

Claudio Soravia
Celia D. DeLozier
Zuzana Dobbie
Claudine Rey Berthod
Eviano Arrigoni
Marie-Anne Bründler
Jean-Louis Blouin
William D. Foulkes
Pierre Hutter

Double frameshift mutations in *APC* and *MSH2* in the same individual

Accepted: 15 March 2005
Published online: 5 April 2005
© Springer-Verlag 2005

C. Soravia
Clinic of Visceral Surgery,
Geneva University Hospital,
Geneva, Switzerland

C. D. DeLozier · J.-L. Blouin
Division of Medical Genetics,
Geneva University Hospital,
Geneva, Switzerland

Z. Dobbie
Division of Medical Genetics,
Department of Clinical and
Biological Sciences,
Basel, Switzerland

C. R. Berthod · P. Hutter
Genetics Unit, ICHV,
Sion, Switzerland

E. Arrigoni
Gastroenterology Practice,
Geneva, Switzerland

M.-A. Bründler
Division of Clinical Pathology,
Geneva University Hospital,
Geneva, Switzerland

W. D. Foulkes
Departments of Medicine,
Oncology and Human Genetics,
McGill University,
Montreal, Québec, Canada

C. Soravia (✉)
Route de Chêne 11,
1207 Geneva, Switzerland
e-mail: csoravia@hin.ch
Tel.: +41-22-7863292
Fax: +41-22-7863285

Abstract Heterozygous germline DNA mismatch repair gene mutations are typically associated with HNPCC. Here we report the case of a proband whose father was known for familial adenomatous polyposis. The number of polyps (<10) was not typical of polyposis, therefore the diagnosis of HNPCC was entertained. Microsatellite instability analyses were performed on peripheral blood and biopsy of a right-sided dysplastic adenoma. The tumour tissue showed high-grade instability and subsequently immunohistochemistry showed that neither *MSH2* nor *MSH6* proteins were expressed in tumour cells. Prophylactic colectomy was performed and an adenocarcinoma developing within the adenoma was diagnosed (pT1N0). Genomic DNA analysis revealed a novel mutation in *MSH2* as: a frameshift mutation in exon 7 (c.1,191_1,192dupG). Both parents of the proband were analyzed for *MSH2* and *APC* mutations, and in the father a truncating mutation in exon 15 of *APC* was identified as del3471-3473GAGA. This mutation was found to be present in the proband. His mother was found to bear the *MSH2* exon 7 mutation. At follow-up, the proband was diagnosed with fundic, antral and duodenal adenomas (one fundic adenoma showed low-grade dysplasia). Several tubular rectal adenomas with low-grade dysplasia were excised. The patient later developed an intra-abdominal desmoid tumour.

Keywords Hereditary colorectal cancer · Mismatch repair · Familial adenomatous polyposis · Hereditary nonpolyposis colorectal cancer · Microsatellite instability

Abbreviations HNPCC: hereditary nonpolyposis colorectal cancer · MMR: mismatch repair · FAP: familial adenomatous polyposis · AFAP: attenuated familial adenomatous polyposis · CRC: colorectal cancer · MSI: microsatellite instability · IHC: immunohistochemistry · ACF: aberrant crypt foci · NF1: neurofibromatosis type 1

Introduction

Among the known genetic predispositions to cancer syndromes, two confer a high risk of colorectal cancer (CRC). Familial adenomatous polyposis (FAP), generally due to mutations in the *APC* gene on chromosome 5, is responsible for approximately 1% of all CRC. Mutations in at least four mismatch repair (MMR) genes, *MLH1*, *MSH2*, *MSH6* and *PMS2*, result in Lynch syndrome, or HNPCC (hereditary nonpolyposis colorectal cancer); together these MMR gene mutations account for 3–5% of all CRC [1–3]. Given the relatively high incidence of these conditions, one would expect an occasional individual to be homozygous for the two *APC* or two MMR mutations, or to be a carrier of compound heterozygous germline mutations in *APC* and MMR genes [4–9] Table 1.

To our knowledge however, the present case is the first patient observed with concomitant inheritance of pathogenic frameshift mutations in *APC* and *MSH2*.

We report here a complete description of this family with 5-year follow-up of the proband. As he presented fewer polyps than expected at age 25 in a patient with FAP: the differential diagnosis was between HNPCC and attenuated FAP (AFAP) [10, 11].

