

The main findings of this trial are that 1) iron and EDTA fortification additively reduce BPb concentrations in lead-exposed children and 2) iron fortification reduces the risk of having elevated BPb concentrations (>3  $\mu\text{g/dL}$ ) and improves iron status. Our results indicate that iron and EDTA act independently and in an additive manner to reduce BPb, and this suggests that they reduce BPb through independent mechanisms. A likely mechanism for reduction in BPb by iron is that the improvements in iron status reduced gastrointestinal lead absorption. Iron-deficient rats absorb a greater fraction of ingested lead (39), but human studies have produced mixed results (8, 9). Improved iron status downregulates the expression of DMT1 (40) and DMT1 may transport lead, although other routes of lead absorption may be more important (10, 11, 41). In contrast, a likely mechanism for the reduction in BPb by EDTA is chelation of lead by EDTA. Because <5% of orally administered EDTA is absorbed (20, 42), the main chelating effect of EDTA in this study was likely to take place in the gut lumen. In radiotracer studies in humans, oral EDTA reduced the absorption of gastrointestinal lead (9). In 23 adults who ingested 55.8  $\mu\text{g}$   $^{203}\text{Pb}$  without and with 1.02 mg oral  $\text{Na}_2\text{EDTA}$ , body lead retention was  $\sim 57\%$  and 8%, respectively (19). However, in our study, it is also possible that absorbed EDTA chelated BPb; the total ingested dose of EDTA over 7 mo was  $\sim 6.5$  g; if this was absorbed at 3–5%,  $\sim 200$ –300 mg absorbed EDTA was available to chelate BPb during the study.

Previous uncontrolled studies of iron supplementation or fortification to reduce BPb have produced conflicting results (43–46). In a controlled study in Mexican children ( $n = 602$ ), 6 mo of oral iron supplementation (30 mg/d as ferrous fumarate) did not reduce

**TABLE 1**  
Baseline characteristics of the intention-to-treat population<sup>1</sup>

	FeSO <sub>4</sub> ( $n = 110$ )	Na <sub>2</sub> EDTA ( $n = 112$ )	NaFeEDTA ( $n = 116$ )	Placebo ( $n = 117$ )
Female sex, $n$ (%)	57 (52)	66 (59)	53 (46)	66 (56)
Age, y	$7.5 \pm 2.9^2$	$7.7 \pm 2.9$	$7.2 \pm 2.7$	$7.1 \pm 2.7$
Height, cm	$116.3 \pm 16.7$	$118.6 \pm 16.5$	$115.7 \pm 14.8$	$115.8 \pm 16.4$
Weight, kg	$21.0 (16.0\text{--}25.0)^3$	$22.0 (16.0\text{--}27.0)$	$20.0 (17.8\text{--}25.0)$	$20.0 (16.0\text{--}25.4)$
Anthropometric indexes, $n$ (%)				
Stunting (HAZ < -2 SDs)	10 (9.1)	13 (11.6)	11 (9.5)	9 (7.7)
Underweight (WAZ < -2 SDs)	10 (9.1)	11 (9.8)	5 (4.3)	7 (6.0)
Overweight (BAZ > 1 SD and < 2 SDs)	12 (10.9)	8 (7.1)	14 (12.1)	12 (10.3)
Obese (BAZ $\geq 2$ SDs)	6 (5.5)	2 (1.8)	3 (2.6)	1 (0.9)
Inflammation status, $n$ (%)				
No inflammation	68 (62)	67 (60)	67 (58)	68 (58)
Inflammation <sup>4</sup>	42 (38)	44 (40)	49 (42)	51 (42)

<sup>1</sup> BAZ, BMI-for-age  $z$  score; FeSO<sub>4</sub>, ferrous sulfate; HAZ, height-for-age  $z$  score; Na<sub>2</sub>EDTA, sodium EDTA; NaFeEDTA, ferric sodium EDTA; WAZ, weight-for-age  $z$  score.

<sup>2</sup> Mean  $\pm$  SD (all such values).

<sup>3</sup> Median; IQR in parentheses (all such values).

<sup>4</sup> Inflammation defined as CRP  $\geq 5$  mg/L or AGP  $\leq 1$  g/L.

