

Dissolution and transformation of cerium oxide nanoparticles in plant growth media

Franziska Schwabe · Rainer Schulin ·
Patrick Rupper · Aline Rotzetter ·
Wendelin Stark · Bernd Nowack

Received: 8 August 2013 / Accepted: 20 September 2014 / Published online: 8 October 2014
© Springer Science+Business Media Dordrecht 2014

Abstract From environmental modeling of engineered nanomaterial (ENM) release, it is clear that ENMs will enter soils, where they interact with soil compounds as well as plant roots. We analyzed three different size groups of cerium dioxide nanoparticles (CeO₂-NPs) in respect to chemical changes in the most common plant growth medium, Hoagland solution. We created a simple environmental model using liquid dispersions of 9-, 23-, and 64-nm-uncoated CeO₂-NPs. We found that CeO₂-NPs release dissolved Ce when the pH of the medium is below 4.6 and in the presence of strong chelating agents even at pH of 8. In addition, we found that in reaction with Fe²⁺-ions, equimolar amounts of Ce were released from NPs. We could

elucidate the involvement of the CeO₂-NPs surface redox cycle between Ce³⁺ and Ce⁴⁺ to explain particle transformation. The chemical transformation of CeO₂-NPs was summarized in four probable reactions: dissolution, surface reduction, complexation, and precipitation on the NP surface. The results show that CeO₂-NPs are clearly not insoluble as often stated but can release significant amounts of Ce depending on the composition of the surrounding medium.

Keywords Nanoparticles · Ce³⁺ and Ce⁴⁺ · Dissolution · Hoagland medium · Fe species · Environmental effects

F. Schwabe · R. Schulin
Soil Protection, Institute of Terrestrial Ecosystems, ETH-Zurich, Universitaetstrasse 16, 8092 Zurich, Switzerland

P. Rupper
Laboratory of Advanced Fibers, Empa-Swiss Federal Laboratories for Materials Science and Technology, Lerchenfeldstrasse 5, 9014 St. Gallen, Switzerland

A. Rotzetter · W. Stark
Department of Chemistry and Applied Biosciences, Institute for Chemical and Bioengineering, ETH Zurich, 8093 Zurich, Switzerland

B. Nowack (✉)
Technology & Society Laboratory, Empa-Swiss Federal Laboratories for Materials Science and Technology, Lerchenfeldstrasse 5, 9014 St. Gallen, Switzerland
e-mail: nowack@empa.ch

Introduction

Engineered nanomaterials (ENMs) were intensely studied in the last 10 years. The broad variety of particle types, high production rates, and the use in costumer products have raised concerns about their release into the environment and their fate and effects after disposal (Gottschalk and Nowack 2011). The properties of ENMs are determined by their high surface-to-volume ratio, their crystal structure, and their reactive surface groups (Baalousha et al. 2010). The reactivity of ENMs is influenced by physico-chemical features such as size, shape, surface coating, surface charge, bioavailability, specific surface area, and aggregation (Batley et al. 2013). Chemical and

biochemical reactions like particle dissolution, adsorption of ions, interaction with cells, and uptake by organisms and combinations of these reactions are crucial to consider when studying the effects of ENMs on organisms in various environmental media (Handy et al. 2012; Baalousha et al. 2010; Scown et al. 2010).

In case of ENM emission (Gottschalk et al. 2009; Mueller and Nowack 2008; Nowack et al. 2012), plants are also of special interest since they are in direct contact with the important sinks: soil, air, and water (Miralles et al. 2012). As plants are the basis of the food chain, accumulation of ENMs in plants will allow them to pass through the food web (Ma et al. 2010). Zhu et al. (2008) were the first to observe ENM uptake by plants. Many studies on plant ENM uptake were carried out using aqueous suspensions or hydroponic media (Miralles et al. 2012). Cerium oxide nanoparticles (CeO₂-NPs) were often used for these studies because they were considered to be stable and insoluble (Madler et al. 2002; Brunner et al. 2006) in dispersion and even after uptake (Zhang et al. 2011). CeO₂-NPs are a good representative of ENMs because they are widely used, e.g., as catalyst, in fuel cells, for ceramic applications, and as diesel fuel additive (Baalousha et al. 2010). Recently, it was shown that CeO₂-NPs can be incorporated into plant roots (Du et al. 2011) and even translocate into shoots (Zhang et al. 2011; Schwabe et al. 2013). Further Zhang et al. (2012) stated that CeO₂-NPs do undergo biotransformation and are soluble to some extent in the presence of cucumber plants in hydroponic culture. Hence, the literature is contradictory about the potential solubility of CeO₂-NPs. In mostly all available CeO₂-NPs—plant studies, biotransformation and solubility of these NPs were not taken into account (Zhao et al. 2012; Wang et al. 2012).

Cerium naturally occurs in the Ce(III) and Ce(IV) oxidation states. CeO₂-NPs are crystallized in a fluorite structure, where each Ce(IV) is surrounded by eight equivalent O²⁻-ions, which is a very stable conformation (Deshpande et al. 2005). In bulk CeO₂, this atom conformation appears in 99.99 % of the material and accounts for its insolubility (Baalousha et al. 2010). CeO₂-NPs have the ability to absorb and release oxygen in a catalytic redox reaction on the surface. This feature plays a critical role in the overall performance of the catalytic activity of CeO₂-NPs and makes it interesting for industrial use. The catalytic activity is due to the change in the oxidation state of Ce atoms on the NP surface between Ce³⁺ and Ce⁴⁺

depending on the oxygen partial pressure and the pH in the surrounding medium (Trovarelli et al. 2001; Ilan et al. 1976). Introduction of Ce³⁺ into the crystalline structure results in oxygen vacancies (Deshpande et al. 2005). Baalousha et al. (2010) found 60 % of Ce³⁺ in CeO₂-NPs of 25 nm. The amount was significantly reduced when the NPs were dispersed in growth media. In a similar medium, van Hoecke et al. (2009) found only 0.3 % Ce³⁺ on the surface of 14-nm CeO₂-NPs. Thill et al. (2006) found that the ratio between Ce³⁺ and Ce⁴⁺ was altered after adsorption on the outer membrane of *E. coli* in 7-nm CeO₂-NPs from 4 % of Ce³⁺ to 29 % of Ce³⁺. Known as the cerium anomaly, oxides of Ce⁴⁺ are considered to be completely insoluble (Kakuwa and Matsumoto 2006), whereas Ce³⁺-oxide can be relatively soluble since it has a solubility product of logK_s = -21.5 (Smith 2004).