Case report

The proband, a 24-year-old man, consulted because of a history of FAP in his father and at least seven other members of his southern Italian family (Fig. 1). His father's medical record was not available when this study begun and no genetic testing had been performed. Five polyps were detected on ileocoloscopy, two in the right colon and three in the sigmoid; four of the polyps were adenomas, one of which was dysplastic.

Table 1 Summary of reported families with *APC* and MMR germline mutations

| References | Genes involved | Clinical findings |
|------------------------------|-----------------------------|--|
| Scheenstra et al. (2003) [9] | <i>APC</i> , <i>MLH1</i> | 10-year old with symptomatic FAP, rapid progression of adenomas |
| Yuan et al. (1998) [7] | <i>APC</i> , <i>MLH1</i> | <i>APC</i> polymorphism I1307K; family not of Jewish origin; does not segregate with HNPCC disease phenotype |
| Yuan et al. (1999) [8] | <i>APC</i> , <i>MSH2</i> | <i>APC</i> polymorphism I1307K; Ashkenazi Jewish individual; variant does not segregate with HNPCC disease phenotype |
| Present case (2004) | <i>APC</i> , <i>MSH2</i> | Right-sided polyps (AFAP), but early age of CRC (24), gastric and rectal adenomas with dysplasia |

As the number of polyps was not typical of polyposis, the diagnosis of HNPCC was entertained. Microsatellite instability analysis was performed on DNA from dysplastic adenoma and from peripheral blood. The tumour tissue showed high-grade instability (MSI-H) at all four markers tested: BAT25, BAT26, D2S123 and D5S346. IHC on four DNA mismatch repair proteins was also performed. *MSH2* and *MSH6* were not expressed in tumor cells, whereas *MLH1*/*PMS2* expression was conserved.

Genetic analysis started with *MSH2* and bidirectional sequencing of the exons and the intron–exon junctions using amplification products obtained by PCR from genomic DNA. A novel mutation was identified in *MSH2*: insertion of a G in codon 398, in exon 7 at position 1,191 (c.1,191_1,192dupG), had created a premature termination 19 codons downstream.

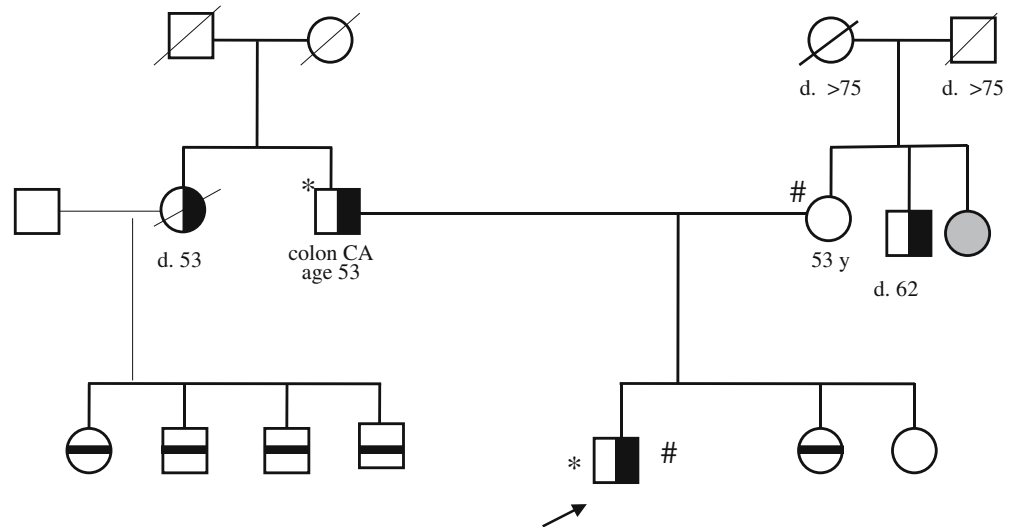
The patient opted for prophylactic surgery at age 25 and underwent total colectomy with ileo–rectal anastomosis. In the prophylactic colectomy specimen, a small well-differentiated adenocarcinoma (pT1N0), developing from a tubulo–villous adenoma was found in the right colon, along with “multiple” flat adenomas and associated aberrant crypts. No intra-abdominal desmoid tumour was found at this time.

