

Habitat suitability models for the conservation of thermophilic grasshoppers and bush crickets—simple or complex?

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Abstract One goal of conservation biology is the assessment of effects of land use change on species distribution. One approach for identifying the factors, which determine habitat suitability for a species are statistical habitat distribution models. These models are quantitative and can be used for predictions in management scenarios. However, they often have one major shortcoming, which is their complexity. This means that they need several, often costly-to-determine parameters for predictions of species occurrence. We first used habitat suitability models to investigate and determine habitat preferences of three different Orthoptera species. Second, we compared the predictive powers of simple habitat suitability models considering only the ‘habitat type’ as predictor with more complex models taking different habitat factors into account. We found that the habitat type is the most reliable and robust factor, which determines the occurrence of the species studied. Thus, analyses of habitat suitability can easily be carried out on the basis of existing vegetation maps for the conservation of the three species under study. Our results can serve as a basis for the estimation of spatio-temporal distribution

and survival probabilities of the species studied and might also be valuable for other species living in dry grasslands.

Keywords Conservation · Habitat selection modelling · Dry grassland · Semi-arid grassland · Model simplicity

Introduction

Anthropogenic land use has contributed to a diversification of the landscape (Settele 1998), while increasing land use has created new habitats for animal and plant species (Huston 1994; Mühlenberg et al. 1996). In Central Europe, species of nature conservation concern, as well as high species diversity in general have been mainly found in extensively managed areas (Kull and Zobel 1991; Bignal and McCracken 1996). The landscape pattern has remained static, since most areas have been utilised in the same way over many years and even centuries. However, due to increased economic pressure, extensively managed areas are nowadays either abandoned and lie fallow, or are fertilized and intensively used (Mühlenberg et al. 1996). In both cases, rare and protected plant and animal species become extinct due to natural succession or increased disturbance (Fuller 1987; Vos and Zonnefeld 1993; Beaufoy et al. 1994; Poschlod et al. 1996).

Within Central Europe these problems apply particularly to dry grasslands such as the ones in the nature reserve ‘Hohe Wann’ in Central Germany, which have only a low agricultural productivity (Van Dijk 1991; Poschlod et al. 1996). On the one hand,

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these grasslands require some level of disturbance to increase small scale environmental heterogeneity and thus species diversity (Huston 1994; McConnaughay and Bazzaz 1987; Jacquemyn et al. 2003). On the other hand, disturbance should not exceed a certain level, with increased management intensity leading to species diversity decline (Kruess and Tscharntke 2002).

For conservation of these areas, different management regimes have been suggested, such as goat and/or cattle grazing, rototilling, burning and mowing (e.g. Schreiber 1977; Bakker 1989; Bobbink and Willems 1993; Kahmen et al. 2002; Kleyer et al. 2002; Redecker et al. 2002). Different management regimes with different return intervals result in a landscape consisting of a mosaic of different habitat patches, with habitat quality constantly changing over time.

For the protection and conservation of insect populations in such dynamic, fragmented landscapes, it is important to know at what successional time a patch is of ideal, or at least acceptable suitability for a specific species or species assemblage. To predict which successional stages are suitable for specific species, reliable information on species-specific habitat requirements are needed. In general, such information is a critical prerequisite for the choice of protected areas, the design of management strategies, and the assessment of possible effects of various land-use changes on the survival of plant and animal species (Fielding and Haworth 1995; Oppel et al. 2004).

In recent years, statistically derived habitat suitability models have become a common tool for the estimation of critical factors, which determine habitat suitability and habitat selection by a species (Lindenmayer et al. 1991; Pearce et al. 1994; Guisan and Zimmermann 2000; Rushton et al. 2004). Such models formalize the relationship between the occurrence of a species and characteristics of a site (Guisan and Zimmermann 2000; Austin 2002) and may be a cost effective alternative to monitoring (Owen 1989; Fraser 1998).

The development of multi-parameter logistic regression models representing several facets of the realised niche investigates habitat preferences of the (insect) species under study in great detail. Unfortunately, such detailed information is not usually available in practical conservation biology and requires enormous amounts of time-consuming field work. In addition, predictors may vary depending on spatial scale, so that different parameters for different questions need to be determined. Thus, for

applied conservation biology, one would like to know a few, easy-to-measure, integrative parameters for the prediction of habitat suitability. Such parameters should ideally be independent of spatial scale. Examples are the management type as well as the habitat type, which are often the only landscape-wide information readily available. Thereby, the habitat type describes the type of vegetation that is typically found in an area, such as dry grassland or forest. Thus, it comprises many aspects of the realised niche of (insect) species living in such areas with regard to abiotic and biotic conditions as well as disturbance regimes.

In contrast to butterflies and some other insect species, grasshoppers and bush crickets are generally regarded as food generalists (i.e. polyphagous to omnivorous, Detzel 1998) and are therefore not usually limited by food resources in natural habitats. Habitat capacity is therefore likely to be determined by other factors such as oviposition sites or ambient temperature. In temperate zones most grasshoppers and bush crickets occur in dry and open habitats with the highest diversity in warm lowland habitats (Detzel 1998). These areas are the focus of many nature conservation efforts in Europe (Poschlod and Schumacher 1998; Pykälä 2003). In open grasslands, grasshoppers and bush crickets utilise different structures during their life cycle such as long lawn structures for food or shelter and short lawn areas with increased temperature for egg development. Grasshoppers and bush crickets are therefore good indicators of structural heterogeneity. Consequently, their habitat requirements cover the habitats of a variety of different other animal species in open grasslands.

In this study, we develop statistical habitat suitability models for the two bush cricket species *Platycleis albopunctata* and *Metrioptera bicolor* (PHILIPPI 1796; Orthoptera: Tettigoniidae) as well as for the grasshopper *Stenobothrus lineatus* (PANZER 1796) (Orthoptera: Acrididae). All three species are typically found on dry grassland. For the development of the models we first use readily available information such as the habitat type. Secondly, we select biotic and abiotic site parameters which are relevant to the habitat preferences of the three species and thus develop more complex multiple models that include specific plot parameters, like vegetation, topography or soil characteristics. Finally, the predictive performances of both types of models are compared to quantify the trade-off between practical applicability and conservation issues.

Methods

The species

From literature a general ‘expert opinion’ on the habitat requirements of the three species can be formed. However, these opinions have not yet been confirmed by thorough field studies of the kind presented in this article (for another method see Lele and Allen 2006). All three species studied are at their northern distribution limit in Germany and thus, might chose distinct habitats within their core distribution area. Current knowledge on species habitat requirements, threat status and distribution can be summarised as follows:

Stenobothrus lineatus

The stripe-winged grasshopper *Stenobothrus lineatus* (PANZER 1796) (Orthoptera: Acrididae) is a medium to large-sized grasshopper species (body length: 15–26 mm). It is thermophilic and xerophilic and inhabits arid and semi-arid grasslands as well as broom heath, juniper heath, and short lawn edges of woods. Sheep-grazed areas and short vegetation structures are its preferred habitat elements (Detzel 1998). *S. lineatus* originates from Siberia and has a euro-asiatic distribution (Detzel 1998; Maas et al. 2002). Populations are reported from France, Southern England, Spain, Italy, Poland and the CIS-countries (Maas et al. 2002). *S. lineatus* is common in South- and Mid-Germany but in North-/Northwest-Germany particularly in the coastal regions, this species only occurs in small populations. Here we find the northern distribution range of the species in Germany. *S. lineatus* is not mentioned in the Red List of Germany (1997), but has been given a ‘near threatened’ status and classified as vulnerable according to Red List of Bavaria, the study region (Bayerisches Landesamt für Umweltschutz 2003).