Taking these studies together, the crystallite structure and lattice changes during redox reactions as well as the Ce³⁺ to Ce⁴⁺ ratios in CeO₂-NPs have been well studied. But not a single report has combined the available information on structure and oxidation state and studied ion release from CeO₂-NPs at the same time. Understanding the reactions of CeO₂-NPs dissolution and transformation is essential for assessing toxicity and environmental fate. To evaluate realistic environmental scenarios, changes in chemical composition and physical features of particles need to be monitored already before they interact with organisms (Nowack et al. 2012). We studied CeO₂-NP dissolution in the most common plant growth medium “Hoagland solution.” Here, we evaluated the individual effects of the constituents of this medium on three different size groups of uncoated CeO₂-NPs and monitored changes in the presence of gum arabic (GA) as representative of dissolved organic matter in such media.

Materials and methods

Nanoparticles

The cerium oxide nanoparticles used in this study were manufactured by flame-spray synthesis. By varying the manufacturing process, three different sizes were produced. They are referred to by their mean diameter size (9, 23, and 64 nm) obtained from TEM image evaluation of 100 single primary particles. Particles were non-coated CeO₂-NPs. Flame-spray synthesis was

performed using cerium 2-ethylhexanoic acid diluted in xylene (2 wt % for the 9-nm CeO₂-NPs and 8 wt % for the 23-nm CeO₂-NPs), which was introduced as a precursor into a methane/oxygen flame (Madler et al. 2002). For the 9-nm NPs, the flow rate of the precursor was 3 mL min⁻¹ (5 mL min⁻¹ for the 23 nm CeO₂-NPs), and 7 mL min⁻¹ oxygen gas (5 mL min⁻¹ for the 23-nm CeO₂-NPs) was used to disperse the liquid leaving a capillary (Stark et al. 2003). The 64-nm CeO₂-NPs were made from 23-nm CeO₂-NPs after sintering for 16 h at 700 °C (Limbach et al. 2005).

BET, TEM, and XRD

Particle size distributions and specific primary particle diameters were determined by Brunauer–Emmett–Teller (BET) via N₂ adsorption using a Micromeritics Tristar 3000, by size surveying a 100 primary particles on images obtained by the means of transmission electron microscopy (TEM) (Tecnai F30 ST, FEI, operated at 300 kV), and by X-ray diffraction (XRD) (X'Pert PRO-MPD, Cu K α radiation, X'Celerator linear detector system, step size of 0.05°, 45 kV, 40 mA, and ambient conditions).

X-ray photoelectron spectroscopy (XPS)

Particle surface speciation and elemental content of surfaces were determined by means of XPS (PHI 5600 spectrometer, Physical Electronics, U.S.A) using non-monochromatized Mg–K α radiation (1253.6 eV). The spectra were collected at a photoemission angle of 45° with respect to the surface normal. The operating pressure of the XPS analysis chamber was approximately 5 × 10⁻⁹ Torr. We acquired survey scan spectra (0–1,100 eV) and higher resolution narrow spectra for Ce. Curve fitting was carried out using CasaXPS software, Version 2.3.14. The potential charging of the sample surfaces was corrected by setting the graphitic and aliphatic (C–C, C–H) photoelectron signal contribution at 285.0 eV. Atomic ratios were calculated from XPS spectra after subtracting a Shirley-type background.

Particle composition

Total Ce was analyzed via X-ray fluorescence (XRF) (Spectro XEPOS Spectro Analytical Instruments, Germany). Samples were acquired by mixing 100 mg

of NPs with 3.9 g quartz purum (Fluka, 83340, Switzerland) in a laboratory shaker (MM200, Retsch, Switzerland). After homogeneous blending with 0.9 g wax (Cereox by Fluxana, BM-0002-1, Switzerland), a pellet was pressed at 15 bar.

To determine total carbon contents, 100 mg samples of pure NP was analyzed in a TOC-L & SSM 5000 A (Shimadzu, Japan).

Preparation of media and dispersions

All solutions were prepared with ultra-pure deionized water taken from a Millipore[®]-unit (TKA GenPure, Germany), subsequently referred to as Millipore. For the preparation of Hoagland medium, different stock solutions were generated. These were 1:1000 diluted and mixed freshly for each experiment, resulting in 20 % Hoagland medium, following Tandy et al. (2006). The final compositions of the medium were 800 μ mol/L Ca(NO₃)₂, 400 μ mol/L MgSO₄, 200 μ mol/L KH₂PO₄, 1,000 μ mol/L KNO₃, 10 μ mol/L Fe(III)-ETDA, 20 μ mol/L H₃BO₃, 4 μ mol/L MnSO₄, 0.4 μ mol/L ZnSO₄, 0.4 μ mol/L CuSO₄, 0.4 μ mol/L Na₂MoO₄, and 10 μ mol/L NaCl. Solutions were buffered using 5.1 mmol/L MES (2-N-morpholino-ethanesulfonic acid) and either 1 M HCl or 1 M KOH for pH adjustment.

Nanoparticles were dispersed using ultrasonication (Ultralab 4000, B. Braun, Germany) for 10 min at 160 W in Millipore. In all experiments which are marked “+ GA”, gum arabic was added during this step. The concentration of GA was varied, proportional to the surface area of the NP group, for 9-nm CeO₂ → 90 mg/L GA, 23-nm CeO₂ → 15 mg/L GA, and for 64 nm-CeO₂ → 10 mg/L GA. The resulting suspension of 1 g/L NPs, was diluted to a final concentration of 100 mg/L CeO₂-NPs in 20 % Hoagland or solution of Hoagland component. All resulting dispersions were again adjusted to experimental requested pH. Dispersions were incubated in 50-ml sample tubes at room temperature for 7 days on a shaded shaking device at 100 rpm (Lab-shaker, Adolf Kühnert AG, Switzerland). The pH values stated in this study are the measured values of samples on d 7. All tested dispersions were generated in at least two individual replicates.

Characterization of CeO₂-NPs in suspensions

Using a Zetasizer 3000 (Malvern, UK), the particle hydrodynamic diameters were determined in nm by

dynamic light scattering (DLS) and the surface charges (zeta potentials in mV) were determined by mobility measurements. For all suspensions 3 individual samples were analyzed, each in 5 technical replicates (4 h after NP ultrasonication).

Analysis of Cerium

Quantification of Ce content of NPs was carried out after a two-step acid digestion with conc. HNO_3 for 60 min and in a 2nd step with 38 % H_2O_2 for another 90 min. The microwave oven was set in both steps (lavis Ethos EM-2, MLS GmbH, Germany) at 165 °C max. temperature. The solutions were analyzed for Ce using ICP-MS (ICP-MS-920, Varian, Switzerland).

For quantification of dissolved Ce (7 days of incubation), the NP dispersions were filtered through 10 kDa Amicon® Ultra-4 Centrifugal Filter (Merck, Millipore, Switzerland) for 45 min at 4,000 rpm. The filtered solutions were measured via ICP-MS after addition of 1 % HNO_3 . The NP-free control media (without NPs) were analyzed in the same way as NP containing suspensions (the nutrient content of pure media was the same before and after filtration). The detection limit for Ce was 0.2 $\mu\text{g/L}$, calculated from the count rate of 10 blank samples as mean + STDV, compared to the mean count rate + STDV of 10 measurements of the standard calibration solutions.

