

Respiratory fluxes and fine root responses in mature *Picea abies* trees exposed to elevated atmospheric CO₂ concentrations

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Abstract With their dominant share in global plant biomass carbon (C), forests and their responses to atmospheric CO₂ enrichment are key to the global C balance. In this free air CO₂ enrichment (FACE) study, we assessed respiratory losses from stems and soil, and fine root growth of ca. 110-year-old *Picea abies* growing in a near-natural forest in NW Switzerland. We anticipated a stimulation of all three variables in response to a ca. 150 ppm higher CO₂ concentration in the tree canopies. During the first 2.5 years of the experiment, stem CO₂ efflux (R_{stem}) remained unresponsive to CO₂ enrichment. This indicates that there is no enhancement of metabolic

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UTAS AG Büro für Landschaft, Natur und Siedlung, Brünigstrasse 64, 6074 Giswil, Switzerland activity in phloem and xylem of these mature trees. Soil CO₂ efflux (R_{soil}) beneath trees experiencing elevated CO_2 (eCO₂) showed a slight but significant reduction compared to R_{soil} under control trees. High CO2 trees did not increase their fine root biomass in ingrowth cores after 20 months under FACE relative to the fine root fractions collected in undisturbed soil. Tree growth (stem radial increment, not shown here) remained completely unchanged although earlier experiments showed largest responses (if any) during the early years after a step increase in atmospheric CO₂ concentration. The data presented here suggest C saturation of the study trees at the current close to 400 ppm CO₂ ambient concentrations. Together with the high local atmospheric N-deposition rates (ca. 20 kg N ha⁻¹ a⁻¹), our findings imply that factors other that C and N supply appear to constrain growth and metabolism of these mature P. abies trees under eCO₂.

Introduction

The rising levels of atmospheric CO_2 are potentially affecting forest biomes not only indirectly via the climatic change, but also directly via potentially enhanced CO_2 uptake by tree canopies. Higher leaflevel CO₂ uptake of forest trees in response to elevated CO_2 (eCO₂) was repeatedly reported (Bader et al. 2010; Darbah et al. 2010; Ellsworth et al. 2012). However, this additional C uptake mostly resulted in a less-than-anticipated, or no long-term increase in growth or net primary productivity in maturing trees (Körner 2006; Norby and Zak 2011; Leuzinger and Hättenschwiler 2013; Sigurdsson et al. 2013). The direct effect of CO₂ via photosynthesis might be masked by a set of counteracting biotic and abiotic effects on tree growth (Körner 2000; Leuzinger and Hättenschwiler 2013). Soil nutrient availability, stand development, and species identity are influencing the potential CO₂ fertilization effect (Hättenschwiler et al. 1997; De Graaff et al. 2006; Norby et al. 2010; Bader et al. 2013). The imbalance between increased foliar C uptake without corresponding aboveground growth response to eCO_2 might be compensated by a stimulation of fine root growth, or by increased respiratory release of CO₂ to the atmosphere.

Although total fine root mass (<2 % of total tree biomass; Körner 1994) contributes little to ecosystem biomass C-stores (<1 %, including in soil organic matter), the turnover of fine roots provides a significant source for soil humus formation. The rapid turnover of fine roots may, in fact, contribute between 20 and 50 % to annual biomass production (Jackson et al. 2009), and thus, plays a significant role in the global C cycle (Matamala et al. 2003). Some studies on young, expanding systems arrived at ca. 40 % increase in fine root biomass at eCO₂ (Curtis and Wang 1998). These results are most likely due to a faster exploration of 'empty' soil when these young trees grew toward canopy closure (unlikely a steadystate signal for mature forests). The Oak Ridge free air CO₂ enrichment (FACE) study in a plantation of Liquidambar styraciflua (10-year-old when the study was initiated in 1997) initially reported several years of increased fine root production (Norby et al. 2004), which led to increased C fluxes to the soil (Jastrow et al. 2005; Iversen et al. 2012). However, this CO₂induced belowground growth stimulation ceased completely towards the end of the 11-year enrichment period, which was explained by the higher nitrogen (N) demand for greater C uptake (Norby et al. 2010; Garten et al. 2011). Intriguingly, a study with *Pinus* taeda (Duke FACE; initiated in 1996 with then 13-year-old trees) did not reveal increased soil C accumulation (Phillips et al. 2012) despite accelerated belowground C fluxes and higher fine root production belowground (Pritchard et al. 2008; Jackson et al. 2009; Drake et al. 2011). Phillips et al. (2012) highlighted that accelerated microbial activity under eCO₂ not only enhances the mineralization of soil organic matter pools (soil priming; Jenkinson et al. 1985) but also offsets the increased input of rootderived C under eCO₂ (rhizodepositions, exudation, and allocation to mycorrhizal fungi) by enhancing the decomposition of these compounds. Similar results were recently obtained in a CO2-enriched scrub-oak community (Hungate et al. 2013). Such priming processes can release additional N, which becomes readily available for tree metabolism (increase N-cycling), and might thus slow the natural, progressive N limitation (PNL) as forests mature. Additionally, deeper soil exploration by roots under eCO₂ might further increase the availability of N (at least transitorily; Pritchard et al. 2008; Iversen et al. 2011). Stimulated microbial activity also accelerated the returns of CO_2 from the soil to the atmosphere in this pine forest (Jackson et al. 2009). CO₂-driven priming effects such as described by Phillips et al. (2012) are in accordance with results of the 8-year FACE study on mature deciduous trees growing under near-natural, but N-saturated conditions at our study site (Bader et al. 2013). Here, soil N availability (Schleppi et al. 2012), and microbial biomass increased significantly under eCO₂ (Bader and Körner 2010). However, in contrast to the P. taeda results at Duke FACE, no aboveground growth (Bader et al. 2013), no stimulation of soil CO₂ efflux (Bader and Körner 2010), and reduced fine root biomass (Bader et al. 2009) were observed in these deciduous trees despite strong photosynthetic stimulation by eCO₂ (Bader et al. 2010).

Higher respiratory CO_2 release from soils (Drake et al. 2011) would be a consequence of increased belowground C supply under eCO_2 (growth and turnover of roots, rhizodeposition, metabolic activity of roots and mycorrhizal partners), assuming that soil microbes are limited by labile C (Fierer et al. 2009). This might reverse the effect of the often-anticipated eCO_2 'fertilization' on forest ecosystems (Raich and Schlesinger 1992). Higher soil CO_2 efflux (R_{soil}) under trees exposed to eCO_2 has been reported frequently (Spinnler et al. 2002; Bernhardt et al. 2006; Comstedt et al. 2006; Pregitzer et al. 2008; Jackson et al. 2009). However, all these test systems contained young trees with expanding root spheres. Not surprisingly, these initial effects declined with time, and were not observed under mature trees in mixed forest stands (Bader and Körner 2010), or in monospecific plantations (King et al. 2004).