**TABLE 2**  
Effects of providing children with fortified biscuits containing iron and EDTA, alone and in combination, for 28 wk on blood lead concentrations<sup>1</sup>

	Estimated intervention effect <sup>2</sup>						
	FeSO <sub>4</sub> (n = 110)	Na <sub>2</sub> EDTA (n = 112)	NaFeEDTA (n = 116)	Placebo (n = 117)	Iron	EDTA	Iron × EDTA, P
Blood lead, µg/dL							
Baseline	4.10 (3.84, 4.40) <sup>3</sup>	4.53 (4.24, 4.84)	4.41 (4.09, 4.74)	4.03 (3.78, 4.31)			
Endpoint	3.30 (3.11, 3.48)	3.29 (3.09, 3.49)	2.87 (2.71, 3.04)	3.66 (3.47, 3.86)	-0.046 (-0.078, -0.013)	-0.047 (-0.080, -0.013)	0.606
Prevalences							
Blood lead >10 µg/dL							
Baseline	1 (0.9) <sup>4</sup>	4 (3.6)	3 (2.6)	1 (0.9)			
Endpoint	1 (0.9)	1 (0.9)	0 (0)	0 (0)	NA	NA	NA
Blood lead >5 µg/dL							
Baseline	30 (27)	39 (35)	37 (32)	29 (25)			
Endpoint	14 (13)	19 (17)	13 (11)	19 (16)	0.695 (0.279, 1.727)	0.777 (0.280, 2.160)	0.969
Blood lead >3 µg/dL							
Baseline	93 (85)	100 (89)	100 (86)	93 (80)			
Endpoint	59 (54)	70 (63)	57 (49)	77 (66)	0.475 (0.250, 0.902)	0.675 (0.355, 1.285)	0.921

<sup>1</sup> FeSO<sub>4</sub>, ferrous sulfate; NA, not applicable; Na<sub>2</sub>EDTA, sodium EDTA; NaFeEDTA, ferric sodium EDTA.

<sup>2</sup> Intervention effects [β values (95% CIs)] were estimated for continuous outcome variables by 2-factor ANCOVA adjusting for respective baseline values, age, sex, grade, and site. Values were log-transformed to perform ANCOVA, and the reported intervention effects [βs (95% CIs)] are reported as log-transformed data. ORs (95% CIs) were estimated for dichotomous outcome variables by using 2-factor binary logistic regression analysis adjusting for respective baseline values, sex, age, grade, and site.

<sup>3</sup> Covariate-adjusted geometric means (95% CIs), calculated from the estimated marginal means obtained by running separate 1-factor (treatment group as factor) ANCOVA on all outcome variables (baseline and endpoint), including respective baseline values (only for endpoint variables), age, sex, site, and grade as covariates (all such values). Data were log-transformed to perform ANCOVA.

<sup>4</sup> n; percentage in parentheses (all such values).

BPb compared with placebo; BPb was determined by atomic absorption spectrophotometry (15). In a controlled trial in Indian schoolchildren (n = 186), subjects who received an iron-fortified rice meal (15 mg Fe/d as ferric pyrophosphate) for 16 wk had lower BPb concentrations at endpoint than did the control group, but baseline BPb was not measured; BPb was measured by anodic stripping voltammetry (38). Differences in study design, such as subject age, iron dose and duration, and analytic method for the determination of BPb, make comparisons between studies difficult.

Our findings on the effects of iron and EDTA fortification on BPb concentrations are generalizable to other populations of lead-exposed children. First, we provided an iron dose (~8 mg/d) comparable to that typically delivered to children by iron-fortification programs (47). Second, an advantage to our study population was that our subjects had only modest elevations in BPb; these are concentrations that are common in children in many low- and high-income countries (3). Finally, we included both preschool- and school-aged children (3–14 y); this may be relevant, because current studies suggest that BPb concentrations at 5–9 y of age are strongly associated with IQ (1, 5, 48), possibly even more so than in earlier childhood (49).

Although the observed decrease in BPb concentrations was modest (35% reduction in the NaFeEDTA group of 1.54 µg/dL from a baseline concentration of ~4.4 µg/dL), cross-sectional data suggest that this change in BPb might result in a measurable increment in child IQ and school performance (1, 5). A current pooled analysis concluded that the increment in IQ points was greatest when decreasing BPb from 10.0 to 2.4 µg/dL (3.9 µg/dL; 95% CI: 2.4, 5.3 µg/dL) than with comparable decreases at higher BPb concentrations (1). Our observed reductions in BPb by the provision of iron (-0.8 µg/dL) and EDTA (-0.8 µg/dL) are within the benchmark dose of 0.1–1.0 µg/dL estimated to lead to a loss of 1 IQ point. This is in agreement with a recent large study (n = 58,650) in US children, which observed a nonlinear negative effect of BPb on reading and math skills, with a steeper failure rate for reading at BPb concentrations <5 µg/dL (5). For children with BPb concentrations <10 µg/dL, a 1-µg/dL difference in BPb was associated with an increased RR for both reading and math failure of 1.06 (95% CI: 1.05, 1.07) (5). However, we did not find an effect of iron or EDTA on KABC-II and HVLT test scores. This may have been due to the short duration of the intervention, the choice of cognitive tests, and/or a β error due to small sample size. Some experts have argued that the cognitive deficits of lead poisoning are irreversible (50), and it is unclear whether an improvement in BPb with iron and/or EDTA fortification can reverse early deficits induced by lead exposure. To our knowledge, this is the first randomized controlled intervention trial evaluating the cognitive effects of improved low-lead exposure in children. Furthermore, it is not known whether the neurodevelopmental effects of lead may be more severe when ID is also present (51), but most of the evidence suggests that their effects are independent of one another (52). Ruff et al. (43) provided parenteral EDTA treatment and oral iron supplements, a combination of both, or neither to 1- to 7-y-old children (n = 154) and measured cognitive outcomes after 6 mo of treatment. A reduction in BPb was associated with higher cognitive scores, but there was no interaction with an increase in iron status. Our study period was only 28 wk; a sustained NaFeEDTA fortification program throughout childhood would likely have a greater impact.