Speciation calculations

Speciation of Ce(III) in solution and precipitation of $\text{Ce}(\text{OH})_3$ and $\text{Ce}(\text{III})\text{PO}_4$ were calculated using VisualMinteq with the stability constants given in the VisualMinteq database (Gustafsson 2011). The total

concentration of dissolvable Ce(III) was set at 23 μM , the values obtained by EDTA-extraction, accounting for 4.2 % of the total Ce in the system.

Results

Particle characterization

TEM images of the three CeO_2 -particle groups used in this study are shown in Fig. 1. Particles were heterogeneous in size, but the size ranges of primary particles clearly separate all three groups. The groups were characterized using different techniques (Table 1). According to the TEM images of primary particles, the largest group (64 nm) was composed of 10 % NPs larger than 100 nm and 2 % under 30 nm but no NPs smaller than 25 nm. In the 23 nm NP group, 50 % of analyzed NPs were smaller than 20 nm, the smallest NPs were 5 nm (1 %) and the largest 80 nm (1 %). The smallest NP group had a mean diameter of 9 nm and consisted of NPs with a minimal diameter of 4 nm to a maximum of 24 nm (Table 2).

The BET results showed that the mean surface area of the smallest group was 10 times higher than the 23-nm NPs and 47 times higher than the surface of 64-nm NPs.

The three particle groups were also characterized in suspension. After ultrasonication of the particles in Millipore water, the pH as well as the zeta potential differed strongly between the three particle groups. The smaller the particle diameter was, the higher was the zeta potential. In Hoagland solution or GA, the zeta potential changed to negative values, and the differences between the three particle groups were reduced.

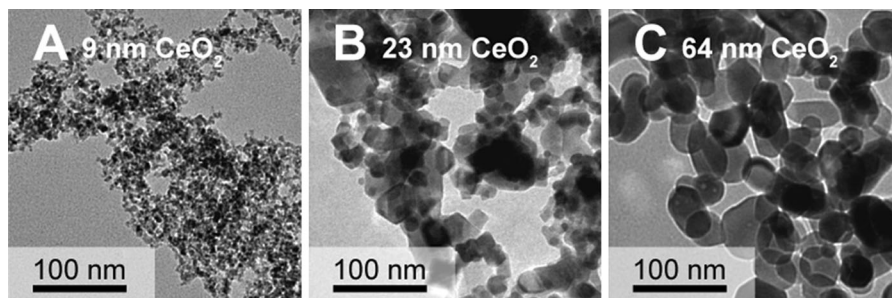


Fig. 1 TEM images of CeO_2 -NPs, after ultrasonic dispersion in Millipore with 100 μL polyacrylamide. **a** 9-nm particles, **b** 23-nm particles, and **c** 64-nm particles

Table 1 Characterization of CeO₂-primary particles as obtained after flame-spray synthesis

| NP group | TEM (nm) | XRD (nm) | BET (g/m ²) | XPS (Ce ³⁺ in %) |
|------------------------|-----------|-----------|-------------------------|-----------------------------|
| 9-nm CeO ₂ | 8.9 ± 4 | 6.2 ± 0.5 | 233 ± 0.4 | 22.9 |
| 23-nm CeO ₂ | 22.8 ± 14 | 27.7 ± 1 | 22.4 ± 0.2 | 11.1 |
| 64-nm CeO ₂ | 63.9 ± 28 | 48.3 ± 5 | 4.9 ± 0.1 | 9.7 |

Primary particle size measured: as diameter in nm by image recognition of TEM-pictures ($n = 100$); XRD-mean from 4 individual runs, given as calculated diameter in nm; BET surface area, XPS for surface speciation of Ce given as % Ce³⁺ on primary particle surface

The DLS size measurements given in Table 3 indicate that particles agglomerated fast and sizes exceeded 1,000 nm in 20 % Hoagland medium when no stabilizing agent (gum arabic) was present.

Surface speciation of Ce NPs

In addition to the elements cerium, oxygen, carbon, and sulfur on the surface of the particles, the XPS survey scans showed significant amounts of sodium in the 23- and 64-nm NP groups. No other elements were found.

Sulfur resulted from the sulfate groups present in the samples, whereas carbon came from the organic matrix. High-resolution spectra were measured for the element cerium. Caution must be taken when estimating the Ce(III) and Ce(IV) amounts from a sample using XPS. An overestimation of the Ce³⁺ concentration in ceria nanoparticles due to surface reduction of ceria in the XPS vacuum chamber enhanced by X-ray radiation has been reported (Zhang et al. 2004b). Therefore, to estimate the amounts of Ce(IV) and Ce(III), a reference sample of Ce(IV) was measured first. No significant amount of Ce(III) was found in the Ce(IV) reference sample. We therefore concluded that, under our experimental XPS conditions, a surface reduction of cerium in our XPS chamber is not a major issue. Based on these findings, spectra of the Ce(IV) speciation of the NP samples were measured. These spectra are given in Fig. 2 for the three NP groups: 9 nm (Fig. 2a), 23 nm (Fig. 2b), and 64 nm (Fig. 2c). The spectrum of cerium is rather complex, (spin-orbit splitting, hybridization with ligand orbitals and partial occupancy of the valence 4f orbital) but well known from the literature (Teterin et al. 1998; Zhang et al. 2004b). The experimental spectra (red lines) were

Table 2 Characterization of CeO₂-NPs in suspensions

| Medium | pH | | | DLS size (nm) | | | Zeta potential (mV) | | |
|-----------------------------|-----|-----|-----|---------------|---------------|----------------|---------------------|-------------|-------------|
| | 9 | 23 | 64 | 9 | 23 | 64 | 9 | 23 | 64 |
| Millipore | 5.0 | 7.8 | 6.0 | 140 ± 3 | 129 ± 1 | 1,182 ± 69 | 44.9 ± 6 | 21.3 ± 1 | 7.4 ± 0.3 |
| Millipore + GA | 5.2 | 7.4 | 6.3 | 244 ± 9 | 146 ± 2 | 260 ± 10 | -24.7 ± 6 | -26.2 ± 1 | -18.1 ± 0.2 |
| Hoagland + GA pH 6 | 5.6 | 5.6 | 6.0 | 184 ± 2 | 166 ± 3 | 948 ± 77 | -11.5 ± 0.5 | -13.6 ± 0.2 | -21.5 ± 0.5 |
| Hoagland without GA pH 6 | 5.6 | 5.6 | 6.0 | 3,023 ± 263 | 8,874 ± 1,637 | 29,410 ± 4,086 | -9.2 ± 4 | -16.4 ± 0.3 | -26.5 ± 0.8 |

Particle characteristics in different solutions 4 h after ultrasonication. pH of analyzed dispersion, DLS (dynamic light scattering) $n = 3$, ZP (zeta potential) mean of 3 replicates, measured 5 times ± STDV