Metriopectera bicolor

The two-colored bush cricket *Metriopectera bicolor* (PHILIPPI 1796; Orthoptera: Tettigoniidae) is medium-sized (body length: 15–18 mm), thermophilic and xerophilic, and mainly inhabits dry grasslands. As it orientates towards vertical structures, the species prefers longlawn biotopes. *M. bicolor* can also be found on juniper heath, poor grasslands, semi-arid and sandy grasslands (Detzel 1998). Kindvall and Ahlen (1992) describe the species as sedentary as it does not often leave its native habitat patches. In the Red List of

Germany (1997) *M. bicolor* is not mentioned and can therefore be regarded as not endangered or threatened. Its conservation status according to Red List of Bavaria (Bayerisches Landesamt für Umweltschutz 2003) is vulnerable. The northern distribution range of *M. bicolor* in Germany runs from Northern Rhineland-Palatinate via South-Hesse, Thuringia to Brandenburg, where only scattered distributed populations can be found. The overall distribution range of *M. bicolor* is from the Ural to France and from Southern Sweden to Northern Italy (Maas et al. 2002).

Platycleis albopunctata

The grey bush cricket *Platycleis albopunctata* (GOEZE 1778; Orthoptera: Tettigoniidae) is a medium- to large-sized bush cricket species (body length: 18–22 mm). It is classified as a thermophilic and xerophilic species (Harz 1969; Ingrisch and Köhler 1998), which inhabits dry locations, especially mesobromion (Detzel 1998). Open soil, sparse vegetation and fringes are its preferred habitat elements. *P. albopunctata* is rated as ‘near threatened’ for the Red List of Germany (1997). In Bavaria it is considered ‘vulnerable’ (Ingrisch and Köhler 1998, Bayerisches Landesamt für Umweltschutz 2003). *P. albopunctata* is mainly distributed in West and Central Europe. The eastern distribution border runs from Silesia to the Northern range of the Alps in Lower Austria. In Western Europe the species is distributed across Portugal, Spain, France and Southern England. The most northern populations have been reported in Southern Scandinavia. Its distribution in Germany is very scattered and the Atlantic regions of Northwest Germany are not colonized. In South and Eastern Germany the species is more abundant in the warmer regions. In Bavaria the species is restricted to the north.

Field work

The study was conducted in August and September 2001 and 2002 in the nature reserve ‘Hohe Wann’ in Northern-Bavaria, Germany (latitude 50°03′, longitude 10°35′). The study area is characterised by a patchwork of vegetation caused by the geological and geomorphological heterogeneity of the area. Small-scale microclimatic differences are caused by differences in inclination, insolation and land use. The most obvious characteristic of the nature reserve is an abundance of dry grassland, formerly used as vine yards (Elsner 1994). These patches are separated by small agricultural fields of different use resulting in a

patchy mosaic of different habitat types. The whole area covers approximately 10 km in NS-direction and 4 km in EW-direction (Binzenhöfer et al. 2005; Rudner et al. 2005).

Incidence of the grasshopper and bush cricket species was recorded on 146 experimental sites selected by stratified random sampling across the ten main habitat types occurring in the region (Table 1). In this study the ‘habitat type’ describes a landscape unit within the studied landscape characterised by its typical vegetation composition, e.g. forest or dry grassland. To increase the resolution of our approach (Vaughan and Ormerod 2003), we sampled habitats with uncertain status regarding the species’ occurrence more intensively (Table 2).

To determine the main habitat types (Table 2) in the area we used a Geographic Information System (ESRI™ ArcView 3.2). The distance between two experimental sites was at least 30 m. In the field, we characterised each site by the vegetation structure of a randomly chosen 1 m² plot. For the analysis of micro-structural preference of the grasshoppers and bush crickets vegetation structure was also recorded in 1 m² plots surrounding the point where individuals of the species under study were found. Vegetation structure analysis included estimates of (i) horizontal plant cover in 10, 20, 30, 40, 50 and 60 cm heights, (ii) vertical plant cover of moss-, herb- and grass-layers and (iii) mean vegetation height (cf. Sundermeier 1999). Additionally we recorded the habitat type, the current management regime, the inclination and exposition of the plots. To determine the impact of ‘landscape’ factors such as solar radiation, slope, soil type and geology, these factors were calculated in a digital terrain model and a landscape model (Schröder et al. 2004; Rudner et al. 2005) and included into the analyses.

For the determination of grasshopper and bush cricket incidence we carried out transect sampling (inter-transect distance = 1.5 m) on the experimental sites (15 × 15 m). The census was terminated either (i) as soon as a specimen was found or (ii) after a maximum of 20 min of sampling. As the activity of grasshoppers and bush crickets strongly depends on weather conditions, we carried out censuses only during ‘good’ weather conditions (i.e. sunshine, cloud cover <3/8; air temperature >17°C; wind speed <4 m/s, according to Mühlenberg 1993) to ensure the same detection probability in all plots.

To test for the transferability of the resulting habitat suitability models in space (cf. Leftwich et al. 1997; Dennis and Eales 1999; Schröder and Richter 1999; Schröder 2000; Fleishman et al. 2003;

Binzenhöfer et al. 2005), we additionally sampled an area approximately 200 km away from our original study site in the Thuringian nature reserve ‘Leuttratal’ near Jena (latitude 50°52’, longitude 11°34’). This area is also characterised by a high fraction of dry grasslands with a wide variety of rare plant and animal species (Heinrich et al. 1998). Here we studied 28 experimental sites across five habitat types (Table 2) in the same manner as in our main study area. *M. bicolor* did not occur in that region, thus the spatial validation of the habitat suitability model for *M. bicolor* was not possible.

Statistical analyses

Development of habitat suitability models

The distribution, or response of an organism in regard to a given environmental variable is generally considered nonlinear (Gauch and Chase 1974; Austin 1976; Heglund et al. 1994). In recent years logistic regression analysis (GLM for a binomial response variable) has gained importance in the analyses of the relationship between independent variables (habitat parameters) and a dichotomous dependent variable (incidence of a specific species, Trexler & Travis 1993). This procedure is the only suitable one for an analysis of categorical variables (Capen et al. 1986). In addition, this method is favoured because of better results in the classification of results and the models are more robust compared to those from discriminant analyses. Additionally, coefficients are easy to interpret and a variety of measures for model calibration and discrimination have been developed (Nagelkerke 1991; Buckland et al. 1997; Fielding and Bell 1997; Hosmer and Lemeshow 2000; Manel et al. 2001; Austin 2002).

In our study, we used single and multiple parameter logistic regression models to predict occurrence probabilities depending on plot parameters (Manel et al. 1999a, b; Hosmer and Lemeshow 2000). For the selection of adequate models, we started with an univariate analysis to assess individual predictor variables independently from each other and to obtain information on each predictor’s performance (Hosmer and Lemeshow 2000). To choose uncorrelated parameters for the development of multiple parameter models we calculated all pairwise Spearman rank correlations. In cases of pairs showing a strong correlation ($\rho_s \geq 0.7$, cf. Fielding and Haworth 1995) only the variable delivering the best AUC-value (i.e. the area under a Receiver Operating Characteristic/ROC-curve as a measure of discriminative

Table 1 Description of the 10 main habitat types in the study area (according to the mapping system of the Bayerisches Landesamt für Umweltschutz; <http://www.bayern.de/ifu/natur/biotopkartierung/index.html>) and classification of these habitat types into EUNIS and CORINE Biotope classification (according to National Biodiversity Network Habitat Dictionary; www.nbn.org.uk/habitats)

Habitat type	EUNIS habitat description	EUNIS habitat type code	CORINE Biotopes Classification 1991	Characterisation of Bavarian habitat type mapping http://www.bayern.de/ifu/natur/biotopkartierung/index.htm
Crop land (CL)	Croplands planted for annually or regularly harvested crops other than those that carry trees or shrubs. They include fields of cereals, of sunflowers and other oil seed plants, of beets, legumes, fodder, potatoes and other forbs. Croplands comprise intensively cultivated fields as well as traditionally and extensively cultivated crops with little or no chemical fertilisation or pesticide application. Faunal and floral quality and diversity depend on the intensity of agricultural use and on the presence of borders of natural vegetation between fields.	I1 arable land and market gardens	82.11 field crops	On the numerous cultivated crop fields in the study region nutrient input is limited. Especially the soils on slopes and clayey Keuper grounds display bad qualities and need to be viewed as low crop areas. Thus, they are often temporally abandoned.
Fallow land (FL)	See crop field.	I1 arable land and market gardens	87 fallow land	Crop field lain fallow at most 3 years. Plant cover very scarce, consisting of annual and biennial pioneer species mixed with former cultivars and weeds.
Intensively managed meadows (IMM)	Lowland and montane mesotrophic and eutrophic pastures and hay meadows of the boreal, nemoral, warm-temperate humid and mediterranean zones. They are generally more fertile than dry grasslands (E1), and include sports fields and agriculturally improved and reseeded pastures.	E2 mesic grassland	38 mesophile grasslands	Species-poor tall stands characterised by a high proportion of grasses and a blue glaucous colour. These meadows are well fertilized and represent several times mown meadows or pastures.
Intensively managed poor meadows (IMMP)	Lowland and montane mesotrophic and eutrophic pastures and hay meadows of the boreal, nemoral, warm-temperate humid and mediterranean zones. They are generally more fertile than dry grasslands (E1), and include sports fields and agriculturally improved and reseeded pastures.	E2 mesic grasslands	38 mesophile grasslands	Fertilized variety of a <i>Arrhenatherum elatius</i> grassland with <i>Salvia pratensis</i> or wet grassland which is dominated by the species typically for intensively managed meadows (see Appendix). In contrast to intensively managed meadows these meadows show sporadic occurrence of pristine indicator species – frequent before fertilization – which give this meadows their characteristic appearance with single coloured spots