Stem CO₂ efflux (R_{stem}) can contribute 13–42 % to the total aboveground C budget of trees (Waring and Schlesinger 1985; Hamilton et al. 2002). R_{stem} responses to eCO_2 have mostly been reported in juvenile trees, and they vary considerably (i.e. reductions and increases; Carey et al. 1996; Janouš et al. 2000; Edwards et al. 2002; Hamilton et al. 2002; Zha et al. 2005; Acosta et al. 2010). So far, we do not see any aboveground growth stimulation following CO₂ enrichment in the tall trees examined here (T. Klein and C. Körner, unpublished), despite indications of increased leaf-level C uptake under eCO_2 (Leuzinger and Bader 2012; Bader et al., in prep.; T. Klein, pers. comm.). Therefore, any CO_2 -driven stimulation of R_{stem} would reflect higher phloem activity or maintenance respiration, or a signal resulting from enhanced CO₂ release in the rhizosphere, from where the respiratory C may find its way into R_{stem} via the xylem sap.

We used the Swiss Canopy Crane (SCC) web-FACE facility (Pepin and Körner 2002) to expose the canopies of 37-m tall, and ca. 110-year-old *P. abies* to increased levels of atmospheric CO₂. The effectiveness of CO₂ enrichment could be confirmed by C isotope signals (Mildner et al. 2014). Here we report the initial responses (i.e. the first 2.5 years of FACE) of *P. abies* to atmospheric CO₂ enrichment, with a focus on stem and soil CO₂ release, and fine root production. We hypothesized (i) a stimulation in fine root production, (ii) enhanced CO₂ efflux from soils, and (iii) greater stem CO₂ efflux under eCO₂ compared to ambient conditions.

Materials and methods

Study site and experimental setup

The experiment was established in a highly diverse, near-natural forest 12 km south-west of Basel, Switzerland (47°33'N, 7°36'E, 500 m a.s.l), dominated by ca. 100–120-year-old deciduous and coniferous trees (dominant species are i.e. *Fagus sylvatica* L., *Quercus petraea* (Matt.) Liebl., *Carpinus betulus* L., *Picea abies* (L.) Karst., *Larix decidua* Mill., *Pinus sylvestris* L., Abies alba Mill.; Fig. S4). The site has a mild temperate climate, with seasonal mean temperatures (May-September) of 14.7 °C, and ca. 800 mm a^{-1} precipitation (Bader and Körner 2010). In 2009, five 37 m tall, 110-year-old Norway spruce (P. abies) individuals were equipped with an improved web-FACE system (Pepin and Körner 2002; Körner et al. 2005; Mildner et al. 2014) using a 45 m tall canopy crane. CO_2 was released into the tree canopies through non-invasive laser-punched tubes (4 mm diameter) woven around the tree branches, allowing for computer-controlled adjustment of the CO₂ release with regard to wind direction by employing sectional control of CO₂. The web-FACE technique applied here provided the best possible means to enrich mature P. abies trees with additional CO₂ given the tree height, and the complexity of the conditions on-site. Those limitations of the web-FACE technique have been discussed in more detail elsewhere (see Pepin and Körner 2002). Our system showed good spatio-temporal performance (Leuzinger, pers. comm.). Median CO₂ concentrations were between 500 and 560 ppm in the canopies (60 sampling points per IRGA reading), with means of 541, 532, and 541 ppm for 2009, 2010, and 2011, respectively (Mildner et al. 2014). We discontinued the FACE treatment if either temperatures were below 4 °C, PPFD was <100 µmol, or wind was above 10 m s^{-1} . So, FACE was largely off during the coldest period from early November until early March (4 months). CO_2 enrichment started on 30 July 2009. Only the tree canopy between 15 and 37 m aboveground was CO₂-enriched, with no downward flow, preventing uncontrolled 'CO₂ pollution' of the understory vegetation and soil surface. Since the CO_2 employed for canopy enrichment carries a constant ¹³C isotope signal (δ^{13} C -30 ‰), it was possible to trace the carbon flows in trees and soils. Together with IRGA-based monitoring of CO₂ concentrations in the canopy air, this isotopic C tracing allowed us to assess the effectiveness of the web-FACE system, and to show that there was no contamination of the control trees by extra CO_2 (Mildner et al. 2014). The CO_2 treated trees (eCO₂-trees) formed a group, facilitating CO₂ enrichment and clear association of signals with investigated trees (Fig. S4). Five similarly tall trees under ambient CO_2 (a CO_2), away from the treated trees, served as controls (aCO₂-trees). All but one of these aCO₂-trees were outside the perimeter of the crane's jib.

Climate variables

Hourly temperatures at different heights (10 cm belowground, T_{soil}; at the soil surface in the litter layer, T_{litter}; 2 m aboveground, T_{air}) were recorded next to an eCO₂-tree using a temperature data logger (HOBO TidbiT v2; Onset Computer Corp., Bourne, MA, USA). Technical failure caused incomplete datasets that could not be complemented by statistical interpolation (see Fig. 1). Starting in August 2008 (a year before FACE), soil moisture (vol. %) at 0-10 cm depth was recorded every 6 h around the investigated trees (11 and 18 sensors arranged around the eCO₂and aCO₂-trees, respectively) using soil moisture probes, connected to a self-contained data logger (10HS and EM50, Decagon Devices Ltd., Pullman, Washington, DC, USA). Precipitation was recorded every 2 min, provided by a weather station situated 2 km from the SCC site (Flüh, Solothurn, Switzerland). Precipitation was summed on a daily basis.