**TABLE 3**

Effects of providing children with fortified biscuits containing iron and EDTA, alone and in combination, for 28 wk on biochemical indicators of iron and inflammatory status<sup>1</sup>

	FeSO <sub>4</sub> (n = 110)	Na <sub>2</sub> EDTA (n = 112)	NaFeEDTA (n = 116)	Placebo (n = 117)	Estimated intervention effects <sup>2</sup>		
					Iron	EDTA	Iron × EDTA, P
Hemoglobin, g/dL							0.309
Baseline	11.9 (11.7, 12.2) <sup>3</sup>	11.8 (11.5, 12.1)	12.0 (11.7, 12.2)	12.1 (11.8, 12.3)			
Endpoint	12.3 (12.2, 12.5)	12.0 (11.8, 12.2)	12.2 (12.1, 12.4)	11.9 (11.7, 12.1)	0.02 (0.01, 0.02)	0.00 (−0.01, 0.01)	
SF, μg/L							0.156
Baseline	16.8 (14.7, 19.2)	16.2 (14.2, 18.5)	17.2 (15.1, 19.6)	17.3 (15.1, 19.7)			
Endpoint	34.5 (31.8, 37.4)	20.4 (18.3, 22.6)	32.6 (29.9, 35.6)	19.2 (17.5, 21.0)	0.25 (0.20, 0.31)	0.03 (−0.02, 0.07)	
Serum TfR, mg/L							0.505
Baseline	5.3 (5.0, 5.7)	5.3 (5.0, 5.7)	5.3 (5.0, 5.7)	5.1 (4.8, 5.4)			
Endpoint	2.9 (2.8, 3.1)	3.1 (3.0, 3.3)	2.8 (2.7, 2.9)	3.1 (3.0, 3.3)	−0.03 (−0.06, −0.01)	−0.01 (−0.03, 0.02)	
Body iron, mg/kg							0.164
Baseline	2.0 (1.1, 3.0)	1.7 (0.8, 2.7)	2.2 (1.3, 3.1)	2.4 (1.5, 3.4)			
Endpoint	7.2 (6.8, 7.7)	5.0 (4.6, 5.5)	7.1 (6.7, 7.6)	4.7 (4.3, 5.0)	0.06 (0.05, 0.07)	0.01 (−0.00, 0.02)	
ZnPP, μmol/mol heme							0.984
Baseline	66.5 (61.1, 72.4)	69.0 (63.4, 75.2)	67.6 (62.1, 73.6)	66.1 (60.7, 71.9)			
Endpoint	75.2 (71.8, 78.5)	87.5 (83.6, 91.8)	75.2 (71.8, 78.7)	87.7 (83.4, 92.0)	−0.07 (−0.10, −0.04)	0.00 (−0.03, 0.03)	
Mean corpuscular volume, fL							0.734
Baseline	78.5 (77.4, 79.6)	78.7 (77.6, 79.8)	78.2 (77.1, 79.3)	79.1 (78.0, 80.4)			
Endpoint	81.1 (80.5, 81.7)	79.8 (79.3, 80.4)	81.1 (80.5, 81.7)	79.6 (79.1, 80.2)	0.01 (0.00, 0.11)	0.00 (−0.00, 0.01)	
CRP, mg/L							0.056
Baseline	0.63 (0.45, 0.82)	0.65 (0.47, 0.85)	0.66 (0.49, 0.86)	0.67 (0.50, 0.87)			
Endpoint	0.93 (0.67, 1.22)	1.23 (0.94, 1.58)	0.80 (0.52, 1.12)	0.81 (0.56, 1.10)	0.03 (−0.05, 0.11)	0.09 (0.01, 0.18)	
AGP, g/L							0.044
Baseline	0.81 (0.78, 0.85)	0.80 (0.76, 0.83)	0.82 (0.79, 0.86)	0.80 (0.76, 0.83)			
Endpoint	0.80 (0.76, 0.84)	0.84 (0.79, 0.89)	0.78 (0.74, 0.83)	0.77 (0.73, 0.81)	0.02 (−0.02, 0.05)	0.04 (0.01, 0.07)	
Anemia <sup>4</sup>							0.236
Baseline	23 (21) <sup>5</sup>	26 (23)	25 (22)	22 (19)	0.21 (0.08, 0.57)	0.69 (0.30, 1.60)	
Endpoint	9 (8)	39 (35)	14 (12)	28 (24)			
ID <sup>6</sup>							0.982
Baseline	38 (35)	32 (29)	42 (36)	35 (30)	0.07 (0.02, 0.24)	1.05 (0.46, 2.41)	
Endpoint	5 (5)	30 (27)	6 (5)	31 (27)			
IDA <sup>7</sup>							0.592
Baseline	14 (13)	15 (13)	18 (16)	12 (10)	0.06 (0.01, 0.38)	0.83 (0.26, 2.60)	
Endpoint	3 (3)	16 (14)	2 (2)	15 (13)			
Elevated CRP (>5 mg/L)							0.064
Baseline	40 (36)	42 (38)	45 (39)	47 (40)	1.36 (0.47, 3.90)	2.11 (0.79, 5.64)	
Endpoint	10 (9)	15 (13)	5 (4)	9 (8)			
Elevated AGP (>1 g/L)							0.356
Baseline	16 (15)	17 (15)	23 (20)	19 (16%)	0.98 (0.43, 2.20)	2.43 (1.16, 5.10)	
Endpoint	17 (16)	34 (30)	26 (22)	19 (16%)			