Table 3 Characterization of 100 mg primary CeO₂ particles as obtained by flame-spray synthesis

| NP group | Total Ce (%) | Total Ce (μmol/100 mg) | Total Carbon (%) | Purity of CeO ₂ (%) | Total Ce (μmol/L) |
|------------------------|--------------|------------------------|------------------|--------------------------------|-------------------|
| 9-nm CeO ₂ | 77.2 | 551 | 1.2 | 96.3 | 488 |
| 23-nm CeO ₂ | 81.4 | 580 | 0.2 | 97.3 | 533 |
| 64-nm CeO ₂ | 85.2 | 607 | 0.05 | 97.8 | 545 |

Particles were analyzed using XRF for total elements and TOC analyzer for total carbon content. The purity of the particles was calculated based on the stoichiometry of CeO₂. For comparison, the Ce content measured by ICP-MS after acid digestion of 100 mg/L NPs is given in the last column as total Ce in mg/L acid digest

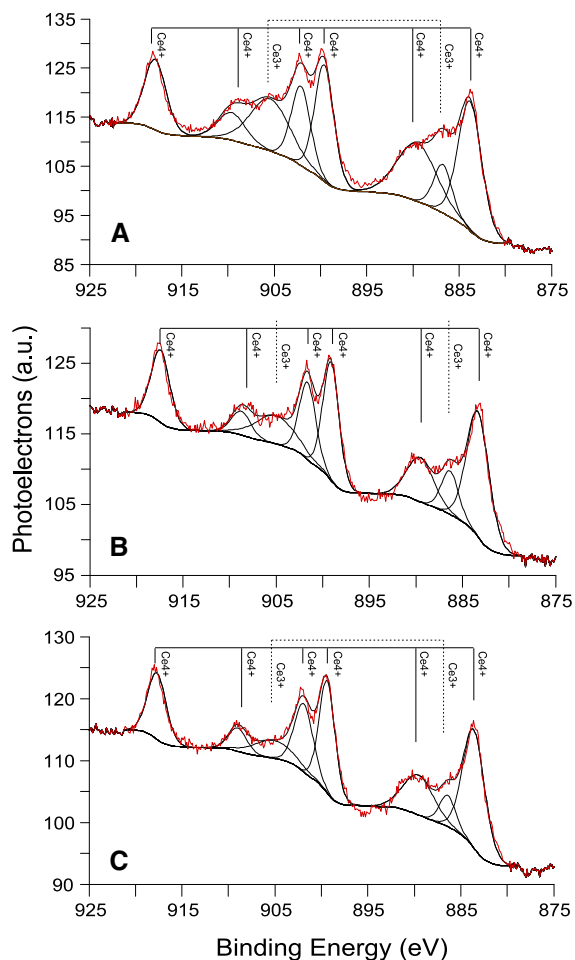


Fig. 2 XPS analysis of the Ce3d electrons in the three particle types. **a** Spectra of 9-nm CeO₂-NPs, **b** spectra of 23-nm CeO₂-NPs, and **c** spectra of 64-nm CeO₂-NP

fitted after deconvolution into eight components (black lines) representing the various Ce peaks (six belonging to Ce⁴⁺ and two to Ce³⁺). The spin–orbit splitting was set to the values given by Teterin et al. (1998). All other energetic positions as well as all intensities and the full width at half maximum were determined by fitting. The single peak at 917.8 eV was well separated from the other peaks and represented Ce⁴⁺ alone. It was used for a quantitative speciation of surface Ce(IV), as described by Li et al. (2004) and Pardo et al. (2007).

$$\text{Ce}^{4+\%} = (\text{Peak}_{917.8\text{eV}}\% / 14) \times 100.$$

The corresponding amount of Ce³⁺ % was calculated as 100 % minus Ce⁴⁺ % because only Ce(III) and Ce(IV) oxidation states of cerium were expected.

Based on literature, we assumed that the surface layer probed in these XPS measurements is 2 nm thick (Seah and Dench 1979). Since the XPS analyses could give the speciation of surface ion, the resulting percentage of Ce(III) in the surface layer is given in Table 1.

We used these amounts to calculate the Ce(III) content of the whole particle, assuming a spherical shape. Since redox-reactions between Ce³⁺/Ce⁴⁺ are known to occur only on the NP surface, we expected that the NP core consists of 100 % Ce⁴⁺ (Baalousha et al. 2010). The particle core was defined as NP diameter—2-nm surface layer (Wu et al. 2004). Based on these assumptions we calculated the overall Ce(III) fraction for each NP group: 19 % in the 9 nm-NPs, 4.8 % in the 23 nm, and 1.7 % in the 64-nm particles.

Dissolution of particles in Hoagland medium

In all tested suspensions, the initial concentration of CeO₂-NPs was 100 mg/L (equal to 0.58 mmol/L). Using the XRF measurements given in Table 3, we calculated the total amount of Ce in solution, which would occur in the case of complete NP dissolution as 551 μmol/L for 9-nm NPs, 580 μmol/L for 23 nm NPs, and 607 μmol/L for 64-nm NPs. Table 3 also gives the amounts of total Ce content of a 100 mg/L NPs after acid digestion. These values were lower than XRF-values probably due to losses during the digestion or dilution. However, both methods indicated that the biggest NP group has the highest purity, which is likely due to the sintering process during production.

Figure 3a shows the dissolved Ce concentration for the three size groups after 7 days of incubation in 20 % Hoagland solution, in Hoagland with additional GA, and in phosphate-free Hoagland solution. At pH 6 without GA, only the 9-nm NPs released a detectable amount of Ce (0.04 μmol/L) into solution, but when GA was present, it released 0.06 μmol/L of Ce. The presence of GA resulted in a significant increase in Ce release from the 9-nm particles at pH 4. When the pH of the suspension was below 4.5, all three CeO₂-NPs released much higher amounts of Ce. The order of Ce release was 9 > 23 > 64 nm. The release of Ce was far higher from 9- to 23-nm NPs, when no phosphate was present in the medium. At pH 4 without phosphate, 38 times more Ce were released from 9-nm NPs (9.96 μmol/L) than in the presence of phosphate. This amount is 1.8 % of the total amount of