Table 1 continued

Habitat type	EUNIS habitat description	EUNIS habitat type code	CORINE Biotopes Classification 1991	Characterisation of Bavarian habitat type mapping http://www.bayern.de/ifu/natur/biotopkartierung/index.htm
Dry grassland (DG)	Well-drained or dry lands dominated by grass or herbs, mostly not fertilized and with low productivity. Included are [Artemisia] steppes. Excluded are dry mediterranean lands with shrubs of other genera where the shrub cover exceeds 10%; these are listed as garrigue (F6).	E1 dry grasslands	34.322D frankonian mesobromion	This habitat type comprises heat and drought tolerant basiphil dry grasslands. These are special habitats, which favour the survival of heliophytic plants by their situation, the climate, the landuse and the soil properties. The species composition depends largely on management. In dry grasslands managed by mowing preferably species like <i>Primula veris</i> and <i>Onobrychis viciifolia</i> will be found. Very often <i>Bromus erectus</i> is the dominating grass species. Sometimes indicators for pasture like <i>Cirsium acule</i> , <i>Carlina vulgaris</i> and <i>Pulsatilla vulgaris</i> can be found.
Extensively managed meadows (EMM)	Lowland and montane mesotrophic and eutrophic pastures and hay meadows of the boreal, nemoral, warm-temperate humid and mediterranean zones. They are generally more fertile than dry grasslands (E1), and include sports fields and agriculturally improved and reseeded pastures.	E2 mesic grasslands	38 mesophile grasslands	Extensively managed meadows are characterised by their regular but not too intensive management. These meadows are mown once- or twice a year but are not fertilized. Pasture occurs as rotational grazing. Often a large spectrum of grass and herb species is present, which gives this habitat type its characteristic colorful appearance in contrast to species-poor intensively managed and fertilised meadows. Extensively managed meadows show a loose stratification. Tall grasses do not cover the whole area. The stands are composed of species characteristic for habitats with mean to low nutrient supply. The indicator species for intensively managed grasslands <i>Alopecurus pratensis</i> , <i>Heracleum sphondylium</i> etc. decline clearly or are absent. Decisive for the characterisation as extensively managed meadow is the occurrence of a significant number of the indicator species (see Appendix).

Table 1 continued

Habitat type	EUNIS habitat description	EUNIS habitat type code	CORINE Biotopes Classification 1991	Characterisation of Bavarian habitat type mapping http://www.bayern.de/lfu/natur/biotopkartierung/index.htm
Fringe vegetation (FV)	Stands of tall herbs or ferns, occurring on disused urban or agricultural land, by water-courses, at the edge of woods, or invading pastures. Stands of shorter herbs forming a distinct zone (seam) at the edge of woods.	E5 woodland fringes and clearings and tall forb stands	34.4 thermophile forest fringe grassland	This habitat type comprises fallow land on nutrient- and nitrogen-poor dry sites, which are light exposed and warm. These sites are often situated between managed areas or close to adjacent forest edges. Sometimes hedge initials are already present. This habitat type occurs most often on areas where former dry grasslands and wine yards were present, in south and west exposition. This type is frequently encountered in the area of abandoned dry grassland; abandoned vineyards in dry and warm locations at south to west exposed forest edges as well as on other abandoned poor sites. Thermophile fringes are often in contact to dry grassland thermophile scrub or forest edges. Mostly fringes and stands rich in woody plants are merging. In hillside situation thermophile fringes may expand into the abandoned slopes. Similarly in extensively used meadows on slopes that are mown late in the year. Fringes where the shrubs were removed are also assigned to this type. Thermophile fringes are not managed and characterised by the occurrence of tall forbs belonging to the alliances <i>Geranium sanguinei</i> and <i>Trifolium medii</i> . In part initial shrubs are present.
Hedges (H)	Woody vegetation forming strips within a matrix of grassy or cultivated land or along roads, typically used for controlling livestock, marking boundaries or providing shelter. Hedgerows differ from lines of trees (G5.1) in being composed of shrub species, or if composed of tree species then being regularly cut to a height <5 m.	F.A.3 species-rich hedgerows of native species	84.2 hedgerows	Linear and plane shrub and tree-shrub occurrence. Species composition is highly dependend on site conditions mostly comprising mesophilic shrub species. Tre4e species with high budding potential and occasionally fruit trees are added. In ageing hedges hazel, rowan berry oak and sycamorepredominate. The understorey of hedges is uneven and reaches from indicators of poor habitats via mesphilic species to nitophytes.

Table 1 continued

Habitat type	EUNIS habitat description	EUNIS habitat type code	CORINE Biotopes Classification 1991	Characterisation of Bavarian habitat type mapping http://www.bayern.de/ifu/natur/biotopkartierung/index.htm
Forest (F)	Forest and woodland of mixed broad-leaved deciduous or evergreen and coniferous trees of the nemoral, boreal, warm-temperate humid and mediterranean zones. They are mostly characteristic of the boreonemoral transition zone between taiga and temperate lowland deciduous forests, and of the montane level of the major mountain ranges to the south. Neither coniferous, nor broadleaved species account for more than 75% of the crown cover. Deciduous forests with an understorey of conifers or with a small proportion of conifers in the dominant layer are included in unit G1. Conifer forests with an understorey of deciduous trees or with a small proportion of deciduous trees in the dominant layer are included in unit G3.	G1.6, G1.7, G3.42	43 mixed woodland	More or less dense woodland with native or introduced wood species, generally minimum width of 20 m and an area of 1 ha minimum.

power, cf. Hosmer and Lemeshow 2000) in univariate analysis was selected for further analysis. We did not use independent factors from principal component analysis (PCA, cf. Vaughan and Ormerod 2003) because these were found to create difficulties in their biological interpretation and were consequently difficult to use in conservation biology. As integrating measures for horizontal and vertical vegetation cover, we used the ‘total horizontal cover’, which describes the plots’ vertical structures (Sundermeier 1999), and the ‘percentage open ground’.

Initial single predictor models included all habitat types investigated. Due to total separation causing numerical instabilities in some habitat types we restricted our analyses to those habitat types with at least minimal variation in occupancy. This was done to achieve a more detailed explanation of the species’ habitat requirements. Eliminated biotopes were included in our models by formulating rules, like ‘*If forest then no suitable habitat*’. These can be easily implemented into the regression equations. Thus, the reduction of our data set increased our error due to the exclusion of observations that could be predicted without error, however, we receive more detailed information on the habitat selection of the species.

Model evaluation and test for spatial autocorrelation

For model calibration, which judges the concurrence between observed and predicted values (Schröder 2000) we used Nagelkerke’s pseudo- R^2 as a measure of goodness of fit (Nagelkerke 1991; Harrell 2001). Model discrimination, the power of the model to separate presence and absence of the species (Schröder 2000), was assessed with a threshold-independent measure, the AUC-value (Hanley and McNeil 1982; Fielding and Bell 1997). According to Hosmer and Lemeshow (2000) values above 0.7 describe an acceptable discrimination, values between 0.8 and 0.9 denote excellent discrimination. For a value above 0.9, discrimination is outstanding. For comparison of different models we used the Akaike Information Criterion (AIC, cf. Buckland et al. 1997; Augustin et al. 2001; Reineking and Schröder 2006).