<sup>1</sup> AGP, α<sub>1</sub>-acid glycoprotein; CRP, C-reactive protein; FeSO<sub>4</sub>, ferrous sulfate; ID, iron deficiency; IDA, iron deficiency anemia; Na<sub>2</sub>EDTA, sodium EDTA; NaFeEDTA, ferric sodium EDTA; SF, serum ferritin; TfR, transferrin receptor; ZnPP, zinc protoporphyrin.

<sup>2</sup> Intervention effects [β values (95% CIs)] were estimated for continuous outcome variables by 2-factor ANCOVA adjusting for respective baseline values, age, sex, grade, and site. Values were log-transformed to perform ANCOVA, and the reported intervention effects [βs (95% CIs)] are reported as log-transformed data. ORs (95% CIs) were estimated for dichotomous outcome variables by using 2-factor binary logistic regression analysis adjusting for respective baseline values, sex, age, grade, and site.

<sup>3</sup> Covariate-adjusted geometric means (95% CIs), calculated from the estimated marginal means obtained by running separate 1-factor (treatment group as factor) ANCOVA on all outcome variables (baseline and endpoint), including respective baseline values (only for endpoint variables), age, sex, site, and grade as covariates (all such values). Data were log-transformed to perform ANCOVA.

<sup>4</sup> Anemia was defined according to the age specific cutoffs for hemoglobin of <11.0, <115, and <120 g/dL for children aged <5 y, 5–11, and ≥12 y of age, respectively.

<sup>5</sup> n; percentage in parentheses (all such values).

<sup>6</sup> ID was defined as SF <12 μg/L for children <5 y of age, SF <15 μg/L for children ≥5 y (28), or TfR >8.3 mg/L. We adjusted SF for inflammation by using the correction factors suggested by Thurnham et al. (29, 30).

<sup>7</sup> IDA was defined as being iron-deficient and anemic.

**TABLE 4**  
Effects of providing children with fortified biscuits containing iron and EDTA, alone and in combination, for 28 wk on cognition<sup>1</sup>