Over the last 4 years, the patient has experienced numerous complications: soon after colectomy, he had frequent episodes of gastro–esophageal reflux which were temporarily lessened by a proton-pump inhibitor (pantoprazol). At the age of 25 oeso–gastroduodenoscopy showed chronic gastritis (negative for *Helicobacter pylori*), hiatal hernia and approximately 30 fundic polyps. Out of three adenomas excised from duodenum, antrum, and gastric fundus, the latter showed low-grade dysplasia. Rectoscopies were carried out at 3-month intervals over the following year, and showed the presence of two to nine polyps, at least one of which was a tubular adenoma with mild dysplasia. The patient was placed on rofecoxib in an attempt to reduce polyp development, but micro-perforations led to peritonitis and emergency surgery. On this occasion, a mesenteric desmoid tumour was diagnosed from biopsy. To eliminate the risk of additional rectal cancer a proctectomy with endo-anal mucosectomy, and implementation of a pelvic pouch procedure was proposed, but was declined by the patient.

Bi-annual rectoscopy and esogastroduodenoscopy are currently carried out as surveillance. At his most recent examination (at age 28), the proband presented with fewer than ten rectal polyps (one serrated adenoma) but presented multiple gastric fundic polyps (excised polyp was adenoma with low-grade dysplasia).

At the time of their son's surgery for colectomy both parents consulted. As already mentioned, the father presented with familial adenomatous polyposis (FAP); he had undergone colectomy at age 53 after FAP with colon carcinoma was diagnosed. An *APC* gene mutation search was initiated in our laboratory. PTT detected a truncating mutation in exon 15, then characterized as c.del3471_3474-GAGA. This frameshift mutation creates a premature stop

Fig. 1 Proband's family pedigree with *APC* and *MSH2* germline mutations. Pedigree of the family segregating an *APC* mutation (father of proband) and *MSH2* mutation (mother of proband). Half-filled symbols indicate colon cancer, filled gray symbols endometrial cancer and horizontal lines clinical polyposis coli. Proven *APC* mutation carriers have an * and proven *MSH2* mutation carriers an #. The arrow indicates the proband



codon six codons downstream. His son (the proband), carrying the *MSH2* exon 7 mutation, was found to have inherited the *APC* exon 15 mutation from his father. The *MSH2* mutation was absent in the father.

We then analyzed the proband's 53-year-old mother, who had no significant medical history but had not undergone colonoscopy, carries the *MSH2* mutation. Clinical screening for digestive and gynecological pathologies was recommended. Of her seven siblings, a brother reportedly died of colon cancer in his fifties and a sister of uterine cancer at age 62. Genetic testing was offered to all at-risk family members.

Discussion

At the age of 25, the proband's clinical presentation was more suggestive of HNPCC or possibly AFAP than of classical FAP, on both clinical and histological grounds. This observation, along with the lack of confirmatory medical records about his father, led us to perform screening tests (MSI, IHC) for HNPCC. The proband was found to harbour two germline mutations predisposing to CRC: an *APC* exon 15 frameshift mutation inherited from his father and a frameshift mutation in *MSH2*, inherited from his mother.

Upon further evaluation, the proband showed clinical characteristics of both disorders: desmoid tumours are indicative of FAP, and the small number of polyps in the proximal colon (less than ten) was suggestive of AFAP or HNPCC. His clinical course has, however, been relatively severe, with an adenocarcinoma of the right colon diagnosed at age 25, and the postoperative development of a mesenteric desmoid tumour. Desmoid tumours occur in 80% of individuals with FAP who undergo prophylactic colon surgery [14]. Several studies have demonstrated an increased risk for desmoid tumours, in patients with *APC* mutations occurring between codons 1,310 and 2,011 [12–14]. In our

patient, the *APC* mutation is located at codon 1,157 and there is no family history of desmoids in this kindred.

Gastric adenoma and/or cancer has been described in both AFAP and HNPCC [15–17]. *Helicobacter pylori* infection may induce atrophic gastritis in FAP patients and trigger adenoma development, but our patient tested negative for *Helicobacter pylori*. As gastric cancer/dysplasia is part of the HNPCC tumour spectrum, *MLH1* and *MSH2* mutations may influence gastric cancer progression.

A more aggressive clinical course has been reported in the handful of reported patients with both *APC* and a second predisposing mutation in a different gene. Zajac and colleagues [18] reported an 18-year-old with inherited FAP and >1,000 polyps. The familial mutation, upstream from the mutation cluster region in the *APC* gene, had not been associated with a particularly severe phenotype in this family. Because of the proband's clinical severity a search for additional mutations was undertaken and a germline *p53* mutation identified. The authors hypothesized that the mother, who died at age 28 of uterine carcinoma, may have also had double mutations.