Spatial autocorrelation has the effect of reducing the number of independent observations, which is not generally reflected by an equivalent decrease in the error degrees of freedom (Legendre 1993). Consequently, error terms are underestimated, leading to over-optimistic estimates of population parameters (Fielding and Haworth 1995) and abetting pseudoreplication (Guisan and Zimmermann 2000). To

Table 2 Overview of experimental sites, their distribution across habitat types and frequency of occupancy for the three species studied. Results are shown for two years at sample site ‘Hohe Wann’ and one year at the sample site ‘Leutratal’

Year	Location	Habitat type	No of plots	<i>P. albopunctata</i>		<i>M. bicolor</i>		<i>S. lineatus</i>	
				occup.	unocc.	occup.	unocc.	occup.	unocc.
2001	Hohe Wann	all	146	23	123	60	86	64	82
		Extensively managed meadow	45	6	39	24	21	25	20
		Intensively managed meadow	7	1	6	1	6	2	5
		Inten. managed poor meadow	24	0	24	5	19	7	17
		Dry grassland	26	9	17	18	8	21	5
		Fringe vegetation	10	7	3	9	1	9	1
		Crop land	8	0	8	1	7	0	8
		Fallow land	6	0	6	1	5	0	6
		Hedge	7	0	7	1	6	0	7
		Forest	7	0	7	0	7	0	7
2002	Hohe Wann	all	143	28	115	70	73	50	93
		Extensively managed meadow	45	13	32	25	20	16	29
		Intensively managed meadow	8	1	7	4	4	1	7
		Inten. managed poor meadow	22	0	22	7	15	3	19
		Dry grassland	26	7	19	22	4	20	6
		Fringe vegetation	10	7	3	8	2	9	1
		Crop land	8	0	8	1	7	0	8
		Fallow land	6	0	6	2	4	1	5
		Hedge	7	0	7	1	6	0	7
		Forest	6	0	6	0	6	0	6
2002	Leutratal	all	28	18	10	0	0	16	12
		Extensively managed meadow	5	2	3	0	0	4	1
		Intensively managed meadow	5	0	5	0	0	2	3
		Inten. managed meadow meagre	5	2	3	0	0	1	4
		Dry grassland	6	4	2	0	0	6	0
		Fringe vegetation	3	2	1	0	0	3	0
		Crop land	2	0	2	0	0	0	2
		Hedge	3	1	2	0	0	1	2

test whether our data show spatial autocorrelation, we calculated Moran’s *I* for standardised residuals as an index of covariance between different point locations (Lichstein et al. 2002; Karagatzides et al. 2003).

Model validation

One problem connected with models based on simple presence/absence data is that these data are only snap-shots from a certain time period and a certain region. Such models are static (Guisan and Zimmermann 2000) and need to be validated in space and time before they can be extrapolated to other areas (Morrison et al. 1998; Schröder and Richter 1999). We first applied a bootstrapping procedure (Verbyla and Litvaitis 1989; Efron and Tibshirani 1993; Reineking and Schröder 2006, Oppel et al. 2004) for internal validation. Additionally, we tested the transferability of the model in space (second study area; Freeman et al. 1997; Manel et al. 1999a, b; Schröder and Richter 1999) and time (second year; Dennis and

Eales 1999; Schröder 2000; Binzenhöfer et al. 2005) for external validation. To test the performance of these model transfers, we applied significance test of AUC-values (Beck and Shultz 1986; Schröder 2004; Binzenhöfer et al. 2005). A model transfer was regarded as successful if the AUC-value significantly exceeded a threshold of 0.7. All analyses were carried out with the statistical software R 1.7.1 (available at <http://cran.r-project.org> using the packages Hmsic and Design provided by F. Harrell).

Influence of spatial scale

To study the effects of the surrounding landscape composition on species occurrence on a larger spatial scale we used a method similar to Binzenhöfer et al. (2005). Therefore, we calculated the relative area of each habitat type in rings (radii *r* = 10 m, 25 m as well as *r* = 50 m) around the plot and weighted them using the predicted occurrence probability determined in the univariate logistic regression analyses with the habitat type as plot parameter. In each case the inner rings

(either the plot itself ($r = 10$ m), or the ring with $r = 25$ m) were subtracted from the outer rings ($r = 25$ m and $r = 50$ m). These calculations were carried out with a GIS (ESRITM ArcView 3.2). If we found overlapping rings we excluded one by random selection to avoid pseudoreplication. This resulted in a reduction of our data sets from $n = 146$ to $n = 118$. The method produces one single metric regression parameter for each radius (instead of categorical variables or percentages) and thus avoids the use of too many degrees of freedom in the analyses. To see whether the immediate surrounding landscape has a significant influence on species occurrence, the values for $r = 25$ m and $r = 50$ m are added to the values for $r = 10$ m (which corresponds to our experimental plot of 15×15 m) in a multiple regression analysis. Expansion of this analysis to scales probably more relevant for dispersal and metapopulation aspects (e.g. $r = 100$ m or 200 m) was not possible in this study. Our experimental sites were restricted to the nature reserve and thus, too many overlapping rings would have resulted in a severe reduction of our data sets.

To check for habitat preferences of our species within one habitat type (smaller spatial scale) we compared the characteristics of occupied plots with those from unoccupied plots.

Results

The prevalence varied between species, but was almost constant over the years (Table 2). In contrast to the main study area, *S. lineatus* as well as *P. albopunctata* exhibited a very high prevalence in the second study area (*S. lineatus*: 57.1%, *P. albopunctata*: 34.3%).

Based on the occupancy pattern across some of habitat types and complete absences in certain habitats (Table 2) we deduced the following rules:

- *Platycleis albopunctata* does not occur in: rich meadows, extensively managed meadows, crop land, fallow land, hedges, and forests.
- *Metrioptera bicolor* does not occur in: rich meadows, crop land, fallow land, hedges, and forests.
- *Stenobothrus lineatus* does not occur in: rich meadows, crop land, fallow land, hedges, and forests.

Analysing the predictors ‘habitat type’ and ‘type of management’

After internal validation, the predictor variable ‘habitat type’ showed a high explanatory power for all three

species in models when it was used as single predictor variable (Table 3). These models were transferable in time for all species, for *S. lineatus* the transfer was also possible in space (Table 3).

For all three species, the ‘fringe vegetation’ has the highest probability of occurrence followed by dry grassland (Fig. 1). The ‘management type’ alone did not yield high explanatory power for the spatial distribution of the species, but was included in some of the multiple models and may be important in terms of conservational aspects. For *P. albopunctata* mowing always resulted in a high incidence. The other two species (*M. bicolor* and *S. lineatus*) are most often found on plots under extensive sheep-grazing management (Fig. 2). Intensively managed areas as well as areas with no management at all are avoided by all three species. Generally, the ‘management type’ cannot explain as much variation in incidence as the predictor ‘habitat type’.

Influence of plot characteristics on the species’ occurrence probabilities

Because habitat suitability may not be determined by a single factor alone, but probably by a combination of different factors we conducted multiple logistic regression analyses for each species with different combinations of independent variables from the reduced data sets.

Stenobothrus lineatus

Multiple logistic regression analyses for the data from 2001 resulted in six significant models with an AUC-value exceeding 0.7. All models were free of residual spatial autocorrelation. Table 2 shows those four models with low AIC-values. The model with the lowest AIC predicts a high occurrence of *S. lineatus* in fringes, dry grassland, extensively managed meadows and grazed areas with low vegetation height. Only two of the multiple models were transferable in time, but none in space (Table 2). Internal validation of the models showed that only those with the variables ‘habitat type’, ‘low vegetation height’ and ‘low total horizontal cover’ were robust, but they were neither transferable in time nor space.