	Estimated intervention effects <sup>2</sup>						
	FeSO <sub>4</sub> (n = 110)	Na <sub>2</sub> EDTA (n = 112)	NaFeEDTA (n = 116)	Placebo (n = 117)	Iron	EDTA	Iron × EDTA, P
<b>KABC-II</b>							
Atlantis (working memory)							
Baseline	31.5 (28.6, 34.7) <sup>3</sup>	30.9 (28.1, 34.0)	28.4 (25.4, 31.7)	28.5 (25.8, 31.4)			0.618
Endpoint	35.2 (32.1, 38.6)	37.1 (34.1, 40.4)	36.4 (33.5, 39.6)	37.5 (34.1, 41.2)	-0.03 (-0.09, 0.03)	-0.01 (-0.06, 0.05)	0.912
Atlantis delayed (long-term memory)							
Baseline	5.4 (4.6, 6.2)	5.5 (4.8, 6.2)	5.5 (4.8, 6.2)	4.9 (4.2, 5.6)			
Endpoint	7.6 (6.9, 8.4)	7.9 (7.1, 8.7)	7.5 (6.7, 8.3)	7.9 (7.1, 8.8)	-0.02 (-0.08, 0.04)	-0.002 (-0.06, 0.06)	0.479
Hand movement (short-term memory)							
Baseline	5.3 (4.6, 6.0)	5.2 (4.6, 5.8)	5.5 (4.8, 6.2)	5.0 (4.3, 5.7)			
Endpoint	5.5 (5.1, 6.1)	5.8 (5.2, 6.3)	5.8 (5.3, 6.4)	5.9 (5.3, 6.5)	-0.02 (-0.07, 0.03)	-0.01 (-0.06, 0.05)	0.372
Triangles test (visuospatial cognition)							
Baseline	9.9 (8.8, 11.1)	9.6 (8.5, 10.8)	9.0 (7.7, 10.4)	9.9 (8.3, 11.8)			
Endpoint	11.0 (10.3, 11.8)	11.8 (11.0, 12.6)	11.0 (10.3, 11.8)	10.8 (10.1, 11.5)	-0.00 (-0.04, 0.03)	0.04 (-0.002, 0.07)	0.666
<b>HVLT</b>							
Recall 1							
Baseline	3.6 (3.1, 4.1)	3.2 (2.8, 3.6)	3.3 (2.9, 3.7)	3.6 (3.2, 4.0)			
Endpoint	4.2 (3.8, 4.6)	3.9 (3.5, 4.4)	4.0 (3.6, 4.4)	4.0 (3.5, 4.5)	0.02 (-0.04, 0.08)	-0.002 (-0.06, 0.05)	0.578
Recall 2							
Baseline	5.1 (4.5, 5.7)	4.7 (4.2, 5.1)	4.5 (4.0, 5.0)	5.4 (4.9, 6.0)			
Endpoint	5.6 (5.2, 6.1)	5.7 (5.2, 6.2)	5.9 (5.4, 6.4)	5.2 (4.7, 5.8)	0.03 (-0.02, 0.08)	0.03 (-0.01, 0.08)	0.590
Recall 3							
Baseline	5.2 (4.6, 5.8)	4.8 (4.2, 5.3)	5.2 (4.6, 5.8)	6.0 (5.2, 6.8)			
Endpoint	6.1 (5.6, 6.6)	6.3 (5.8, 6.9)	6.4 (5.8, 7.0)	6.1 (5.6, 6.7)	0.00 (-0.05, 0.05)	-0.01 (-0.03, 0.06)	0.650
Total scores of recalls							
Baseline	10.7 (9.2, 12.5)	11.2 (9.6, 13.1)	11.5 (9.8, 13.4)	11.9 (10.2, 13.9)			
Endpoint	12.3 (10.3, 14.6)	11.3 (9.5, 13.5)	12.1 (10.2, 14.3)	10.6 (8.9, 12.6)	0.06 (-0.04, 0.16)	-0.03 (-0.07, 0.03)	0.723
Recognition							
Baseline	10.2 (9.6, 10.8)	10.5 (9.8, 11.1)	10.1 (9.6, 10.7)	10.1 (9.5, 10.7)			
Endpoint	10.4 (9.9, 10.9)	10.6 (10.1, 11.2)	10.4 (9.9, 10.8)	10.8 (10.3, 11.3)	-0.02 (-0.04, 0.01)	-0.01 (-0.03, 0.02)	0.333
Discrimination index							
Baseline	6.5 (5.7, 7.4)	5.8 (5.1, 6.7)	5.7 (4.9, 6.5)	6.1 (5.3, 6.9)			
Endpoint	6.1 (5.3, 7.0)	6.1 (5.3, 7.0)	6.6 (5.8, 7.5)	6.5 (5.7, 7.4)	-0.02 (-0.07, 0.04)	-0.02 (-0.07, -0.04)	

<sup>1</sup> FeSO<sub>4</sub>, ferrous sulfate; HVLT, Hopkins Verbal Learning Test; KABC-II, Kaufman Assessment Battery for Children, Second Edition; Na<sub>2</sub>EDTA, sodium EDTA; NaFeEDTA, ferric sodium EDTA.  
<sup>2</sup> Intervention effects [ $\beta$  values (95% CIs)] were estimated for continuous outcome variables by 2-factor ANCOVA adjusting for respective baseline values, age, sex, grade, and site. Values were log-transformed to perform ANCOVA, and the reported intervention effects [ $\beta$ s (95% CIs)] are reported as log-transformed data.  
<sup>3</sup> Covariate-adjusted geometric means (95% CIs), calculated from the estimated marginal means obtained by running separate 1-factor (treatment group as factor) ANCOVA on all outcome variables (baseline and endpoint), including respective baseline values (only for endpoint variables), age, sex, site, and grade as covariates (all such values). Data were log-transformed to perform ANCOVA.