Fine root sampling

On 24 March 2010, 8 months after the onset of FACE, or 4 months of effective canopy CO₂ enrichment, we took 9 soil cores (12 cm in depth \times 3.6 cm diameter) per tree in the main rooting sphere (2 m around the tree trunks) to ensure that we captured the fine roots of P. abies. The 9 soil cores were organized into three groups of three soil cores (triplets). The triplets were placed at an angle of 120° around each trunk, with 10 cm distance between each soil core in a triplet. The fine root biomass found in the soil cores was averaged per tree to account for microscale heterogeneity. We used these coring holes to install equally sized in-growth cores (cylinders made of a 2 mm stiff mesh), filled with sieved, root-free soil collected on-site. The soil in the in-growth cores was gently compacted to match the in situ bulk soil density (mass to volume ratio). The in-growth cores were extracted 20 months later (6 December 2011) by means of a knife. The soil and in-growth cores were kept frozen



Fig. 1 Seasonal variation of daily soil surface T under litter (*solid black line* in the *upper four panels*), precipitation (*vertical bars*), and soil moisture in the top 10 cm (*lower four panels*) measured either at the swiss canopy crane (SCC) site (T and soil

moisture), or taken from a nearby weather station 2 km away from the SCC site (precipitation) in the years 2008 to 2011 (*left* to *right*). Soil moisture was measured either under control *Picea abies* trees (*dashed line*), or under CO₂ treated trees (*solid line*)

at -20 °C until further analysis. The cores were defrosted in cold water for 48 h at 4 °C before processing to slow microbial degradation of fine roots. Fine roots were extracted using a sieve (1 mm mesh) and tweezers. P. abies fine roots were selected on the basis of a P. abies reference root collection. The distinct morphology of P. abies roots warranted the separation of P. abies roots from roots of other species (as later confirmed by δ^{13} C signals; Mildner et al. 2014). We did not quantify the fraction of non-P. abies fine roots at the time of harvest. However, we revisited the fine root fraction matter and re-sampled the same location, and weighed the non-P. abies fine root fraction in autumn 2014. We found that half of the fine roots were from *P*. *abies*, and the other half belonged to the surrounding trees of this semi-natural mixed forest. Fine roots were classified into three diameter classes (<0.5, 0.5-1, 1-2 mm), dried at 80 °C for 48 h, and weighed for biomass determination. No differentiation of still intact dead and live fine roots was made.

Soil respiration

We measured CO_2 release from the forest floor, hereafter referred to as soil respiration (R_{soil} ; µmol $CO_2 m^{-2} s^{-1}$), with two identical custom-made, closed, non-steady-state, non-through-flow chambers. The chambers were equipped with open path, non-dispersive infrared gas analysers, and relative humidity/T sensors (GMP343 carbon dioxide probe, HMP75 rH/T probe; Vaisala, Vantaa, Finland; detailed description of the system in Bader and Körner 2010). Polypropylene collars (\emptyset 20, 5–7 cm height), inserted ca. 2–3 cm into the soil, served as a socket and seal for the chambers. We installed three collars per tree in 2 m distance to the stem base at a 120° angle around each tree, serving as replicates for each tree. These collars were left in place throughout the course of the experiment. Photosynthetically active tissue inside the collars (very minor understorey herbs) was removed prior to R_{soil} measurements, but litter was left in place to ensure natural conditions. Monthly measurements started in July 2008, 1 year before FACE initiation, and were intensified after the onset of FACE on 30 July 2009. During winter, measurements were suspended when snow covered the ground. Chamber recordings were performed at maximum daytime R_{soil} rates (i.e. from 1 to 6 pm), alternating between eCO₂- and aCO₂-trees to reduce any temporal bias. R_{soil} rates were calculated by applying a linear regression to the increase of the CO_2 concentration inside the chamber headspace over 4 min (60 recordings per 5 min, the first minute of each measurement were discarded to account for potential chamber placement effects). Soil temperature at 10 cm soil depth (T_{soil}) was recorded simultaneously adjacent to the collars using a KM20REF thermometer (Comark, Instruments, Norwich, UK).

Stem respiration

Stem CO₂ release, hereafter referred to as stem respiration (R_{stem} ; µmol CO₂ m⁻² s⁻¹), was measured using the LI-COR 6400-09 Soil CO₂ Flux Chamber connected to a LI-6400XT Portable Photosynthesis System (LI-COR, Lincoln, Nebraska, USA). The soil chamber operated in a closed system mode, and CO₂ drawdown inside the headspace allowed us to measure multiple cycles. We recorded 2-3 cycles per measurement and calculated the average. Four circular polyethylene collars (\emptyset 10, 4–5 cm high) were attached to the stem surface of each tree at ca. 1.3 m above ground, facing the cardinal directions (N, E, S, W). We used hot-melt adhesive and sealent (Terostat-IX, Teroson, Ludwigsburg, Germany) to ensure airtight collar connection to the stem surface. These collars served as chamber sockets. We did not install T sensors inside the stem sapwood. Thus, air temperature measured directly on the bark (T_{bark}) served as temperature reference using the LI-COR 6000-09TC Soil Probe Thermocouple (LI-COR, Lincoln, Nebraska, USA). We started measurements prior to the start of FACE (pre-treatment). We regarded R_{stem} signals 7 days after the onset of FACE as pre-treatment signals since the lag between leaf-level C assimilation and signal detection in R_{stem} is ca. 12 days (Mildner et al. 2014). Measurements were taken in 1-3 month intervals in 2009 and 2010, with two final measurements early in 2011.

Data analysis

The T dependency of respiratory fluxes (soil and stem) was modeled using a nonlinear least squares regression following Lloyd and Taylor (1994):

$$R = R_{10} e^{\mathrm{E}_{0} \left(\frac{1}{56.02} - \frac{1}{\mathrm{T} - 227.13}\right)},\tag{1}$$

where *R* is the measured respiration rate (either R_{soil} or R_{stem}) and R_{10} the respiration rate at 10 °C (T_{soil} for

soil respiration, and T_{bark} for stem respiration), E_O is the activation energy. The Lloyd and Taylor (LT) approach outperforms conventional *R* versus T correlation models (Arrhenius, van't Hoff). The T sensitivity of R_{soil} or R_{stem} (Q_{10}) was modeled following:

$$Q_{10} = \left(\frac{R_2}{R_1}\right)^{\left(\frac{10^5C}{T_2-T_1}\right)},$$
 (2)

where R_1 and R_2 are the respiration rates at temperatures of 10 °C (T_1) and 20 °C (T_2) (at 10 cm soil depth or at the bark surface), respectively, derived from the modeled LT regression. Pre-treatment differences in $R_{\rm soil}$ between trees later exposed to eCO₂ and aCO₂ were accounted for by assigning a temperature dependent correction factor to R_{soil} of eCO₂-trees in the FACE period. This correction factor was calculated from the difference of the modeled LT curves (Eq. 1) between eCO_2 and aCO_2 during the pre-treatment years (2008 and 2009). However, since the pre-treatment data for R_{stem} did not allow for modeling a R versus T relationship (insufficient T range), R_{stem} of eCO₂-trees in the FACE period was standardized by the mean pretreatment aCO₂/eCO₂ difference, thus assuming that the T response of R_{stem} did not change. These corrected respiratory fluxes were used to model the LT regression of eCO₂-trees under FACE (Eq. 1), and further parameters (Q_{10} , R_{10} ; Eq. 2). Confidence limits for the modeled Q_{10} and R_{10} values were obtained from bootstrapped 95 % confidence intervals. Annual release of C by stems, or soil, respectively, was calculated based upon the modeled LT regression (Eq. 1) by summing the estimated hourly R rates from continuously logged temperatures for all investigated years. We used either hourly records of Tair to calculate the annual C release by stems, or continuously available groundlitter (soil surface) temperature, correlating well with T_{soil} to calculate annual C release by the soil (missing values in the second half of 2011 were reconstructed from T_{air}; see Fig. 1). We know that the T response of R_{soil} does not differ between day and night based on diurnal respiration measurements (Bader et al. in prep.). Data analysis was performed using the software R, version 2.15.0 (R Development Core Team 2011).