Metrioptera bicolor

Of the six significant multiple parameter models, which all showed no residual spatial autocorrelation, the highest occurrence of *M. bicolor* was predicted

Table 3 Model characteristics for significant models ($p < 0.05$) with an AUC-value ≥ 0.7 for the three investigated species: AUC with SE, $R^2_{\text{Nagelkerke}}$, AIC, $AUC_{\text{bootstrapped}}$ after internal validation of the whole model, predictors included in more than 50% of the 300 bootstrap models (stable predictors), $AUC_{\text{bootstrapped}}$ for the model only including the stable predictors. Transferability in space (s) or time (t) is indicated by indices

Species	Model	Predictor variables	AUC \pm SE	$R^2_{\text{Nagelkerke}}$	AIC	full model	bootstrapping	
							$AUC_{\text{bootstrapped}}$	stable predictors
<i>S. lineatus</i>	S1 ^t	Habitat type; Management; Vegetation height (quadratic term)	0.848 \pm 0.036	0.508	102.35	0.807	habitat type	0.725
	S2 ^t	Habitat type; Vegetation height (quadratic term); Vegetation height	0.809 \pm 0.042	0.366	114.85	0.779	Habitat type Vegetation height (quadratic term)	0.728
	S3	Habitat type; Vegetation height (quadratic term)	0.762 \pm 0.046	0.367	115.92	0.735	Whole model	
	S4	Management; Total horizontal cover	0.786 \pm 0.045	0.335	117.73	0.766	Total horizontal cover	0.758
	S5 st	Habitat type (all data)	0.846 \pm 0.03	0.505	133.14	0.834	Whole model	
	M1	Habitat type; Cosine exposition; Vegetation height; Sheep lifestock	0.799 \pm 0.044	0.347	120.30	0.757	Habitat type	0.709
<i>M. bicolor</i>	M2	Habitat type; Cosine exposition; Vegetation height	0.780 \pm 0.045	0.301	122.99	0.750	Habitat type	0.709
	M3	Habitat type; Sheep lifestock; Vegetation height	0.782 \pm 0.044	0.310	123.39	0.751	Habitat type	0.709
	M4	Habitat type; Sheep lifestock	0.747 \pm 0.047	0.277	124.63	0.724	Habitat type	0.709
	M5	Habitat type; Vegetation height	0.772 \pm 0.046	0.267	125.65	0.749	Habitat type	0.709
	M6	Management; Cosine exposition; Vegetation height; Total horizontal cover	0.741 \pm 0.048	0.268	128.32	0.675	No stable predictor found	
	M7 ^t	Habitat type (all data)	0.806 \pm 0.04	0.387	150.3	0.787	Whole model	
<i>P. albopunctata</i>	P1	Habitat type; Management; Vegetation height	0.864 \pm 0.047	0.475	68.58	0.807	No stable predictor found	
	P2	Habitat type; Sine exposition; Vegetation height	0.855 \pm 0.047	0.434	71.98	0.806	Whole model	
	P3	Habitat type; Vegetation height	0.812 \pm 0.054	0.354	76.09	0.787	Whole model	
	P4 ^t	Habitat type (all data)	0.855 \pm 0.04	0.415	88.9	0.838	No stable predictor found	

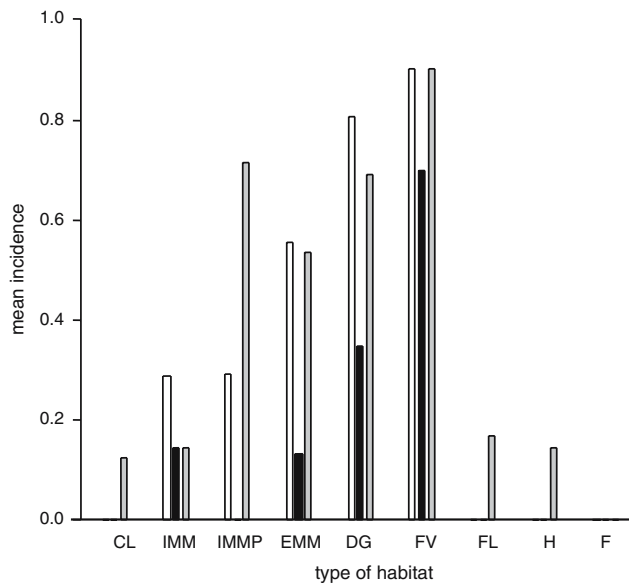


Fig. 1 Mean incidence in the habitat types: crop land (CL), intensively managed meadows (IMM), intensively managed poor meadows (IMMMP), extensively managed meadows (EMM), dry grasslands (DG), fringe vegetation (FV), fallow land (FL), hedges (H), forest (F) for *S. lineatus* (white bars), *M. bicolor* (grey bars) and *P. albopunctata* (black bars)

for sheep-grazed fringe vegetation with tall vegetation and south-facing exposition. However, after internal validation with backwards variable selection, all multiple parameter models were identified as unstable (i.e. specific predictors were considered in

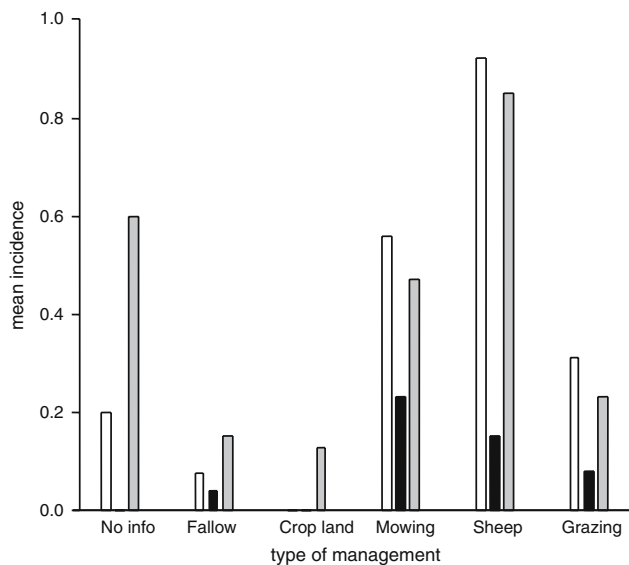


Fig. 2 Mean incidence for different types of management for *S. lineatus* (white bars), *M. bicolor* (grey bars) and *P. albopunctata* (black bars)

less than 50% of 300 bootstraps and thus excluded) and had to be reduced to single predictor models with ‘habitat type’ as the single explanatory variable (Table 2). None of the more complex models was transferable in time. Spatial validation was not possible because *M. bicolor* could not be found in the second area.

Platycleis albopunctata

The model considering ‘habitat type’, ‘management type’ and ‘vegetation height’ yielded the smallest AIC-value (Table 3). But after internal validation with backwards variable selection none of these variables was included in more than 50% of the 300 bootstraps. Out of the three significant multiple models with the low AIC-values (all without residual spatial autocorrelation) the model considering ‘habitat type’, ‘sine exposition’ and ‘vegetation height’ was considered to be the best one. *P. albopunctata* prefers sites of mown fringe vegetation, south-west exposition and general low vegetation height. None of the multiple parameter models was transferable in time or space.

Influence of spatial scale

Finally, we carried out a comparison of occupied and unoccupied plots separately for each ‘habitat type’. This allowed a closer look at habitat selection and yields habitat-specific models. Low ‘vegetation height’, low ‘total horizontal cover’ as well as low ‘cover at the heights of 20/30/40 cm’ are attributes preferred by *S. lineatus* on dry grasslands (Mann–Whitney *U*-test, $p < 0.05$ for all cases). By conducting the same analyses for *M. bicolor* we could not detect any significant differences between occupied and unoccupied plots (Mann–Whitney *U*-test, $p > 0.05$ for all cases). *P. albopunctata* prefers extensively managed west-facing meadows (Mann–Whitney *U*-test, $p < 0.05$ for all cases).

For *P. albopunctata* and *M. bicolor* no additional influence of the surroundings on habitat occupancy could be detected for the radii analysed in this study. Instead, for *S. lineatus* the surrounding area between 25 and 50 m was a significant variable in the model together with the plots’ own habitat type ($AUC_{bootstrapped} = 0.823$ compared to $AUC_{bootstrapped} = 0.725$ for the habitat type as single predictor variable).