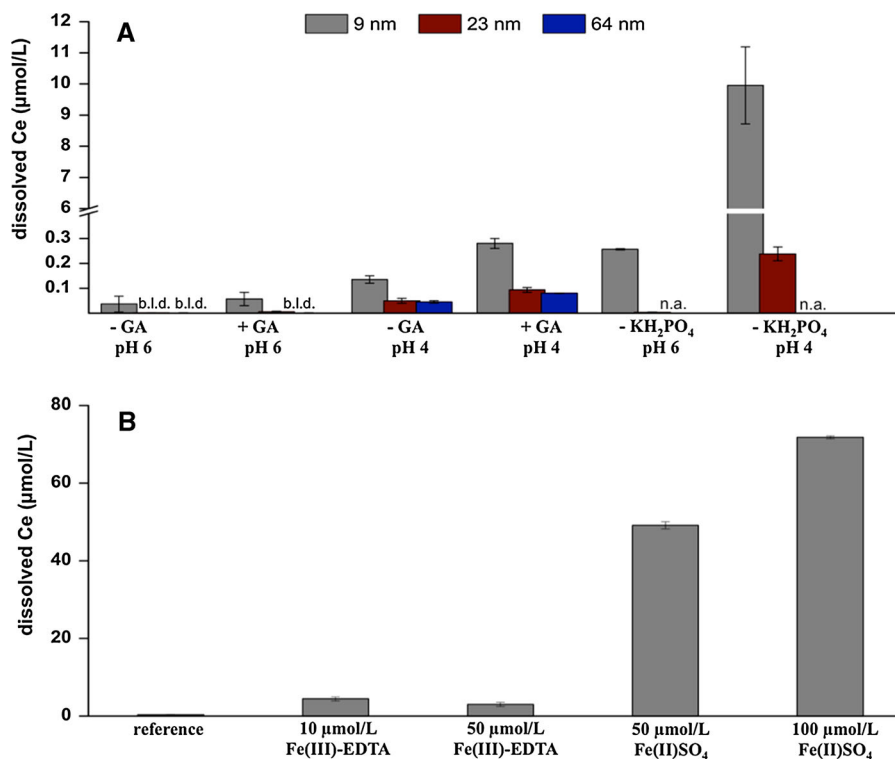


Fig. 3 Amount of dissolved Ce from 100 mg/L CeO₂-NPs. **a** in 20 % Hoagland solution after 7 days of incubation. -GA = no gum arabic was added, +GA = gum arabic was added; -KH₂PO₄ = no phosphate and no gum arabic was added. All dispersions were prepared at least in duplicate; error bars indicate standard error over all replicates per dispersion. b.l.d = below limits of detection, n.a. = not annotated (64-nm NPs were not measured in 20 % Hoagland without KH₂PO₄ because no detectable amount of Ce was expected). Break in y-axis:

0.4–6 µmol/L **b** Amount of dissolved Ce from 9-nm CeO₂ group (100 mg/L) after 7 days of incubation, in the presence of different Fe species. The “reference” sample shows the mean value of dissolved Ce in different Hoagland components. pH values of dispersions: reference, pH 4.3; 10 µmol/L Fe(III)EDTA, pH 4.5; 50 µmol/L Fe(III)EDTA, pH 4.5; 50 µmol/L Fe(II)SO₄, pH 4.8; and 100 µmol/L Fe(II)SO₄, pH 4.5; *n* = 3, error bars indicate standard error of replicates

Ce in suspension. The bigger 23-nm particles released less ions, but in the absence of phosphate, these NPs released 2.5 times more Ce compared to the same dispersion with phosphate.

Dissolution of CeO₂-NPs in presence of single Hoagland components

Since the smallest particles released the highest amounts of Ce, they were further used to analyze the influence of single Hoagland components on Ce on solubility. To get a clear picture of the chemistry behind NPs dissolution, 9-nm NPs were tested at pH 4.5 with the single components present in 20 % Hoagland. In Fig. 3b, the average amount of released Ce in 800 µmol/L CaNO₃, 800 µmol/L MgSO₄,

1,000 µmol/L KNO₃, and 1 g/L MES hydrate is used as the *reference* value, because there were no significant differences between Ce dissolution in the presence of all these compounds. Besides phosphate, we found that Fe species strongly influenced NPs dissolution. 10 or 50 µmol/L Fe(III)EDTA resulted in 4.4 and 3.0 µmol/L detectable Ce ions, respectively. Very high amounts of Ce were released when Fe was supplied as Fe(II). In dispersion of NPs with 50 µmol/L Fe²⁺, we detected 49.1 µmol/L Ce ions and in 100 µmol/L Fe²⁺ 71.8 µmol/L. This means that almost equimolar amounts of Ce were released when Fe²⁺ was present. These results were obtained in dispersions under standard atmosphere. When O₂ was displaced by N₂, the amounts of released Ce did not change (data not shown).

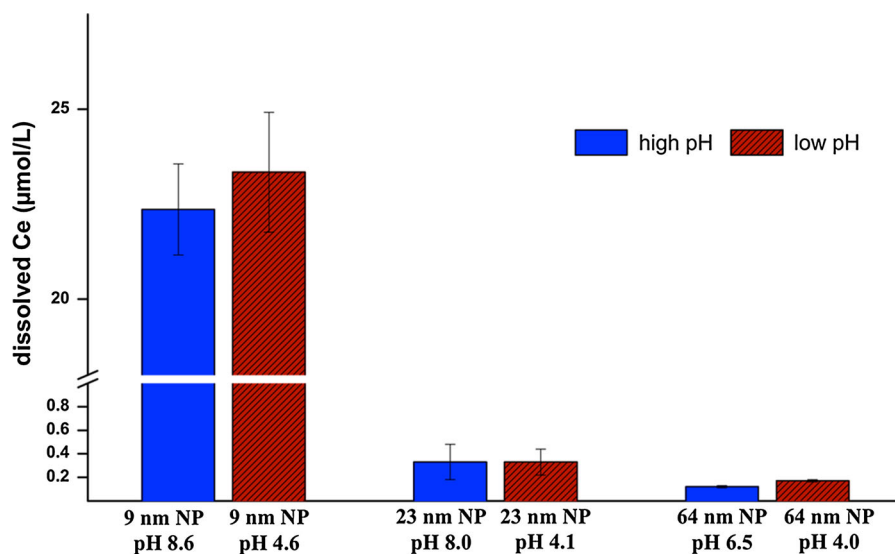
Estimating dissolvable Ce content of NPs by EDTA-complexation

EDTA is a very strong chelating agent that forms strong complexes with trivalent ions and thus enhances the dissolution of oxides (Stumm 1992). For an estimation of the dissolvable Ce, particles were incubated with 50 $\mu\text{mol/L}$ EDTA-acid (Fig. 4). We detected 23 $\mu\text{mol/L}$ Ce with 9-nm NPs, 0.33 $\mu\text{mol/L}$ Ce with 23-nm NPs, and 0.12 $\mu\text{mol/L}$ with the 64-nm NP group, after 7 days of incubation. The results were the same at pH values below 4.6 or above 6. From these results, the percentage of maximum dissolvable Ce was calculated as 4.2 % for 9-nm NPs, 0.06 % for 23-nm NPs, and 0.02 % for 64-nm NPs. The concentration of dissolved Ce after incubation with EDTA-acid is higher compared to incubation with Fe-EDTA, because Fe^{3+} is a competing ion for complexation (Fig 5).

Discussion

Our results show that CeO_2 -NPs are not stable in Hoagland medium but do readily dissolve. We centralize four main factors that influenced the dissolution of CeO_2 -NPs: (i) size of primary particle; (ii) pH of suspension; (iii) extent of agglomeration/aggregation in suspension; and (iv) the presence of chemical binding or reaction partners like phosphate or reducing agents like Fe(II)SO_4 .