Statistical analysis

Linear mixed effects models fitted by restricted maximum likelihood were applied in all statistical

analyses using R, version 2.15.0 (R Development Core Team 2011; R package nlme). The replicated unit in this project was a single 'tree' (five control trees under aCO_2 , and five trees subjected to eCO_2). Therefore, all measurements per tree were averaged prior to analysis. Since both groups of trees were studied before FACE and after the onset of FACE, we defined a 'pretreatment' factor to account for any change that might have occurred between the pre-treatment and FACE period. We assessed the significance of the main effects using a backwards selection procedure that progressively removes all non-significant terms until the optimal model is attained. This means that all terms not contained in the final model were statistically not significant. Model selection was validated by likelihood ratio tests and the akaike information criterion. The random factor'tree' was included in all models. Where necessary, homogeneity violations were modeled using adequate variance function structures (power, constant power, exponential and constant variance structures, or a combination thereof), and independence violations were corrected by implementing temporal autocorrelation structures. Model assumptions were examined using diagnostic plots (i.e. residual and quantile-quantile plots).

Results

Climatic conditions

Annual T_{air} averaged 8.2 °C in 2010 (min. -8.6, and max. 24.4 °C), and at 10.1 °C in 2011 (min. -5.7, and max. 25.2 °C). The mean T_{litter} was 9.1 °C (min. -1.0, and max. 23.1 °C), and 8.1 °C (min. -1.3, and max. 19.4 °C) for 2009 and 2010, respectively. We fitted linear regression models with T_{litter} as response variable, and either Tair or Tsoil as predictors. The amount of explained variation seen in T_{litter} increased when T_{soil} was used instead of T_{air}. This is reflected in the lower R^2 between T_{litter} and T_{air} ($R^2 = 0.841$) compared to T_{litter} and T_{soil} ($R^2 = 0.967$). The T_{soil} record had some gaps so that T_{litter} could be used for $T_{\text{soil}}.$ Given the strong correlation between T_{soil} and T_{litter} , we only show T_{litter} (Fig. 1). Precipitation was normal during the study period (no exceptionally dry period; for daily precipitation records see Fig. 1). Soil moisture was always high and tended to be slightly lower (-1.1 vol. %) under the CO₂-treated spruce

trees compared to control trees before FACE (Fig. 1). This pattern did not change over the course of this experiment (Leuzinger and Bader 2012).

Tree fine root biomass

Irrespective of the later ongoing treatment (eCO₂ or aCO₂), there was significantly more biomass in the finest root fraction (78 \pm 8 g m⁻² in <0.5 mm) compared to the biomass of 0.5-2 mm fine roots $(18 \pm 3 \text{ g m}^{-2})$ collected in late March 2010 after only three months of late season CO2-enrichment ('Diameter' effect: P < 0.001; Table 2; Fig. 2). These amounts of fine roots, collected from undisturbed soil, are supposed to depict the initial steady-state situation for this forest. Total fine root biomass (all diameter classes combined) of eCO₂-trees and aCO₂-trees did not differ (115 vs. 112 g m⁻²; no 'site' effect; P = 0.211; Table 2). However, we determined 27 % lower fine root biomass under eCO₂ compared to aCO₂ in the <0.5 mm diameter class (99 \pm 17 vs. 135 \pm 10 g m⁻²), whereas there was 29-61 % higher biomass under eCO₂ relative to aCO₂ in the 0.5–1 mm (51 \pm 8 vs. 39 \pm 4 g m⁻²), and



Fig. 2 Fine root biomass under *Picea abies* trees. *White bars* indicate data from trees exposed to ambient CO₂, and *grey bars* specify initial root data for trees exposed to elevated CO₂ (in situ content of boreholes later used for in-growth cores), *black bars* show the fine roots accumulated in in-growth cores after 20 months (ambient CO₂: n = 5 trees, elevated CO₂: n = 5 trees, mean \pm SE). *Left panel*, fine roots in soil cores sampled at the beginning of the experiment. *Right panel*, fine roots in in-growth cores after 20 months of FACE

1–2 mm (45 \pm 9 vs. 28 \pm 5 g m⁻²) categories at the start of the experiment (significant 'root thickness × site' interaction at *P* = 0.001; Table 2; Fig. 2).

Generally, there was a high proportionality between in-growth fine root mass and initial mass under in situ conditions (soil cores). The <0.5 mm diameter category had far more fine root biomass than the 0.5-2 mm classes $(117 \pm 18 \text{ vs. } 41 \pm 5 \text{ g m}^{-2}; P < 0.001; \text{ Table 2};$ Fig. 2). Overall, fine roots (all diameter categories combined) expanding into root-free soil exclusively during FACE (in-growth cores) produced similar biomass under eCO_2 and aCO_2 (202 vs. 195 g m⁻² under eCO_2) relative to aCO_2 ; P = 0.575; Table 2). However, similar to the soil cores, the in-growth cores showed more fine root biomass under eCO₂ relative to aCO₂ in the 0.5-1 and 1-2 mm diameter fractions (0.5-1 mm: 23 ± 8 vs. 21 ± 2 g m⁻²; 1–2 mm: 19 ± 5 vs. 8 ± 2 g m⁻²; Fig. 2). Yet, in the <0.5 mm root fraction we found 18 % lower fine root biomass in eCO2-trees relative to aCO₂-trees (70 \pm 19 vs. 86 \pm 13 g m⁻²; Fig. 2), which was again similar to the initial pattern seen in undisturbed soil cores ('Diameter \times CO₂' effect: P = 0.036; Table 2). Irrespective of the CO₂ treatment and diameter class, fine root dry mass in in-growth cores (227 g m^{-2}) arrived at only 57 % of the fine root mass previously found in soil cores (397 g m^{-2} ; Fig. 2). This finding suggests that the soil space in in-growth cores was not fully explored after 20 months under FACE.