Discussion

Influence of plot characteristics on the occurrence probability of species

In our study, the habitat-specialist exhibiting the most restricted habitat requirements was *P. albopunctata*. It only occurred on fringe vegetation, dry grasslands and ‘intensively managed poor meadows’. The two other species, *M. bicolor* and *S. lineatus*, were also found on extensively managed meadows. This result corresponds well with the literature on habitat requirements of these species (Detzel 1998).

After the reduction of the data set to the habitat types that yielded some incidence, the variable ‘habitat type’, which comprises several aspects of the realised niches, significantly contributes to many models. ‘Habitat type’ obviously has a great influence on the occurrence probability of the species. Additionally for *S. lineatus*, low vegetation height as well as low total horizontal cover, strongly influence occurrence positively. This may be explained by the fact that egg development depends on temperature (van Wingerden et al. 1991) and females lay their eggs in the upper ground layer or at the bottom of grasses (Oschmann 1993).

For *M. bicolor* a south-facing position, tall vegetation and sheep grazing are the factors, which in combination with the habitat type, explained most of the variance in the data. For this species, fringes, dry grasslands, and extensively managed meadows offer good food resources as the larvae feed on grasses as well as on flowers of grasses and herbs (Ingrisch 1976). This holds especially when the management is sheep grazing. As males of this species call in tall vegetation (Detzel 1998) and females lay their eggs in grass stems (Hartley and Warne 1972), a preference for high vegetation may correspond to the vertical orientation of the species.

The occurrence of *P. albopunctata* is best described by the combination of the variables ‘habitat type’, ‘sine exposition’ and ‘vegetation height’ in the plots. This corresponds to the finding of Detzel (1998) who describes that low vegetation height as well as southwest exposition result in an increased temperature that promotes the larval development of *P. albopunctata*. Apart from the influence of the habitat type, management regime contributes to habitat suitability for all three species. *P. albopunctata* was mainly found on mown dry grasslands and fringes, which provide an elevated ground temperature, and low vegetation due to their position on the

steeper, upper south-facing hillsides. The other two species preferred areas, which were extensively grazed by sheep. Detzel (1998) also describes a preference by *P. albopunctata* for areas managed by sheep grazing, but our results seem to be in contrast with this. Our result may, however, be biased by the fact that the dry grasslands as well as the fringes investigated are all managed by mowing, therefore mowing and fringes cannot be separated. Although we cannot precisely predict the best management practice for the conservation of *P. albopunctata*, it appears that mown habitats may be suitable for this species.

In general, grazing by sheep mainly occurs in extensively managed meadows, where it produces a heterogeneous mosaic of tall and short vegetation patches within a site (Adler et al. 2001). Such complex habitats offer a variety of different local conditions with varying microclimates for different activities like feeding, mating or reproduction. Intensively managed, agricultural or silvicultural areas, as well as abandoned areas with no management at all are avoided by all three species. For intensively managed areas and agricultural land, the regular disturbance intervals at times when adults are reproducing or eggs are developing, rather than negative microclimatic conditions, appear to be the reason for an absence of the species. This is supported by the fact that adults of *P. albopunctata* did not leave crop land, but females were observed ovipositing after being released there artificially (Hein et al. 2003). Woodland as well as abandoned areas are most likely too cold and wet for the development of offspring.

Both, the ‘habitat type’ as well as the ‘management type’ are proxies for the real resources such as microclimatic differences based on factors like plant species composition, vegetation structure and density and are driving factors for the incidence of grasshoppers and bush crickets. In particular the categories of the habitat type display various aspects of the realised niche for the studied species (see also Samways and Moore (1991) who found no significant relationship between grasshopper assemblage and microclimatic temperature but a strong positive correlation with grass species richness). Thus, the influence of the parameter habitat type on occurrence probability is determined by a variety of other factors such as predation risk, temperature regime, oviposition and microclimatic conditions that are relevant for the species’ habitat selection and survival. This may

explain the contrasting effects of management regime for *P. albopunctata*. It is neither the ‘habitat type’ nor the exact management regime that determines species occurrence, but the resulting level of the relevant factors, such as increased temperature for egg development, which determines habitat quality for the species.

Model validation

Validation either internally or externally is the best means of determining the robustness and generality of a model (cf. Freeman et al. 1997; Glozier et al. 1997; Leftwich et al. 1997; Dennis and Eales 1999; Roloff and Kernoban 1999; Schröder and Richter 1999; Zimmermann and Kienast 1999; Bio et al. 2002; Lehmann et al. 2002a; Fleishman et al. 2003; Opper et al. 2004; Pepler-Lisbach and Schröder 2004; Binzenhöfer et al. 2005). For all three species the simple model including ‘habitat type’ as the explanatory variable could be validated internally and showed a good transferability in time. For *S. lineatus* it could also be transferred in space. Spatial validation was not possible in any other case due to the low sample size in the second study area (28 plots) and the fact that the ranges of tested categories for ‘habitat type’ and ‘management’ in the second study area were too small (see Table 2). In such a case, it is more likely that we compared two different sampling designs rather than performing an external validation (Lehmann et al. 2002b). For future studies we would recommend the use of the same sampling design with similar sampling effort to perform a spatial validation of habitat suitability models. The failure of transferability in time for the multiple parameter models of *M. bicolor* might be caused by their instability. None of these models were stable with respect to the selected explanatory variables and only the variable ‘habitat type’ resulted in stable models after bootstrapping with stepwise backward variable selection.

Methodological aspects of the study

Experimental design

Although the type of stratified random sampling used in our study is an adequate and commonly used method (see also Wessels et al. 1998; Hirzel and Guisan 2002), we would recommend the use of a two-step approach whenever possible for further studies. This should start with a preliminary study (which was

not possible in our case) with which clear non-habitat structures can be determined to exclude them from the analyses (Dufrene and Legendre 1991; Aspinall and Lees 1994). The intensive work on the detailed habitat requirements of a species can then be conducted in a main study with a high sample size in habitats with intermediate occupancy (see also Hirzel and Guisan 2002). Such an approach with higher sample sizes always improves the quality of the results (Hirzel and Guisan 2002). In the case of the bush cricket *P. albopunctata* with its low prevalence of around 20% in the nature reserve ‘Hohe Wann’, a higher sample size would almost certainly have improved the precision of our model. In the case of the other two investigated species, prevalence was around 40–50%. This is regarded as optimal for the development of logistic regression models (Hosmer and Lemeshow 2000).

The influence of spatial scale

For the determination of species specific habitat preferences, one should always take into account that habitat selection behaviour is a scale-dependent process (Johnson 1980; Orians and Wittenberger 1991; Mackey and Lindenmayer 2001; Opper et al. 2004). Thus an analysis of occurrence probability should be carried out on different spatial scales (Orians and Wittenberger 1991). This fact has been studied separately in a number of analyses (Poff 1997; Lindenmayer 2000; Cushman and McGarigal 2002; Luck 2002; Thompson and McGarigal 2002; Sergio et al. 2003; Store and Jokimaki 2003; Grand and Mello 2004; Opper et al. 2004; Parody and Milne 2004; Poirazidis et al. 2004; Aubry et al. 2005; Graf et al. 2005; Legalle et al. 2005; Mörtberg and Karlström 2005). To account for this, we further expanded our analyses and looked for microhabitat preferences within one experimental plot. Therefore, we used only data from occupied plots and compared the parameters from the random point with those of the ‘cricket (detection) point’ in the same experimental plot. In our study, none of the species showed any significant preference for distinct microhabitat parameters (Wilcoxon match paired test after Bonferroni correction according to Rice (1989) for multiple comparisons, all comparisons $p > 0.05$). Regarding *M. bicolor*, this contrasts with the findings of Krätzel (1999) and Krätzel et al. (2002) who found evidence for the preference for distinct microstructures. This may be due to the fine-grained scale of our study. Our plots

were rather small (just 225 m²) compared to the 800–1320 m² in the study of Krätzel (1999) and Krätzel et al. (2002). Thus, our plots may have been rather homogeneous. We would nevertheless expect that our species do actively select distinct microhabitats in a larger experimental site. This argument is supported by our separate analyses of each habitat type which showed a clear preference of *P. albopunctata* and *S. lineatus* for some structural aspects of occupied compared to unoccupied plots.