The only other patient reported to harbor concomitant mutations in *APC* and a MMR gene was a 10-year-old child with symptomatic polyposis; the *APC* mutation was a maternally-inherited prevalent frameshift in exon 15, and the paternally-inherited MMR mutation was a splice site mutation in *MLH1* [9]. The authors hypothesized that the rapid progression to high-grade adenomas in this patient was due to the influence of the *MLH1* mutation; high-grade dysplasia was associated with loss of *MLH1* expression by IHC criteria. Surely, the severe phenotype may simply be a reflection of the clinical variability seen in both FAP and HNPCC.

It is worth noting that in mice that are compound heterozygotes for *APC* and *MLH1* germline mutations, cancer phenotypes are also enhanced [19, 20].

Two families have been described [7, 8] with an MMR gene mutation plus the *APC* missense variant I1307K. The latter change, present in 6% of the Jewish population, does not result in FAP, but in some individuals appears to predispose to colorectal polyps, as a possible result of hypermutability of the altered region of the *APC* gene; the relative risk of CRC with the I1307K polymorphism has been reported as 1.5–2.0 [21]. In the first of the two families, a French Canadian CRC kindred with an *MLH1* truncating mutation, there was no clear association between the *APC* I1307K polymorphism and the development of cancer [7]. The same was true in a second such family, of Ashkenazi origin, which fulfilled the Amsterdam criteria for HNPCC. Affected individuals had a missense mutation of *MSH2*, which was later proven to be pathogenic [22]; however, presence or absence of the *APC* I1307K genotype did not correlate with disease. Thus, in these two families, one clear mutation (in a MMR gene which correlated with the HNPCC phenotype) was defined, whereas the *APC* polymorphism was transmitted independently of the CRC phenotype.

Kinzler and Vogelstein [23] proposed that the *APC* gene serves as the “gatekeeper” of epithelial proliferation, thus exerting a major influence on initiation of the neoplastic process. By contrast, the MMR mutations underlying HNPCC alter “caretaker” genes, and would accelerate carcinogenesis, once the initiation step has occurred. However, the

distinction between gatekeeper and caretaker roles has recently become somewhat blurred, as the mutation of *APC* could not be shown to be a compulsory step in all CRC. Indeed, a significant proportion of colorectal cancer may be initiated through the epigenetic silencing of alternative genes implicated in apoptosis and mismatch repair [24]. In dysplastic microscopic epithelial lesions referred to as aberrant crypt foci (ACF), flat tubular adenomas, and polypoid tubular adenomas, the frequency of *APC* mutations is reported to be 0, 7 and 36%, respectively [25]. Moreover, genomic instability has actually been detected in ACF. These observations led to two contrasting hypotheses: a “dysplasia ACF–adenoma–carcinoma sequence”, or a “heteroplastic ACF–adenoma–carcinoma sequence”. These observations may suggest the *APC* mutation consistently triggers adenoma formation before the *MSH2* mutation participates in tumorigenesis through MSI. Molecular analysis of ACF or the demonstration that very early adenomas show abnormal expression of beta-catenin [26], should help to clarify this issue.

Acknowledgements This work was supported by a grant from the Geneva University Hospitals (PRD-01-2-23 to JLB and CDD). PH was supported by the Fonds National Suisse de la Recherche Scientifique (No 3138-051088) and by la Recherche Suisse Contre le Cancer (No AKT1446).