Soil respiration

Pre-treatment measurements (all records collected during the 12 months before the start of FACE on 30 July 2009) revealed 0.6 \pm 0.1 $\mu mol~CO_2~m^{-2}~s^{-1}$ higher $R_{\rm soil}$ rates under later eCO₂-trees compared to aCO₂trees (3.5 \pm 0.2 vs. 2.9 \pm 0.2 $\mu mol~CO_2~m^{-2}~s^{-1}$ in later eCO₂-trees vs. aCO₂-trees; Fig. S1). A trend towards slightly lower rates of R_{soil} under eCO₂ relative to aCO₂ was observed when standardizing R_{soil} of eCO₂trees during FACE by the T-dependent difference during the pre-treatment years (reduction of $0.2 \pm 0.1 \ \mu mol$ $\rm CO_2\,m^{-2}~s^{-1}$ in 2010, and 0.3 \pm 0.1 $\,\mu mol\,\rm CO_2\,m^{-2}~s^{-1}$ in 2011; mean \pm SE; Fig. 3). This corresponds to 90 \pm 4 and 86 \pm 3 % of R_{soil} under aCO₂ in 2010 and 2011, respectively. The annual mean of R_{soil} of aCO₂-trees was 2.4 ± 0.1 (2009), 2.1 ± 0.1 (2010), and $2.2 \pm 0.1 \text{ }\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ (2011), whereas R_{soil} of eCO₂-trees averaged at 2.3 \pm 0.1 (2009), 1.9 \pm 0.1 (2010), and $2.0 \pm 0.1 \ \mu mol \ CO_2 \ m^{-2} \ s^{-1}$ (2011).



Fig. 3 Soil respiration (R_{soil}) and soil temperature at 10 cm depth of mature *Picea abies* exposed to ambient, or elevated atmospheric CO₂ concentrations in 2008, 2009, 2010, and 2011 (ambient CO₂: n = 5 trees; elevated CO₂: n = 5 trees; mean \pm SE). R_{soil} of trees exposed to elevated CO₂ was

Hence, the significant main 'CO₂' effect (P = 0.025; Table 2), and the 'CO₂ \times pre-treatment' interaction (P < 0.001) indicate that the pattern observed before the initiation of FACE differed significantly from the pattern observed after the onset of FACE, with significantly lower R_{soil} under eCO₂-trees relative to aCO2-trees. Also the cumulative annual C release was lower under eCO₂-trees (Fig. 5; Table 1). During the pre-treatment period, designated eCO₂-trees showed 19–23 % higher annual R_{soil} but, after correcting for the pre-treatment differences, this signal reversed, resulting in 8-11 % lower levels during the FACE periods in 2010-2011 after correcting for the pre-treatment differences (Fig. S3; Fig. 5; Table 1). R_{soil} revealed a distinct seasonality that was determined by the seasonal course of T_{soil} . T_{soil} explained 38–88 % of the variation in R_{soil} (P < 0.001; Fig. 3; Tables 1, 2). Maximum R_{soil} was measured in July 2009 just before the onset of FACE (eCO₂: 4.2 ± 0 ; and aCO₂: $4.2 \pm 0.2 \mu mol$ CO₂ $m^{-2} s^{-1}$ at a T_{soil} of c. 16 °C). Soil moisture influenced R_{soil} only in interaction with T_{soil} (P = 0.036). The statistically insignificant two-way interactions $(CO_2 \times T_{soil} P = 0.114; CO_2 \times soil moisture P =$ 0.287; Table 2) indicate that the observed CO_2 enrichment effect was independent of these parameters.

Stem respiration

Instantaneous mid-summer rates of R_{stem} were 4.5 \pm 0.2 and 3.5 \pm 0.3 µmol CO₂ m⁻² s⁻¹ in later eCO₂-trees and in aCO₂-trees, respectively, before the

corrected for the pre-treatment difference observed between control and treated trees (see materials and methods). Soil temperature under elevated and ambient CO₂ did not differ (*n.s.*). Therefore, the mean of all trees is plotted (n = 10). The grey-shaded areas on top of the panels denote the FACE periods

FACE treatment became effective (Fig. S2). Thus, R_{stem} in designated eCO₂-trees was 1.0 µmol CO₂ $m^{-2} s^{-1}$ higher relative to aCO₂-trees during this period of peak R_{stem} (a 29 % higher signal; Fig. S2). Since T_{bark} was similar among the treatments, the different rates of R_{stem} observed before FACE initiation reflects tree-specific differences (Fig. S2; Fig. S3). Accounting for this pre-treatment difference, R_{stem} of eCO₂-trees was slightly but not significantly $(0.1 \pm 0.1 \text{ }\mu\text{mol CO}_2 \text{ }\text{m}^{-2} \text{ }\text{s}^{-1})$ lower than in aCO₂trees across all years under FACE (mean \pm SE of R_{stem} in eCO₂-trees and aCO₂-trees, respectively: 1.8 ± 0.4 and $1.9 \pm 0.4 \ \mu mol CO_2 \ m^{-2} \ s^{-1}$; Table 2; Fig. 4). The mean Q_{10} for 2010 was 1.9 for both treated and control trees (95 % CI: 1.5-2.3 for eCO₂trees, and 1.6–2.3 for aCO₂-trees; Fig. 5; Fig. S3; Table 1). Q_{10} data for 2009 and 2011 suffered from insufficient sample size (large 95 % CI; Table 1). Accounting for the ca. 29 % higher peak season pretreatment signal under eCO_2 compared to aCO_2 (Fig. S2), the cumulative C release from stems over the year 2009 was similar (4 % higher) under eCO₂ compared to aCO₂ (n.s.). During FACE, eCO₂-trees respired less than aCO₂-trees (-7 % in 2010, and -14 % in 2011; Table 1). Irrespective of the CO₂ treatment, R_{stem} correlated with T_{bark} (P < 0.001), accounting for 60–81 % of the seasonal variation in R_{stem} (Table 1). Accordingly, R_{stem} peaked in late summer and coincided with highest T_{bark} (eCO_2: 4.0 \pm 0.6 $\mu mol~CO_2$ m^{-2} at 29.2 °C, and aCO_2: 4. 4 \pm 0.5 $\mu mol~CO_2~m^{-2}$ at 28.9 °C in July 2010; mean \pm SE).