Fig. 4 Amount of dissolved Ce from 100 mg/L CeO_2 -NPs in 50 $\mu\text{mol/L}$ EDTA after 7 days of incubation. Break in y-axis: 1–17 $\mu\text{mol/L}$; $n = 3$, error bars indicate standard error over all replicates per dispersion



Concerning factor 1: we found that the smaller the primary particle size was, the more dissolved Ce was released. In our study, the largest NP group (64 nm) was almost insoluble. The smaller particles contained far more Ce(III). Deshpande et al. (2005) demonstrated that small CeO_2 -NPs face a high lattice strain when Ce^{3+} is introduced because of a larger ionic radius compared to Ce^{4+} . This lattice strain increases exponentially when particle size is below 25 nm (Zhang et al. 2004a). Particles below 20 nm, like in our case, have a strong need for lattice relaxation, which is acquired by insertion of oxygen vacancies into the crystal lattice. As a result, even more Ce(III)

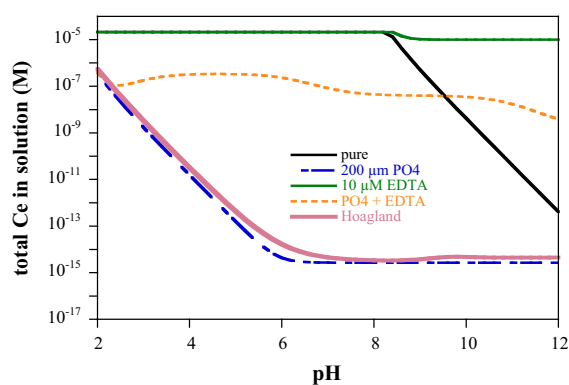


Fig. 5 Solubility of CeOH_3 (total Ce 23 μM) in the presence of different media at the point of thermodynamic equilibrium in pure water, 200 $\mu\text{mol/L}$ PO_4 , 10 $\mu\text{mol/L}$ EDTA, 200 $\mu\text{mol/L}$ PO_4 + 10 $\mu\text{mol/L}$ EDTA, and complete 20 % Hoagland solution

can be introduced into the crystal lattice (Tsunekawa et al. 1999).

In alkaline conditions, the oxidation of Ce^{3+} to Ce^{4+} is preferred, whereas in acidic media, the reduction of Ce is higher (Trinidad et al. 2008). Since Ce^{3+} is the more soluble ion, a decrease in the pH of the dispersion leads to higher Ce release from NPs. Our data confirmed this for all three NP groups in Hoagland medium. Only when the pH was below 5, all three size groups released detectable amounts of Ce.

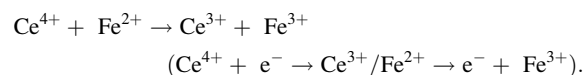
The modeling of $\text{Ce}(\text{OH})_3$ solubility indicated that it is completely soluble up to pH 8 in pure water. Consequently, any Ce^{3+} present on the surface of CeO_2 could be completely dissolved from the NPs up to this pH. The speciation calculation showed the maximum possible dissolved concentration at the point of thermodynamic equilibrium. But dissolution reactions of metal oxides are known to be kinetically slow, especially at higher pH (Stumm 1992) and the thermodynamic equilibrium had presumably not been reached in our samples after 7 days, which may explain the lower dissolved Ce concentrations compared to the modeled values.

The influence of GA as an organic matter representative was well pronounced in Hoagland medium by increasing Ce solubility. There are two possible explanations for these results. Firstly, GA prevents agglomeration and aggregation (Schwabe et al. 2013), thus NP sedimentation is slowed and a large reaction surface is maintained for a longer period, as indicated in this study by the smaller hydrodynamic diameter of all NPs when GA was present. Or secondly, the reactive groups of GA form a complex with Ce in the (III+) oxidation state and keep it stable in solution, similar to EDTA-complexation. Besides natural organic matter, small organic ligands might dissolve ions from CeO_2 -NPs, like oxalate, citrate, and ascorbate (Cervini-Silva et al. 2005).

In the presence of EDTA, equimolar Ce amounts can be solubilized (up to pH 12). This high solubility of Ce^{3+} in the presence of EDTA (log K $\text{Ce}(\text{III})\text{EDTA}$ of 15.93) (Smith 2004) forms the basis for assessing the possible amount of Ce that can be solubilized from CeO_2 -NPs. For 9-nm particles, this fraction was 4.2 %; the fraction of Ce^{3+} calculated from the XPS data was 19 %, thus approximately one fourth of the Ce^{3+} of NPs is close enough to the surface to be solubilized under optimal conditions in the presence of a strong chelating agent. The calculations also suggest

that 200 $\mu\text{mol/L}$ phosphate (the concentration in 20 % Hoagland solution) would result in very low dissolved Ce concentrations above pH 4, caused by the very low solubility of $\text{Ce}(\text{III})\text{PO}_4$. Singh et al. 2011 stated that the redox cycle between Ce^{3+} and Ce^{4+} can be blocked by phosphate since it can trap Ce in the $\text{Ce}(\text{III})$ oxidation state (Singh et al. 2011). Zhang et al. 2012 found $\text{Ce}(\text{III})\text{PO}_4$ formation in hydroponic medium around the roots of cucumber. $\text{Ce}(\text{III})\text{PO}_4$ immobilizes free Ce by precipitation on the particle surface (Zhang et al. 2012). In Hoagland solution, the phosphate content determined the solubility of Ce; the presence of EDTA had only a small influence because it was added as $\text{Fe}(\text{III})\text{EDTA}$ (log K 27.2), which is a more stable complex than $\text{Ce}(\text{III})\text{EDTA}$ (log K 15.9) (Smith 2004). Summarizing our recent findings and the existing literature, $\text{Ce}(\text{III})\text{PO}_4$ is the most likely form into which CeO_2 is transformed.

When $\text{Fe}(\text{II})$ was present as a reducing agent in acidic conditions, the dissolved concentrations of Ce were almost equimolar to the initial $\text{Fe}(\text{II})$, even in the presence of oxygen, which could have driven the oxidation of Ce^{3+} back into Ce^{4+} . These results indicate that once Ce^{3+} is formed on the particle surface, it is released from the particle and subjacent Ce can be reduced, as illustrated in the following chemical equation:



The high solubility of CeO_2 in the presence of Fe^{2+} may have profound implications for the stability of CeO_2 -NPs in natural systems. Free Fe^{2+} can be present in wastewater and occur in sediments under anoxic conditions. CeO_2 -NPs may therefore not be stable in two of the most relevant systems for ENM as determined from modeling studies (Gottschalk et al. 2009), wastewater treatment plants and sediments (Phillips and Lovley 1987).