Our study has limitations. We did not measure fecal or renal excretion of EDTA-chelated lead because of the difficulties of obtaining multiple fecal and 24-h urine collections from children in our study setting. The fortificants were provided in a noninhibitory food matrix; the effects of iron fortification of inhibitory foods on BPb may differ because of the presence of native chelators, such as phytic acid and polyphenols. The actual change in BPb was lower than that used for the power calculation; however, we were able to discriminate this smaller change because the variance in BPb was much lower than in the Indian study used to estimate the original sample size; we also enrolled ~20% more subjects than required from the sample size calculation. Although ZnPP was significantly lower in the iron groups at endpoint than in the Na<sub>2</sub>EDTA or placebo groups, ZnPP remained elevated in all 4 groups. This suggests the persistence of iron-deficient erythropoiesis despite a clear reduction in ID on the basis of SF and TfR in the 2 groups who received iron fortification. Lead inhibits the activity of the ferrochelatase that catalyzes insertion of iron into the protoporphyrin ring to form heme; in severe lead poisoning (BPb  $\geq 30$   $\mu\text{g/dL}$ ), this can increase ZnPP (53), but the BPb elevations in our subjects were much lower, suggesting that the persisting elevations of ZnPP in our study were unlikely due to lead exposure.

In conclusion, our findings suggest that, in lead-exposed populations, NaFeEDTA should be the iron fortificant of choice, because it reduces both ID and BPb. Because the cognitive deficits of lead poisoning may be irreversible (50), primary prevention is critical. Iron-fortification programs are rapidly expanding worldwide, and the urban poor of Latin America, Africa, and Asia are major target groups. An advantage of iron fortification of staple foods as a BPb mitigation strategy is that it may reach even the poorest and most disadvantaged groups (i.e., those most likely to be lead-exposed). In addition, iron fortification can reduce the risk of IDA in women of reproductive age and pregnant women, and thereby might reduce fetal lead exposure.