Table 1 Nonlinear regression estimates (Lloyd and Taylor 1994) of annual CO₂ efflux rates at 10 °C (R_{10}), temperature sensitivity (Q_{10}) with bootstraped 95 % confidence intervals

(CI), and annual cumulative C fluxes from stems and soil of Picea abies under elevated or ambient CO_2

CO ₂ efflux	Period	Treatment (CO ₂)	<i>R</i> ₁₀	95 % CI of <i>R</i> ₁₀	<i>Q</i> ₁₀	95 % CI of <i>Q</i> ₁₀	Explained variation (%)	Annual C release $(g m^{-2} a^{-1})^a$	Change (%) ^c
Stem	PF 2009 ^b	С	1.6	0.8-3.1	2.0	1.2-3.7	35	596	n.a.
	PF 2009 ^b	Е	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	
	F 2009	С	0.7	0.6–0.9	2.7	1.8-4.3	80	318	4
	F 2009	Е	0.8	0.5-0.9	2.7	1.7-6.8	63	329	
	F 2010	С	1.3	1.1-1.7	1.9	1.6-2.3	68	450	-7
	F 2010	Е	1.3	1.0-1.6	1.9	1.5-2.3	60	418	
	F 2011	С	0.9	0.1-1.6	2.5	1.5-30.0	74	460	-14
	F 2011	Е	0.9	0.2-1.2	2.4	1.7-10.1	81	396	
Soil	PF 2008 ^b	С	2.5	2.2-2.8	2.0	1.6-2.6	58	887	19
	PF 2008 ^b	Е	2.9	2.6-3.3	1.7	1.2-2.2	38	1059	
	PF 2009 ^b	С	2.4	2.2-2.7	2.2	1.7-2.8	52	906	23
	PF 2009 ^b	Е	3.0	2.9-3.3	2.0	1.7-2.4	62	1117	
	F 2009	С	2.4	2.2-2.6	2.0	1.7-2.4	66	909	4
	F 2009	Е	2.3	2.1-2.5	2.4	2.0-2.9	73	948	
	F 2010	С	2.2	2.0-2.4	2.5	2.2-3.0	84	784	-8
	F 2010	Е	2	1.8-2.1	3.3	2.8-3.8	88	723	
	F 2011	С	2.1	1.9-2.3	2.5	2.1-3.0	80	845	-11
	F 2011	Е	1.8	1.6-2.0	2.8	2.3-3.5	77	750	

FACE Free air CO2 enrichment, PF pre-FACE, F FACE, C control trees, E CO2-treated trees

^a based on temperature records in the litter layer (soil CO₂ efflux), or 2 m above ground (stem CO₂ efflux)

^b Pre-FACE: uncorrected values

^c Percentage increase in the annual carbon release from trees subjected to elevated CO₂ versus control trees

Discussion

This project aimed at identifying respiratory and root growth responses in tall P. abies trees to elevated atmospheric CO₂ concentration. Stable carbon isotope signals allowed us to trace the fate of new carbon from the tree tops to the soil, and these data confirmed the effectiveness of our CO₂ treatment (Mildner et al. 2014). We expected that these 110-year-old spruce trees have reached a steady-state annual canopy and fine root renewal. A sudden exposure to a 150 ppm higher CO₂ concentration may thus cause strong initial, but declining long-term responses (Leuzinger et al. 2011; Norby and Zak 2011). In fact, we did not observe any significant downward adjustment of leaf-level photosynthesis at eCO₂ in current and previous year needles shortly after the onset of FACE in 2009 (n.s.; Fig. S5). In the fifth year of FACE, rates of photosynthesis remained enhanced at eCO₂, and the photosynthetic enhancement ratio was similar in control and CO_2 -treated trees (Klein, pers. comm.). This indicates that there is no photosynthetic acclimation to higher levels of CO_2 . Also, stomatal conductance remained unchanged (Leuzinger and Bader 2012). These results suggest that more C entered the trees under eCO₂. However, we found no stimulation of fine root accumulation, a reduced CO_2 efflux from soils, and unchanged CO_2 efflux from stems. In the following we will discuss these findings in the light of the results of other CO_2 enrichment experiments.

Fine root biomass

The in-growth core fine root samples reached slightly more than half the initial steady-state biomass, which suggests that 20 months are not enough to arrive at a new steady-state. Therefore, roots in in-growth cores were still in an expanding stage. Fine root growth showed no stimulation by eCO_2 until December 2011

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0.002**

able 2 Linear mixed	Factor	Df	F value	Р
<i>cea abies</i> fine root	Fine root biomass in soil cores (2010)			
omass (soil cores	Diameter	2,16	86.59	< 0.001***
-growth cores March 2010	Site	1,8	1.85	0.211
December 2011), and	Diameter \times site	2,16	14.27	0.001**
D_2 efflux from stems and	Fine root biomass in in-growth cores (2010-2011)			
il under ambient and	Diameter	2,16	31.13	< 0.001***
	CO_2	1,8	0.34	0.575
	Diameter \times CO ₂	2,16	4.14	0.036*
	Stem CO ₂ efflux 2009–2011			
	Bark temperature	1,106	24.99	< 0.001***
	Pre-treatment	1,106	12.70	0.001**
	Soil CO ₂ efflux 2008–2011			
	CO_2	1,8	7.55	0.025
	Pre-treatment	1,308	251.06	< 0.001***
	Soil temperature	1,308	1655.21	< 0.001***
	Soil moisture	1,308	1.29	0.257
	$CO_2 \times pre-treatment$	1,308	13.44	< 0.001***
	$CO_2 \times soil$ temperature	1,308	2.51	0.114
	Pre-treatment \times soil temperature	1,308	0.78	0.378
	$CO_2 \times soil moisture$	1,308	1.14	0.287
	Pre-treatment × soil moisture	1,308	0.12	0.733
	Soil temperature \times soil moisture	1,308	4.44	0.036*

* *P* < 0.05; ** *P* < 0.01; *** P < 0.001



Soil moisture \times soil temperature \times pre-treatment

Fig. 4 Stem respiration (R_{stem}) and bark surface temperature of mature Picea abies exposed to ambient, or elevated atmospheric CO₂ concentrations in 2009, 2010, and 2011 (ambient CO₂: n = 5 trees; elevated CO₂: n = 5 trees; mean \pm SE). R_{stem} of trees exposed to elevated CO2 was corrected for the pre-

treatment difference observed between control and treated trees (see materials and methods). Bark surface temperature under elevated and ambient CO₂ did not differ (n.s.). Therefore, the mean of all trees is plotted (n = 10). The grey-shaded areas on top of the panels denote the FACE periods