In summary, our results indicate that the following four reactions are involved in the transformation of CeO_2 -NPs in the environment (Fig. 6). Firstly, the dissolution of Ce in the (III+) oxidation state occurs due to its relatively high solubility (Kakuwa and Matsumoto 2006). Further Ce^{4+} can be reduced to Ce^{3+} and then be released. Reduction occurs predominantly in acidic media and may need an electron donor-like Fe^{2+} . Complexation is another, but very

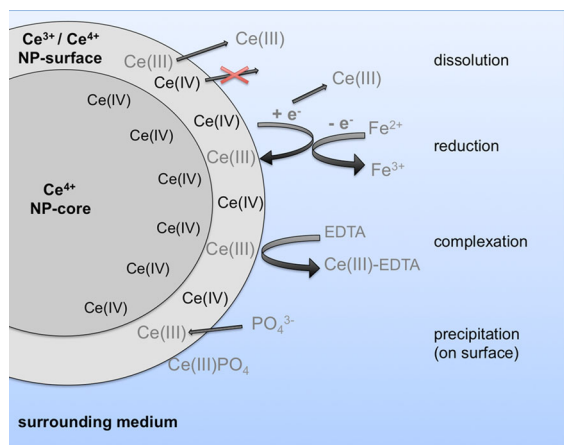


Fig. 6 Scheme of possible chemical reactions taking place on the surface and in the surrounding medium of CeO_2 -NPs when free ions/chelating agents are present

effective, way of transforming CeO_2 -NPs. As we demonstrated in the presence of EDTA, very high amounts of dissolved Ce can be detected. Different anthropogenic chelating agents are found in natural water. In the effluents of wastewater treatment plants, high amounts of EDTA of up to $17 \mu\text{M}$ have been measured (Nowack and VanBriesen 2005). We assume that GA might function in a similar way as EDTA in increasing the soluble Ce fraction.

The fourth important reaction was precipitation. Similar to Baalousha et al. (2010), we found less released Ce when phosphate was present in the media. This indicates that Ce^{3+} is not dissolved from the surface but trapped most likely by CePO_4 formation on the NP surface.

Conclusions

For estimating the fate and effect of nanoparticles in ecosystems, chemical transformation especially dissolution needs to be considered. We emphasize that CeO_2 -NPs, which are often described to be very stable, like in waste incineration plants (Walser et al. 2012), can undergo dissolution or other chemical transformations. To estimate the dissolvable amount of NPs, we suggest the easy method of extracting surface ions via a strong chelating agent (e.g. EDTA).

Acknowledgments We kindly acknowledge Dr. Thomas Bucheli and Alexander Gogos from Agroscope Reckenholz-

Tänikon (ART) for their support in DLS and zeta potential measurements. We thank Dr. Susan Tandy for language corrections. Financial support by the ETH-Zurich (ETHIIRA Project, ETH- 21 08-3) was kindly acknowledged.

References

- Baalousha M, Le Coustumer P, Jones I, Lead JR (2010) Characterisation of structural and surface speciation of representative commercially available cerium oxide nanoparticles. *Environ Chem* 7(4):377–385
- Batley GE, Kirby JK, McLaughlin MJ (2013) Fate and risks of nanomaterials in aquatic and terrestrial environments. *Acc Chem Res* 46(3):854–862
- Brunner TJ, Wick P, Manser P, Spohn P, Grass RN, Limbach LK, Bruinink A, Stark WJ (2006) In vitro cytotoxicity of oxide nanoparticles: comparison to asbestos, silica, and the effect of particle solubility. *Environ Sci Technol* 40:4374–4381
- Cervini-Silva J, Fowle DA, Banfield J (2005) Biogenic dissolution of a soil cerium-phosphate mineral. *Am J Sci* 305(6–8):711–726
- Deshpande S, Patil S, Kuchibhatla SVNT, Seal S (2005) Size dependency variation in lattice parameter and valency states in nanocrystalline cerium oxide. *Appl Phys Lett* 87(13):133113
- Du WC, Sun YY, Ji R, Zhu JG, Wu JC, Guo HY (2011) TiO_2 and ZnO nanoparticles negatively affect wheat growth and soil enzyme activities in agricultural soil. *J Environ Monit* 13(4):822–828
- Gottschalk F, Nowack B (2011) The release of engineered nanomaterials to the environment. *J Environ Monit* 13(5):1145–1155
- Gottschalk F, Sonderer T, Scholz RW, Nowack B (2009) Modeled environmental concentrations of engineered nanomaterials (TiO_2 , ZnO , Ag, CNT, fullerenes) for different regions. *Environ Sci Technol* 43:9216–9222
- Gustafsson JP (2011) Visual MINTEQ version 3.0. <http://www2.lwr.kth.se/English/Oursoftware/vminteq/index.html>. Accessed 1 Oct 2014
- Handy RD, Cornelis G, Fernandes T, Tsyusko O, Decho A, Sabo-Attwood T, Metcalfe C, Steevens JA, Klaine SJ, Koelmans AA, Home N (2012) Ecotoxicity test methods for engineered nanomaterials: practical experiences and recommendations from the bench. *Environ Toxicol Chem* 31(1):15–31
- Hoecke KV, Quik JTK, Mankiewicz-Boczek J, Schampelaere KACD, Elsaesser A, Meeren PVd, Barnes C, McKerr G, Howard CV, Meent DVD, Rydzynski K, Dawson KA, Salvati A, Lesniak A, Lynch I, Silversmit G, Samber BrD, Vincze L, Janssen CR (2009) Fate and effects of CeO_2 nanoparticles in aquatic ecotoxicity tests. *Environ Sci Technol* 43(12):4537–4546. doi:10.1021/es9002444
- Ilan YA, Czapski G, Meisel D (1976) The one-electron transfer redox potentials of free radicals. I. The oxygen/superoxide system. *Biochim Biophys Acta* 430(2):209–224
- Kakuwa Y, Matsumoto R (2006) Cerium negative anomaly just before the Permian and Triassic boundary event—the upward expansion of anoxia in the water column. *Palaeogeogr Palaeoclimatol Palaeoecol* 229(4):335–344