In many other studies, scale dependency of predictors is discussed and the influence of the surrounding of the tested areas has been shown (e.g. Binzenhöfer et al. 2005). In our case no additional influence of the plot surrounding on habitat occupancy could be detected for *P. albopunctata* and *M. bicolor*. This is probably because the information given by the plots' habitat type is the same as that of a radius of 25 m. Thus, our radii do not represent the true surroundings in the sense of dispersal distance of the insects. Whether the inclusion of the surrounding between 25 and 50 m in the model of *S. lineatus* is based on metapopulation effects or the spatial heterogeneity of *S. lineatus* habitats cannot be deduced using the data from this study.

Implications for conservation

From our analyses we would conclude that for the conservation of these grasshopper and bush cricket species in the nature reserve 'Hohe Wann' management should aim to maintain the extensively managed meadows and especially dry grasslands and fringe vegetation. Whether this is achieved by mowing, sheep grazing or another studied management regime does not seem to be of great importance for the studied species as long as management does not occur too often (more than twice a year) and at times when reproduction or development of eggs is not disturbed (early or late in the season; for comparison see Chambers and Samways 1998). Regarding our results on parameters determining species occurrence, we know that we found no new relevant parameters for species occurrence compared with literature data for the temperate zones, but we gathered quantitative information on the parameters' influences. This information can be used in modelling approaches to test different management

scenarios in their outcome for species survival (see Rudner et al. 2005; Schröder et al. submitted).

We are able to show that the habitat type is always a powerful predictor of species occurrence when we use it as a single predictor variable or even in the reduced data sets and multiple regression analyses. Thus, we can, on the one hand, conclude that the habitat type as a highly integrating variable, accounts for some parameters we did not survey (e.g. microclimate). On the other hand simple vegetation type maps (depicting habitat types) are sufficient to predict species occurrence with satisfactory precision, which is especially useful for nature conservation. Because the habitat type also represents a specific successional stage respectively a certain kind of management our models are able to predict in an indirect way the quality of habitats under different management. The use of these models would provide a quick and efficient possibility to determine the future/unknown distribution of species under different management regimes (see application of our data by Schröder et al. submitted) as well as in other regions where species distribution data are not available (Wilson et al. 2005, see also Binzenhöfer et al. 2005; Strauss and Biedermann 2005). Additionally, habitat models can be used to extrapolate ecological knowledge from point data to the landscape scale by creating habitat suitability maps (see Schröder 2000; Strauss and Biedermann 2005). These could be used as a basis for planning nature reserves and for the identification of core habitats (Cabeza et al. 2004). In such cases, the approach of statistical habitat suitability models is especially appealing as the data from our species could be combined with those of other animals (Hein et al. in press) and/or plant species and thus a multi-species approach could be used to identify the most "valuable" areas in terms of species diversity.

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Appendix

Table 4 Plant indicator species of the 10 main habitat types in the study area

Habitat type	Plant indicator species
Crop field (CF)	Crops, corn, root crop, legumes, oil- and fibre plants. Herbs like: <i>Adonis aestivalis</i> , <i>Apera spica-venti</i> , <i>Campanula arvensis</i> , <i>Conringia orientalis</i> , <i>Consolida regalis</i> , <i>Galinoga parviflora</i> , <i>Lolium multiflorum</i> , <i>Lolium perenne</i> , <i>Matricaria maritima</i> , <i>Melampyrum arvense</i> , <i>Papaver dubium</i> , <i>Papaver rhoeas</i> , <i>Polygonum laplatifolia</i> , <i>Polygonum persicaria</i> , <i>Senecio vernalis</i> , <i>Sinapis alba</i> , <i>Sinapis arvensis</i> , <i>Sonchus asper</i> , <i>Veronica arvensis</i> , <i>Viola arvensis</i> , <i>Viola arvensis</i> , <i>Viola tricolor</i>
Fallow land (FL)	<i>Anagallis arvensis</i> , <i>Apera spica-venti</i> , <i>Bromus arvensis</i> , <i>Bromus sterilis</i> , <i>Capsella bursa-pastoris</i> , <i>Chrysanthemum leucanthemum</i> , <i>Cirsium arvense</i> , <i>Erigeron annuus</i> , <i>Euphorbia div. spec.</i> , <i>Geranium molle</i> , <i>Lepidium campestre</i> , <i>Lepidium ruderales</i> , <i>Matricaria discoidea</i> , <i>Matricaria maritima</i> , <i>Matricaria recutita</i> , <i>Papaver dubium</i> , <i>Papaver rhoeas</i> , <i>Poa annua</i> , <i>Potentilla anserina</i> , <i>Potentilla reptans</i> , <i>Rapistrum rugosum</i> , <i>Scleranthus annuus</i> , <i>Senecio vernalis</i> , <i>Sinapis alba</i> , <i>Sinapis arvensis</i> , <i>Spergularia rubra</i> , <i>Stellaria media</i> , <i>Veronica persica</i> , <i>Vicia hirsuta</i> , <i>Vicia sativa</i> , <i>Vicia tetrasperma</i> , <i>Viola arvensis</i> , <i>Viola tricolor</i>
Intensively managed meadows (IMM)	<i>Achillea millefolium</i> , <i>Alopecurus pratensis</i> , <i>Arrhenatherum elatius</i> , <i>Dactylis glomerata</i> , <i>Festuca pratensis</i> , <i>Heracleum sphondylium</i> , <i>Lolium multiflorum</i> , <i>Lolium perenne</i> , <i>Phleum pratense</i> , <i>Pheum pratense</i> , <i>Ranunculus repens</i> , <i>Ranunculus acris</i> , <i>Taraxacum officinale</i> agg., <i>Trifolium dubium</i> , <i>Trifolium repens</i> , <i>Trifolium pratense</i>
Intensively managed poor meadows (IMMP)	Species listed above under intensively managed meadows. In addition: <i>Anthoxanthum odoratum</i> , <i>Anthriscus sylvestris</i> , <i>Avenula pubescens</i> , <i>Bromus hordeaceus</i> , <i>Bromus sterilis</i> , <i>Centaurea jacea</i> , <i>Daucus carota</i> , <i>Festuca pratensis</i> , <i>Festuca rubra</i> , <i>Holcus lanatus</i> , <i>Knautia arvensis</i> , <i>Lathyrus pratensis</i> , <i>Leucanthemum vulgare</i> , <i>Poa pratensis</i> , <i>Poa trivialis</i> , <i>Potentilla reptans</i> , <i>Rumex acetosa</i> , <i>Salvia pratensis</i> , <i>Centaurea jacea</i> , <i>Plantago div. spec.</i> , <i>Silvaum silaus</i>
Dry grassland (DG)	<i>Anthericum ramosum</i> , <i>Anthyllis vulneraria</i> agg., <i>Asperula cynanchica</i> , <i>A. tinctoria</i> , <i>Aster amellus</i> , <i>Aster linosyris</i> , <i>Bromus erectus</i> , <i>Carlina acaulis</i> , <i>Carlina vulgaris</i> , <i>Carex ericetorum</i> , <i>C. humilis</i> , <i>C. montana</i> , <i>C. caryophylla</i> , <i>Cirsium acule</i> , <i>Dianthus carthusianorum</i> , <i>Euphorbia verrucosa</i> , <i>Festuca ovina</i> , <i>Galium boreale</i> , <i>Geniella ciliata</i> , <i>G. germanica</i> , <i>Helianthemum nummularium</i> agg., <i>Hippocrepis comosa</i> , <i>Hypochoeris maculata</i> , <i>Linum austriacum</i> , <i>Orchis militaris</i> , <i>O. morio</i> , <i>O. purpurea</i> , <i>O. ustulata</i> , <i>Ophrys div. spec.</i> , <i>Pimpinella saxifraga</i> , <i>Polygala comosa</i> , <i>Prunella grandiflora</i> , <i>Pulsatilla vulgaris</i> agg., <i>Ranunculus bulbosus</i> , <i>Teucrium chamaedrys</i> , <i>T. montanum</i> , <i>Trifolium montanum</i> , <i>Veronica teucrium</i>
Extensively managed meadows (EMM)	Besides characteristic species of intensively managed meadows like <i>Arrhenatherum elatius</i> , <i>Achillea millefolium</i> and <i>Trifolium pratense</i> the following species are characteristic for this habitat type: <i>Agrostis capillaris</i> , <i>Anthoxanthum odoratum</i> , <i>Arrhenatherum elatius</i> , <i>Avena pratensis</i> , <i>Avena pubescens</i> , <i>Avenella flexuosa</i> , <i>Brachypodium pinnatum</i> , <i>Briza media</i> , <i>Bromus erectus</i> , <i>Campanula glomerata</i> , <i>Campanula rotundifolia</i> , <i>Centaurea jacea</i> , <i>Centaurea scabiosa</i> , <i>Chrysanthemum leucanthemum</i> , <i>Cynosurus cristatus</i> , <i>Dactylis glomerata</i> , <i>Festuca ovina</i> agg., <i>Festuca pratensis</i> , <i>Festuca rubra</i> , <i>Holcus lanatus</i> , <i>Hypochoeris maculata</i> , <i>Knautia arvensis</i> , <i>Leucanthemum ircutianum</i> , <i>Linum catharticum</i> , <i>Lotus corniculatus</i> , <i>Luzula campestris</i> , <i>Meum athamanticum</i> , <i>Onobrychis viciaefolia</i> , <i>Ononis spinosa</i> , <i>Pimpinella saxifraga</i> , <i>Plantago div. spec.</i> , <i>Poa angustifolia</i> , <i>Poa pratensis</i> , <i>Potentilla tabernaemontani</i> , <i>Primula veris</i> , <i>Prunella vulgaris</i> , <i>Ranunculus auricomus</i> , <i>Ranunculus bulbosus</i> , <i>Rhinanthus alectorolophus</i> , <i>Salvia pratensis</i> , <i>Sanguisorba minor</i> , <i>Saxifraga granulata</i> , <i>Thymus pulegioides</i> , <i>Trisetum flavescens</i>
Fringe vegetation (FV)	<i>Brachypodium pinnatum</i> , <i>Briza media</i> , <i>Bromus erectus</i> , <i>Campanula glomerata</i> , <i>Campanula rotundifolia</i> , <i>Carex brizoides</i> , <i>Carex leporina</i> , <i>Centaurea jacea</i> , <i>Centaurea scabiosa</i> , <i>Chrysanthemum leucanthemum</i> , <i>Cynosurus cristatus</i> , <i>Dactylis glomerata</i> , <i>Deschampsia cespitosa</i> , <i>Festuca ovina</i> , <i>Festuca pratensis</i> , <i>Festuca rubra</i> , <i>Galium verum</i> , <i>Holcus lanatus</i> , <i>Knautia arvensis</i> , <i>Lotus corniculatus</i> , <i>Luzula campestris</i> , <i>Molinia caerulea</i> , <i>Agrimonia eupatoria</i> , <i>Anemone sylvestris</i> , <i>Anthericum liliago</i> , <i>Anthericum ramosum</i> , <i>Aster amellus</i> , <i>A. linosyris</i> , <i>Astragalus cicer</i> , <i>Astragalus glycyphyllos</i> , <i>Bupleurum falcatum</i> , <i>Calamintha clinopodium</i> , <i>Campanula persicifolia</i> , <i>Campanula rapunculoides</i> , <i>Chrysanthemum corymbosum</i> , <i>Coronilla varia</i> , <i>Dianthus armeria</i> , <i>Dicamnus albus</i> , <i>Fragaria viridis</i> , <i>Geranium sanguineum</i> , <i>Inula salicina</i> , <i>Laserpitium latifolium</i> , <i>Laserpitium siler</i> , <i>Lychnis viscaria</i> , <i>Malva alcea</i> , <i>Melampyrum arvense</i> , <i>Melampyrum cristatum</i> , <i>Origanum vulgare</i> , <i>Peucedanum cervaria</i> , <i>Polygonatum odoratum</i> , <i>Bupleurum falcatum</i> , <i>Seseli libanotis</i> , <i>Silene nutans</i> , <i>Teucrium scorodonia</i> , <i>Thalictrum minus</i> , <i>Thesium bavarum</i> , <i>Trifolium alpestre</i> , <i>Trifolium medium</i> (high cover >20%), <i>Veronica teucrium</i> , <i>Vincetoxicum hirsutum</i> , <i>Viola hirta</i>