1,308

9.80



Fig. 5 *Picea abies* stem respiration (R_{stem}) response to bark surface temperature (*upper panels*), and soil respiration (R_{soil}) response to soil temperature 10 cm below ground (*lower panels*) during the FACE periods of the years 2009, 2010, and 2011. R_{stem} and R_{soil} of trees exposed to elevated CO₂ were corrected for the pre-treatment difference observed between control and treated trees (see materials and methods). The *inset* diagrams in 2009 depict the pre-treatment uncorrected R_{stem} (*upper inset*) and R_{soil} (*lower inset*) response in the period before the initiation

when compared to the pre-treatment signals. These results contrast observations in juvenile trees where increased fine root production at eCO_2 was found in *P. abies* grown in open top chambers (Lebègue et al. 2004), or in Glass Domes (Pokorný et al. 2013), regenerating birch/aspen stands, three *Populus* species, and young deciduous trees of three species, all grown under FACE (Lukac et al. 2003; Pregitzer et al. 2008; Smith et al. 2013). In the Oak Ridge FACE experiment, a plantation of *Liquidambar styraciflua* (21-year-old in 2009) showed an increase in fine root production and mortality during the first 7 years (Norby et al. 2004), but the signal disappeared after

of FACE in 2009. All respiration measurements were fitted with Lloyd and Taylor (1994) functions. Trees were exposed to ambient (*open symbols, dashed line*), or elevated atmospheric CO₂ concentrations (*filled symbols, solid line*). Each *symbol* represents the mean R_{stem} or R_{soil} rates measured per tree (n = 2-4) and measurement campaign. The Q_{10} values indicate the mean increase in the R_{stem} or R_{soil} rate per 10 °C temperature increase (from 5 to 15 °C)

11 years due to progressive N-limitation (Norby et al. 2010). An initial stimulation of fine root production by eCO_2 was also reported for a young closed-canopy *Pinus taeda* plantation at the Duke FACE (Allen et al. 2000). These studies investigated young trees, which may not have completely explored the available soil volume, and mostly grew under ample nutrient supply (expanding systems; Körner 2006).

The soil space beneath mature trees in a fully-grown forest can be expected to be fully explored by roots and to have arrived at a steady-state fine root turnover, which would prevent stimulation by eCO₂ (Norby et al. 1999; Körner 2006). The 110-year-old trees studied in our

web-FACE experiment operated at constant annual needle renewal rates (unpublished litter production data) and, thus, should also be in a steady-state of fine root renewal, not affected by eCO₂ (Körner 2006; Norby and Zak 2011). A Swiss treeline FACE study on 35-year-old L. decidua and Pinus uncinata, both in a quasi steady-state development, did not reveal any fine root growth following high CO2 exposure despite higher soil CO₂ efflux (Handa et al. 2008; Dawes et al. 2013; Hagedorn et al. 2013). Additionally, a CO₂ enrichment experiment in a scrub-oak system in Florida showed an initial burst of fine root production under eCO₂ after disturbances (fire and hurricane), a signal that gradually vanished in the following years (Day et al. 2013) with canopy closure (full LAI recovery; Palmroth et al. 2006). The former web-FACE study at our site on mature deciduous forest trees showed even reduced fine root production after 7 years of eCO₂ (Bader et al. 2009). This was explained by stand maturation, and (stomata driven) reduced canopy transpiration. Thus, soil moisture savings reduced the need for intensified soil exploration by fine roots (Leuzinger and Körner 2007; Bader et al. 2009). In contrast, our spruce trees showed no reduction of sap flow when exposed to eCO_2 and, hence, exhibited no soil moisture savings that might be responsible for the missing fine root growth response (Leuzinger and Bader 2012).

Abundance of soil nutrients, especially the availability of N, determines how fine roots will respond to eCO₂ (Pregitzer et al. 1995; Curtis and Wang 1998; De Graaff et al. 2006; Dieleman et al. 2010), regardless of tree or stand age. Recently two meta-analyses investigated the interactive effects of high CO₂ and N availability in soils, with high soil N fueling the CO₂ effect on fine root growth (De Graaff et al. 2006; Dieleman et al. 2010). This contradicts our results since we found no fine root response to eCO₂ despite decades of N-deposition of ca. 20 kg N $ha^{-1}a^{-1}$ at our site. Additionally, CO₂ enrichment induced soil nitrate release both in the present study (unpublished data), and in the former web-FACE experiment on mature deciduous trees (Schleppi et al. 2012). In *Pinus* taeda at the Duke FACE site, N-fertilization reduced fine root biomass by ca. 12 % compared to unfertilized plots, accompanied by reductions in soil respiration (Jackson et al. 2009; Drake et al. 2011). In 6-8-yearold P. abies saplings, N-addition reduced fine root production in comparison to plots without extra N in CO₂-enriched plots (Spinnler et al. 2002). It appears that the trees in this near-natural, mature forest do not exhibit such N-mediated fine root responses to eCO₂.

Given the substantial atmospheric N deposition in the test region, PNL, caused by accelerated soil N withdrawal during long-term CO_2 enrichment (Luo et al. 2004), is unlikely to occur here and stimulate fine root expansion under high CO_2 (see Franklin et al. 2009; Garten et al. 2011 for PNL effects).

 CO_2 fertilization may also induce deeper rooting, a phenomenon commonly observed in CO_2 enrichment experiments (Lukac et al. 2003; Norby et al. 2004; Jackson et al. 2009; Iversen 2010; Smith et al. 2013). However, we could not explore this possibility here, because the accessible soil profile at the SCC site is maximal 25 cm deep, with extremely rocky subsoil.

Soil respiration

In the short term, R_{soil} is mainly controlled by soil moisture and soil temperature (Raich and Schlesinger 1992; Davidson et al. 1998). When accounting for these covariates plus pre-treatment signals (Fig. S1), we detected a trend towards reduced R_{soil} in response to FACE. Spruce trees under eCO₂ also showed continuously decreasing annual C returns to the atmosphere compared to control trees (Table 1). The (moderate) reduction of CO₂ release compared to pretreatment conditions is surprising, given that soil CO₂ efflux carried a clear ¹³C signal that indicates effective CO₂ enrichment and fast belowground allocation of new C (Mildner et al. 2014). The absolute reduction in $R_{\rm soil}$ in response to eCO₂ might be even more pronounced, had the soil space been fully occupied by P. abies fine roots instead of a ca. 50 % fraction of all fine roots, including those from neighboring deciduous trees. The finding of reduced R_{soil} contrasts with many examples for very young stands (mostly obtained in open top chambers, OTC) that showed increased but highly variable R_{soil} under eCO₂ (plus 5–93 %) compared to aCO_2 (Zak et al. 2000). Forest FACE experiments in young plantations initially showed increases in R_{soil} rates at eCO₂, but these signals declined with time (Hamilton et al. 2002; King et al. 2004; Comstedt et al. 2006; Jackson et al. 2009; Norby and Zak 2011). Dieleman et al. (2010) summarized the results for 32 OTC and FACE sites using trees and found an average 19 % increase in $R_{\rm soil}$, with soil N fertilization enhancing the CO₂ effect. However, a few CO₂ enrichment experiments