- Li ZM, Zhu FR, Zhang ZB, Ren XJ, Deng H, Zhai LH, Zhang LX (2004) Laser resonance ionization spectroscopy of even-parity autoionization states of cerium atom. *Guang pu xue yu guang pu fen xi =Guang pu* 24(12):1494–1498
- Limbach LK, Li YC, Grass RN, Brunner TJ, Hintermann MA, Muller M, Gunther D, Stark WJ (2005) Oxide nanoparticle uptake in human lung fibroblasts: effects of particle size, agglomeration, and diffusion at low concentrations. *Environ Sci Technol* 39(23):9370–9376. doi:10.1021/es051043o
- Ma XM, Geiser-Lee J, Deng Y, Kolmakov A (2010) Interactions between engineered nanoparticles (ENPs) and plants: phytotoxicity, uptake and accumulation. *Sci Total Environ* 408(16):3053–3061
- Madler L, Stark WJ, Pratsinis SE (2002) Flame-made ceria nanoparticles. *J Mater Res* 17(6):1356–1362
- Miralles P, Church TL, Harris AT (2012) Toxicity, uptake, and translocation of engineered nanomaterials in vascular plants. *Environ Sci Technol* 46(17):9224–9239
- Mueller NC, Nowack B (2008) Exposure modeling of engineered nanoparticles in the environment. *Environ Sci Technol* 42(12):4447–4453
- Nowack B, VanBriesen J (eds) (2005) Chelating agents in the environment. In: *Biogeochemistry of chelating agents*, vol 910. ACS Symposium Series, pp 1–18
- Nowack B, Ranville JF, Diamond S, Gallego-Urrea JA, Metcalfe C, Rose J, Horne N, Koelmans AA, Klaine SJ (2012) Potential scenarios for nanomaterial release and subsequent alteration in the environment. *Environ Toxicol Chem* 31(1):50–59
- Pardo A, Merino MC, Arrabal R, Viejo F, Munoz JA (2007) Ce conversion and electrolysis surface treatments applied to A3xx.x alloys and A3xx.x/SiCp composites. *Appl Surf Sci* 253(6):3334–3344
- Phillips EJP, Lovley DR (1987) Determination of Fe(III) and Fe(II) in Oxalate Extracts of Sediment. *Soil Sci Soc Am J* 51(4):938–941
- Schwabe F, Schulin R, Limbach LK, Stark W, Burge D, Nowack B (2013) Influence of two types of organic matter on interaction of CeO₂ nanoparticles with plants in hydroponic culture. *Chemosphere* 91(4):512–520. doi:10.1016/j.chemosphere.2012.12.025
- Scown TM, Santos EM, Johnston BD, Gaiser B, Baalousha M, Mitov S, Lead JR, Stone V, Fernandes TF, Jepson M, van Aerle R, Tyler CR (2010) Effects of aqueous exposure to silver nanoparticles of different sizes in rainbow trout. *Toxicol Sci* 115(2):521–534. doi:10.1093/toxsci/kfq076
- Seah MP, Dench WA (1979) Quantitative electron spectroscopy of surfaces: a standard data base for electron inelastic mean free paths in solids. *Surf Interface Anal* 1(1):2–11
- Singh S, Dosani T, Karakoti AS, Kumar A, Seal S, Self WT (2011) A phosphate-dependent shift in redox state of cerium oxide nanoparticles and its effects on catalytic properties. *Biomaterials* 32(28):6745–6753. doi:10.1016/j.biomaterials.2011.05.073
- Smith RM, Martell AE (2004) NIST critically selected stability constants of metal complexes, version 2.0. U.S. Department of Commerce, Gaithersburg
- Stark WJ, Madler L, Maciejewski M, Pratsinis SE, Baiker A (2003) Flame synthesis of nanocrystalline ceria-zirconia: effect of carrier liquid. *Chem Commun* 5:588–589. doi:10.1039/b211831a
- Stumm W (1992) *Chemistry of the solid-water interface: processes at the mineral-water and particle-water interface in natural systems*. Wiley, New York
- Tandy S, Schulin R, Nowack B (2006) The influence of EDDS on the uptake of heavy metals in hydroponically grown sunflowers. *Chemosphere* 62(9):1454–1463
- Teterin YA, Teterin AY, Lebedev AM, Utkin IO (1998) The XPS spectra of cerium compounds containing oxygen. *J Electron Spectrosc* 88:275–279
- Thill A, Zeyons O, Spalla O, Chauvat F, Rose J, Auffan M, Flank AM (2006) Cytotoxicity of CeO₂ nanoparticles for *Escherichia coli*. Physico-chemical insight of the cytotoxicity mechanism. *Environ Sci Technol* 40:6151–6156
- Trinidad P, de Leon CP, Walsh FC (2008) The use of electrolyte redox potential to monitor the Ce(IV)/Ce(III) couple. *J Environ Manage* 88(4):1417–1425
- Trovarelli A, Boaro M, Rocchini E, de Leitenburg C, Dolcetti G (2001) Some recent developments in the characterization of ceria-based catalysts. *J Alloy Compd* 323:584–591
- Tsunekawa S, Sahara R, Kawazoe Y, Ishikawa K (1999) Lattice relaxation of monosize CeO_{2-x} nanocrystalline particles. *Appl Surf Sci* 152(1–2):53–56
- Walser T, Limbach LK, Brogioli R, Erismann E, Flamigni L, Hattendorf B, Juchli M, Krumeich F, Ludwig C, Prikopsky K, Rossier M, Saner D, Sigg A, Hellweg S, Gunther D, Stark WJ (2012) Persistence of engineered nanoparticles in a municipal solid-waste incineration plant. *Nat Nanotechnol* 7(8):520–524
- Wang Z, Xie X, Zhao J, Liu X, Feng W, White JC, Xing B (2012) Xylem- and phloem-based transport of CuO nanoparticles in maize (*Zea mays* L.). *Environ Sci Technol* 46(8):4434–4441. doi:10.1021/es204212z
- Wu LJ, Wiesmann HJ, Moodenbaugh AR, Klie RF, Zhu YM, Welch DO, Suenaga M (2004) Oxidation state and lattice expansion of CeO_{2-x} nanoparticles as a function of particle size. *Phys Rev B* 69(12):125415
- Zhang F, Jin Q, Chan SW (2004a) Ceria nanoparticles: size, size distribution, and shape. *J Appl Phys* 95(8):4319–4326
- Zhang F, Wang P, Koberstein J, Khalid S, Chan SW (2004b) Cerium oxidation state in ceria nanoparticles studied with X-ray photoelectron spectroscopy and absorption near edge spectroscopy. *Surf Sci* 563(1–3):74–82
- Zhang ZY, He X, Zhang HF, Ma YH, Zhang P, Ding YY, Zhao YL (2011) Uptake and distribution of ceria nanoparticles in cucumber plants. *Metallomics* 3(8):816–822
- Zhang P, Ma YH, Zhang ZY, He X, Zhang J, Guo Z, Tai RZ, Zhao YL, Chai ZF (2012) Biotransformation of ceria nanoparticles in cucumber plants. *ACS Nano* 6(11):9943–9950
- Zhao LJ, Peralta-Videa JR, Varela-Ramirez A, Castillo-Michel H, Li CQ, Zhang JY, Aguilera RJ, Keller AA, Gardea-Torresdey JL (2012) Effect of surface coating and organic matter on the uptake of CeO₂ NPs by corn plants grown in soil: insight into the uptake mechanism. *J Hazard Mater* 225:131–138
- Zhu H, Han J, Xiao JQ, Jin Y (2008) Uptake, translocation, and accumulation of manufactured iron oxide nanoparticles by pumpkin plants. *J Environ Monit* 10:713–717