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## REFERENCES

- Lanphear BP, Hornung R, Khoury J, Yolton K, Baghurst P, Bellinger DC, Canfield RL, Dietrich KN, Bornschein R, Greene T, et al. Low-level environmental lead exposure and children's intellectual function: an international pooled analysis. *Environ Health Perspect* 2005;113:894-9.
- EFSA Panel on Contaminants in the Food Chain. Scientific opinion on lead in food. Parma (Italy): European Food Safety Authority; 2013.
- Budtz-Jørgensen E, Bellinger D, Lanphear B, Grandjean P; International Pooled Lead Study Investigators. An international pooled analysis for obtaining a benchmark dose for environmental lead exposure in children. *Risk Anal* 2013;33:450-61.
- Joint FAO/WHO Expert Committee on Food Additives. Evaluation of certain food additives and contaminants. Seventy-third report of the Joint FAO/WHO Expert Committee on Food Additives. *World Health Organ Tech Rep Ser* 2011;(966):1-136.
- Evens A, Hryhorczuk D, Lanphear BP, Rankin KM, Lewis DA, Forst L, Rosenberg D. The impact of low-level lead toxicity on school performance among children in the Chicago public schools: a population-based retrospective cohort study. *Environ Health* 2015;14:21.
- Joint FAO/WHO Expert Committee on Food Additives. Joint FAO/WHO Expert Committee on Food Additives (JECFA) meeting. Fifty-third meeting. Rome (Italy): FAO; 1999. [cited 2016 Sep 21]. Available from: <ftp://ftp.fao.org/esn/jecfa/jecfa61sc.pdf>.
- American Academy of Pediatrics Committee on Environmental Health. Lead exposure in children: prevention, detection, and management. *Pediatrics* 2005;116:1036-46.
- Watson WS, Morrison J, Bethel MI, Baldwin NM, Lyon DT, Dobson H, Moore MR, Hume R. Food iron and lead absorption in humans. *Am J Clin Nutr* 1986;44:248-56.
- Flanagan PR, Chamberlain MJ, Valberg LS. The relationship between iron and lead absorption in humans. *Am J Clin Nutr* 1982;36:823-9.
- Bannon DI, Abounader R, Lees PS, Bressler JP. Effect of DMT1 knockdown on iron, cadmium, and lead uptake in Caco-2 cells. *Am J Physiol Cell Physiol* 2003;284:C44-50.
- Gunshin H, Mackenzie B, Berger UV, Gunshin Y, Romero MF, Boron WF, Nussberger S, Gollan JL, Hediger MA. Cloning and characterization of a mammalian proton-coupled metal-ion transporter. *Nature* 1997;388:482-8.
- Barton JC, Conrad ME, Nuby S, Harrison L. Effects of iron on the absorption and retention of lead. *J Lab Clin Med* 1978;92:536-47.
- Kwong WT, Friello P, Semba RD. Interactions between iron deficiency and lead poisoning: epidemiology and pathogenesis. *Sci Total Environ* 2004;330:21-37.
- Wright RO, Tsaih SW, Schwartz J, Wright RJ, Hu H. Association between iron deficiency and blood lead level in a longitudinal analysis of children followed in an urban primary care clinic. *J Pediatr* 2003;142:9-14.
- Rosado JL, Lopez P, Kordas K, Garcia-Vargas G, Ronquillo D, Alatorre J, Stoltzfus RJ. Iron and/or zinc supplementation did not reduce blood lead concentrations in children in a randomized, placebo-controlled trial. *J Nutr* 2006;136:2378-83.
- Centers for Disease Control and Prevention. Managing elevated blood lead levels among young children: recommendations from the Advisory Committee on Childhood Lead Poisoning Prevention. Atlanta (GA): CDC; 2002.
- Bothwell TH, MacPhail AP. The potential role of NaFeEDTA as an iron fortificant. *Int J Vitam Nutr Res* 2004;74:421-34.
- Bradberry S, Vale A. A comparison of sodium calcium edetate (edetate calcium disodium) and succimer (DMSA) in the treatment of inorganic lead poisoning. *Clin Toxicol (Phila)* 2009;47:841-58.
- James HM, Hilburn ME, Blair JA. Effects of meals and meal times on uptake of lead from the gastrointestinal tract in humans. *Hum Toxicol* 1985;4:401-7.
- Foreman H, Trujillo TT. The metabolism of C14 labeled ethylenediaminetetraacetic acid in human beings. *J Lab Clin Med* 1954;43:566-71.
- EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS). Scientific opinion on the use of ferric sodium EDTA as a source of iron added for nutritional purposes to foods for the general population (including food supplements) and to foods for particular nutritional uses. *EFSA J* 2010;8(1):1414-46.
- Andersson M, Theis W, Zimmermann MB, Foman JT, Jakel M, Duchateau GS, Frenken LG, Hurrell RF. Random serial sampling to evaluate efficacy of iron fortification: a randomized controlled trial of margarine fortification with ferric pyrophosphate or sodium iron edetate. *Am J Clin Nutr* 2010;92:1094-104.
- Lu Y, Kippler M, Harari F, Grandner M, Palm B, Nordqvist H, Vahter M. Alkali dilution of blood samples for high throughput ICP-MS analysis-comparison with acid digestion. *Clin Biochem* 2015;48:140-7.
- Centers for Disease Control. CDC response to Advisory Committee on Childhood Lead Poisoning Prevention recommendations. Low level lead exposure harms children: a renewed call for primary prevention. Atlanta (GA): CDC; 2012 (updated 7 June). (updated 2012 Jun 7). [cited 2016 Sep 21]. Available from: [http://www.cdc.gov/nceh/lead/acclpp/final\\_document\\_030712.pdf](http://www.cdc.gov/nceh/lead/acclpp/final_document_030712.pdf).
- Baumgartner J, Smuts CM, Malan L, Kvalsvig J, van Stuijvenberg ME, Hurrell RF, Zimmermann MB. Effects of iron and n-3 fatty acid supplementation, alone and in combination, on cognition in school children: a randomized, double-blind, placebo-controlled intervention in South Africa. *Am J Clin Nutr* 2012;96:1327-38.