showed no stimulation or a decline of R_{soil} , e.g. soil under mature deciduous trees subjected to web-FACE at our study site (N-fertilized soil) did not release more CO₂ under eCO₂. This was attributed to higher soil moisture at eCO₂ that may have impeded soil CO₂ efflux (Bader and Körner 2010). Furthermore, Tingey et al. (2006) reported declining rates of R_{soil} in Ponderosa pine seedlings subjected to eCO₂ in growth chambers, caused by altered R_{soil} sensitivity to soil temperature and soil moisture at eCO₂.

The extent to which R_{soil} responds to eCO₂ has been found to be strongly related to responses of fine roots (Zak et al. 2000; Jackson et al. 2009; Drake et al. 2011). Root respiration (and associated mycorrhizal fungal respiration) can contribute 50–65 % to total R_{soil} (Andrews et al. 1999; Högberg et al. 2001, 2002; Bhupinderpal-Singh et al. 2003), and is fueled by fresh aboveground assimilates (Högberg et al. 2001). Therefore, the relative reduction in R_{soil} is likely to reflect reduced belowground C transfer under eCO2 (Palmroth et al. 2006; but see Jastrow et al. 2005). Generally, C supply to belowground microorganisms, or fungal symbionts was found to either increase with CO₂ fertilization, or did not change (Fransson 2012). In the short term, extra C is likely to increase the abundance of microorganisms (e.g. fungi and bacteria; Blankinship et al. 2011) which may become competitors for essential plant nutrients (Diaz et al. 1993; Hättenschwiler and Körner 1998; Inauen et al. 2012). Likewise, heterotrophic rhizomicrobial respiration could decline when exudates alter the microbial community (Bader and Körner 2010), its activity (Drake et al. 2011), or species composition (Carney et al. 2007; Drigo et al. 2008; reviewed in Zak et al. 2000). A higher release of nitrate under eCO₂ relative to aCO₂ (Schleppi and Textor, pers. comm.; similar to Schleppi et al. 2012) could also contribute to reduced microbial activity. However, we expected the 'priming effect' (Jenkinson et al. 1985) to dominate, as was found in the Duke FACE study (Drake et al. 2011; Phillips et al. 2012) that reported slowly increasing R_{soil} over the course of 12 years of FACE (Jackson et al. 2009). The tall, 110-year-old trees in our study may either respond more slowly, or have their roots spread over such a large area that R_{soil} signals get diluted.

Stem respiration

During the first 2.5 years of web-FACE, there was no indication of a CO_2 -driven decline or increase of R_{stem}

in these mature P. abies trees, although a strong stable C isotope signal in respiratory CO_2 evidences that the novel C derived from web-FACE (Mildner et al. 2014). The lack of any stem growth stimulation at eCO₂ in these trees (the 2009–2014 mean basal area increment standardized by mean pre-treatment rates was 1.4 ± 0.1 at aCO₂ and 1.5 ± 0.3 at eCO₂; n = 6 years, mean \pm SE; Klein and Körner; unpublished), given the assumption that the stem diameter increment largely determines the magnitude of R_{stem} signals under eCO_2 (Zha et al. 2005; Moore et al. 2008), co-explains why we also see no R_{stem} signal in response to web-FACE. In contrast to these results, juvenile trees exposed to a step increase in CO₂ on fertile ground, or with ample soil space, grew faster and their stems respired more. For instance, R_{stem} was 16 % higher in eCO₂ in 16-year-old P. abies (Acosta et al. 2010). Similarly, an increase in R_{stem} in response to eCO₂ was observed in 15-year-old Liquidambar styraciflua (Edwards et al. 2002), and in 20-year-old P. sylvestris (Zha et al. 2005). Stem growth and the associated R_{stem} responses to eCO₂ are largely determined by the developmental stage (age) of a tree, the species investigated, and the nutrient supply (Körner 2006). CO_2 enrichment may also contribute to higher maintenance respiration (Carey et al. 1996; Edwards et al. 2002; Zha et al. 2005), and mature trees exhibit higher maintenance respiration rates than juvenile trees (Ryan and Waring 1992). Hence, mature trees might be expected to yield even greater responses, but this is in contrast to what we found. R_{stem} signals might become diminished by translocation of dissolved CO₂ in sap flow (Negisi 1979; Teskey and McGuire 2002; Moore et al. 2008; Bloemen et al. 2013). However, sap flow measured prior to web-FACE was similar in the trees examined, and this relation did not change at eCO_2 (Leuzinger and Bader 2012). Whatever the reason, these tall trees did not exhibit a greater R_{stem} response under web-FACE.

Conclusions and outlook

Previous and ongoing works revealed that photosynthesis, was, and still is, enhanced under eCO_2 , and stomatal conductance remained unaffected by eCO_2 (Leuzinger and Bader 2012). Therefore, we expected strong and positive initial responses to a step increase in CO_2 in both types of respiratory CO_2 release, and in fine root growth in these tall trees. The fact that we did not detect such a stimulation, despite clear isotopic evidence of successful canopy CO2 enrichment, by default, suggests other pathways of C-dissipation under eCO₂. We expected such overflow responses because we (seemingly correctly) anticipated no stem growth response for reasons related to tree nutrition (other than by N), and tissue element stoichiometry (ongoing research). It remains to be seen if accelerated root growth will occur at a later stage, as was the case in other FACE works (Allen et al. 2000; Spinnler et al. 2002; Norby et al. 2004). The fine root data should be highly sensitive to CO₂ because fine roots from ingrowth cores had not yet arrived at steady-state root density, and the signal should still capture the root expansion process. The data presented here add to the growing evidence that mature trees or trees growing in stands that arrived at steady-state leaf and root turnover are unlikely to take benefit from eCO₂. These trees are likely to be C saturated at current ambient CO₂ concentrations, as has been shown for boreal spruce trees (Sigurdsson et al. 2013). We observed highly homeostatic stem respiratory signals, and soil CO₂ efflux even declined slightly in response to web-FACE.

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