Table 4 continued

Habitat type	Plant indicator species
Hedges (H)	<i>Acer campestre</i> , <i>Acer pseudoplatanus</i> , <i>Alnus glutinosa</i> , <i>Berberis vulgaris</i> , <i>Betula pendula</i> , <i>Carpinus betulus</i> , <i>Clematis vitalba</i> , <i>Cornus sanguinea</i> , <i>Corylus avellana</i> , <i>Crataegus div. spec.</i> , <i>Euonymus europaeus</i> , <i>Frangula alnus</i> , <i>Fraxinus excelsior</i> , <i>Ligustrum vulgare</i> , <i>Lonicera xylosteum</i> , <i>Malus sylvestris</i> , <i>Populus tremula</i> , <i>Prunus avium</i> , <i>Prunus domestica</i> , <i>Prunus spinosa</i> , <i>Pyrus communis</i> , <i>Quercus petraea</i> , <i>Quercus robur</i> , <i>Rhamnus catharticus</i> , <i>Ribes uva-crispa</i> , <i>Rosa div. spec.</i> , <i>Rubus div. spec.</i> , <i>Salix div. spec.</i> , <i>Sambucus nigra</i> , <i>Sambucus racemosa</i> , <i>Sorbus aucuparia</i> , <i>Viburnum opulus</i> , <i>Cotoneaster integerrimus</i> , <i>Cotoneaster tomentosus</i> , <i>Hippophae rhamnoides</i> , <i>Viburnum lantana</i> , <i>Pyrus pyrastrer</i> , <i>Sorbus aria</i> , <i>Sorbus torminalis</i> , <i>Sorbus domestica</i> .
Forest (F)	Herb and grass species: <i>Agrimonia eupatoria</i> , <i>Brachypodium pinnatum</i> , <i>Coronilla varia</i> , <i>Medicago falcata</i> , <i>Origanum vulgare</i> , <i>Trifolium mediuminter alia</i> , <i>Agropyron repens</i> , <i>Aegopodium podagraria</i> , <i>Alliaria petiolata</i> , <i>Anthriscus sylvestris</i> , <i>Chaerophyllum div. spec.</i> , <i>Galium aparine</i> , <i>Geum urbanum</i> , <i>Glechoma hederacea</i> , <i>Lamium maculatum</i> , <i>Torilis japonica</i> , <i>Urtica dioica</i> ; <i>inter alia</i> >5% Tree- and shrub layer: <i>Acer campestre</i> , <i>Acer platanoides</i> , <i>Acer pseudoplatanus</i> , <i>Betula pendula</i> , <i>Carpinus betulus</i> , <i>Fagus sylvatica</i> , <i>Hedera helix</i> , <i>Picea abies</i> , <i>Pinus sylvestris</i> , <i>Prunus padus</i> , <i>Prunus serotina</i> , <i>Quercus petraea</i> , <i>Quercus robur</i> , <i>Quercus rubra</i> , <i>Robinia pseudoacacia</i> , <i>Sorbus aucuparia</i> , <i>Tilia cordata</i> , <i>Tilia platyphyllos</i> Herb- and grass layer: <i>Allium ursinum</i> , <i>Arum maculatum</i> , <i>Avenella flexuosa</i> , <i>Convallaria majalis</i> , <i>Dryopteris carthusiana</i> , <i>Galium odoratum</i> , <i>Galium sylvaticum</i> , <i>Holcus mollis</i> , <i>Hordelymus europaeus</i> , <i>Lathyrus vernus</i> , <i>Leucobryum glaucum</i> , <i>Lonicera periclymenum</i> , <i>Luzula luzuloidea</i> , <i>Melica uniflora</i> , <i>Oxalis acetosella</i> , <i>Polygonatum multiflorum</i> , <i>Polytrichum formosum</i> , <i>Pteridium aquilinum</i> , <i>Stellaria holostea</i> , <i>Teucrium scorodonia</i> , <i>Milium effusum</i> , <i>Trientalis europaea</i> , <i>Viola hirta</i> , <i>Viola reichenbachiana</i> , <i>inter alia</i>

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