References

1. Altonen LA, Salovaara R, Kristo P et al (1998) Incidence of hereditary nonpolyposis colorectal cancer and the feasibility of molecular screening for the disease. *N Engl J Med* 338:1481–1487
2. Lynch HT, de la Chapelle A (2003) Hereditary colorectal cancer. *N Engl J Med* 348:919–932
3. Soravia C, Cohen Z (2005) Familial adenomatous polyposis. In: Fazio VW, Church JM, Delaney CP (eds) *Current therapy in colon and rectal surgery*. Elsevier Mosby, Philadelphia, pp 349–353
4. Kariola R, Otway R, Lonnqvist KE et al (2003) Two mismatch repair gene mutations found in a colon cancer patient—which one is pathogenic? *Hum Genet* 112(2):105–109
5. Whiteside D, McLeod R, Graham G et al (2002) A homozygous germ-line mutation in the human *MSH2* gene predisposes to hematological malignancy and multiple café-au-lait spots. *Cancer Res* 62:359–362
6. Gallinger S, Aronson M, Shayan K et al (2004) Gastrointestinal cancers and neurofibromatosis type 1 features in children with a germline homozygous *MLH1* mutation. *Gastroenterology* 126:576–585
7. Yuan ZQ, Kasprzak L, Gordon PH, Pinsky L, Foulkes WD (1998) I1307K *APC* and h*MLH1* mutations in a non-Jewish family with hereditary nonpolyposis colorectal cancer. *Clin Genet* 54:369–370
8. Yuan ZQ, Wong N, Foulkes WD et al (1999) A missense mutation in both h*MSH2* and *APC* in an Ashkenazi Jewish HNPCC kindred: implications for clinical screening. *J Med Genet* 36:790–793
9. Scheenstra R, Rijcken FE, Koornstra JJ et al (2003) Rapidly progressive adenomatous polyposis in a patient with germline mutations in both the *APC* and *MLH1* genes: the worst of two worlds. *Gut* 52:898–899
10. Soravia C, Berk T, Madlensky L et al (1998) Genotype–phenotype correlations in attenuated adenomatous polyposis coli. *Am J Hum Genet* 62:1290–1301
11. Cao Y, Pieretti M, Marshall J, Khattar NH, Chen B, Kam-Morgan L, Lynch H (2002) Challenge in the differentiation between attenuated familial adenomatous polyposis and hereditary nonpolyposis colorectal cancer: case report with review of the literature. *Am J Gastroenterol* 97(7):182
12. Soravia C, Berk T, McLeod RS, Cohen Z (2000) Desmoid disease in patients with familial adenomatous polyposis (FAP). *Dis Colon Rectum* 43:363–369
13. Davies DR, Armstrong JG, Thakker N et al (1995) Severe Gardner syndrome in families with mutations restricted to a specific region of the *APC* gene. *Am J Hum Genet* 57:1151–1158
14. Caspari R, Olschwang S, Friedl W et al (1995) Familial adenomatous polyposis: desmoid tumours and lack of ophthalmic lesions (CHRPE) associated with *APC* mutations beyond codon 1444. *Hum Mol Genet* 4:337–340

15. Bertario L, Russo A, Sala P et al (2003) Multiple approach to the exploration of genotype–phenotype correlations in familial adenomatous polyposis. *J Clin Oncol* 21:1698–1707
16. Aarnio M, Salovaara R, Aaltonen LA, Mecklin JP, Järvinen H (1997) Features of gastric cancer in hereditary non-polyposis colorectal cancer syndrome. *Int J Cancer* 74:551–555
17. Nakamura S, Matsumoto T, Kobori Y, Iida M (2002) Impact of *Helicobacter pylori* infection and mucosal atrophy on gastric lesions in patients with familial adenomatous polyposis. *Gut* 51:485–489
18. Zajac V, Tompka M, Ilencikova D, Majek P, Stevurkova V, Kirchoff T (2000) A double germline mutations in the APC and p53 genes. *Neoplasma* 47 (6):335–341
19. Edelmann W, Yang K, Kuraguchi M et al (1999) Tumorigenesis in Mlh1 and Mlh1/Apc1638N mutant mice. *Cancer Res* 59:1301–1307
20. Shoemaker AR, Haigis KM, Baker SM et al (2000) Mlh1 deficiency enhances several phenotypes of Apc(Min)/+ mice. *Oncogene* 19:2774–2779
21. Niell BL, Long JC, Rennert G, Gruber SB (2003) Genetic anthropology of the colorectal cancer-susceptibility allele APC I1307K: evidence of genetic drift within the Ashkenazim. *Am J Hum Genet* 73:1250–1260
22. Foulkes WD, Thiffault I, Gruber SB et al (2002) The founder mutation MSH2*1906G->C is an important cause of hereditary nonpolyposis colorectal cancer in the Ashkenazi Jewish population. *Am J Hum Genet* 71:1395–1412
23. Kinzler KW, Vogelstein B (1996) Lessons from hereditary colorectal cancer. *Cell* 87:159–170
24. Yuen ST, Chan TL, Ho JW et al (2002) Germline, somatic and epigenetic events underlying mismatch repair deficiency in colorectal and HNPCC-related cancers. *Oncogene* 21 (49):7585–7592
25. Jass JJ, Whitehall VLJ, Young J, Legget BA (2002) Emerging concepts in colorectal neoplasia. *Gastroenterology* 123:862–876
26. Mirabelli-Primdahl L, Gryfe R, Kim H et al (1999) Beta-catenin mutations are specific for colorectal carcinomas with microsatellite instability but occur in endometrial carcinomas irrespective of mutator pathway. *Cancer Res* 59:3346–3351