26. Eisinger J, Blumberg WE, Fischbein A, Lilis R, Selikoff IJ. Zinc protoporphyrin in blood as a biological indicator of chronic lead intoxication. *J Environ Pathol Toxicol* 1978;1:897–910.
27. Grantham-McGregor S. Early child development in developing countries. *Lancet* 2007;369:824.
28. World Health Organization. Serum ferritin concentrations for the assessment of iron status and iron deficiency in populations. Geneva (Switzerland): World Health Organization; 2011.
29. Thurnham DI, McCabe LD, Haldar S, Wieringa FT, Northrop-Clewes CA, McCabe GP. Adjusting plasma ferritin concentrations to remove the effects of subclinical inflammation in the assessment of iron deficiency: a meta-analysis. *Am J Clin Nutr* 2010;92:546–55.
30. Thurnham DI, Northrop-Clewes CA, Knowles J. The use of adjustment factors to address the impact of inflammation on vitamin a and iron status in humans. *J Nutr* 2015;145:1137S–43S.
31. World Health Organization. Haemoglobin concentrations for the diagnosis of anaemia and assessment of severity. Geneva (Switzerland): World Health Organization; 2011.
32. Baumgartner J. Interactions between iron and omega-3 fatty acids: effects of deficiency and repletion on brain monoamines and cognition [dissertation]. Zurich (Switzerland): Swiss Federal Institute of Technology; 2012.
33. Kaufman AS, Lichtenberger EO, Fletscher-Janzen E, Kaufman LN. Essentials of KABC-II assessment. 2nd ed. Hoboken (NJ): John Wiley & Sons; 2005.
34. Brandt J. The Hopkins Verbal Learning Test: development of a new memory test with six equivalent forms. *Clin Neuropsychol* 1991;5:125–42.
35. Ogunlade AO, Kruger HS, Jerling JC, Smuts CM, Covic N, Hanekom SM, Mamabolo RL, Kvalsvig J. Point-of-use micronutrient fortification: lessons learned in implementing a preschool-based pilot trial in South Africa. *Int J Food Sci Nutr* 2011;62:1–16.
36. Dalton A, Wolmarans P, Witthuhn RC, van Stuijvenberg ME, Swanevelder SA, Smuts CM. A randomised control trial in schoolchildren showed improvement in cognitive function after consuming a bread spread, containing fish flour from a marine source. *Prostaglandins Leukot Essent Fatty Acids* 2009;80:143–9.
37. Muthayya S, Eilander A, Transler C, Thomas T, van der Knaap HC, Srinivasan K, van Klinken BJ, Osendarp SJ, Kurpad AV. Effect of fortification with multiple micronutrients and n-3 fatty acids on growth and cognitive performance in Indian schoolchildren: the CHAMPION (Children's Health and Mental Performance Influenced by Optimal Nutrition) study. *Am J Clin Nutr* 2009;89:1766–75.
38. Zimmermann MB, Muthayya S, Moretti D, Kurpad A, Hurrell RF. Iron fortification reduces blood lead levels in children in Bangalore, India. *Pediatrics* 2006;117:2014–21.
39. Robertson IK, Worwood M. Lead and iron absorption from rat small intestine: the effect of dietary Fe deficiency. *Br J Nutr* 1978;40:253–60.
40. Leong WI, Bowlus CL, Tallkvist J, Lonnerdal B. Iron supplementation during infancy—effects on expression of iron transporters, iron absorption, and iron utilization in rat pups. *Am J Clin Nutr* 2003;78:1203–11.
41. Elsenhans B, Janser H, Windisch W, Schümann K. Does lead use the intestinal absorptive pathways of iron? Impact of iron status on murine <sup>210</sup>Pb and <sup>59</sup>Fe absorption in duodenum and ileum in vivo. *Toxicology* 2011;284:7–11.
42. Foreman H, Vier M, Magee M. The metabolism of C14-labeled ethylenediaminetetraacetic acid in the rat. *J Biol Chem* 1953;203:1045–53.
43. Ruff HA, Bijur PE, Markowitz M, Ma YC, Rosen JF. Declining blood lead levels and cognitive changes in moderately lead-poisoned children. *JAMA* 1993;269:1641–6.
44. Angle CRSK, McIntire MS. Lead and iron deficiencies. In: Hemphill DD, editor. Trace substances in environmental health. Columbia (MO): University of Missouri, 1975.
45. Wolf AW, Jimenez E, Lozoff B. Effects of iron therapy on infant blood lead levels. *J Pediatr* 2003;143:789–95.
46. Choi JW, Kim SK. Association between blood lead concentrations and body iron status in children. *Arch Dis Child* 2003;88:791–2.
47. Lynch SR. The impact of iron fortification on nutritional anaemia. *Best Pract Res Clin Haematol* 2005;18:333–46.
48. Chen A, Dietrich KN, Ware JH, Radcliffe J, Rogan WJ. IQ and blood lead from 2 to 7 years of age: are the effects in older children the residual of high blood lead concentrations in 2-year-olds? *Environ Health Perspect* 2005;113:597–601.
49. Hornung RW, Lanphear BP, Dietrich KN. Age of greatest susceptibility to childhood lead exposure: a new statistical approach. *Environ Health Perspect* 2009;117:1309–12.
50. Rischitelli G, Nygren P, Bougatsos C, Freeman M, Helfand M. Screening for elevated lead levels in childhood and pregnancy: an updated summary of evidence for the US Preventive Services Task Force. *Pediatrics* 2006;118:e1867–95.
51. Kordas K. Iron, lead, and children's behavior and cognition. *Annu Rev Nutr* 2010;30:123–48.
52. Wasserman G, Graziano JH, Factor-Litvak P, Popovac D, Morina N, Musabegovic A, Vrenezi N, Capuni-Paracka S, Lekic V, Preteni-Redjepi E, et al. Independent effects of lead exposure and iron deficiency anemia on developmental outcome at age 2 years. *J Pediatr* 1992;121:695–703.
53. Goering PL. Lead-protein interactions as a basis for lead toxicity. *Neurotoxicology* 1993;14:45–60.