

Jaw musculature during the dawn of turtle evolution

Ingmar Werneburg

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Abstract Using a new approach to study muscle anatomy in vertebrates, the fully differentiated jaw musculature of 42 turtle species was studied and character mappings were performed. Soft tissue arrangements were correlated to the temporal openings (emarginations) of the skull and the trochlearis system of the jaw apparatus among turtle taxa. When compared to the cranial anatomy of stem Testudines, most characters detected as diagnostic of particular extant groups have to be considered as being evolved first within Testudines. Hence, jaw muscle anatomy of extant turtles is difficult to compare to that of other reptilian taxa. Moreover, the high number of apomorphic character changes speaks for a divergating turtle and saurian morphotype of jaw musculature, which could indicate either a position of turtles outside of Sauria or a highly derived, undetectable origin within that group. In general, a low direct correlation of most soft and hard tissue characters was detected. This finding could imply that both character complexes are more integrated to each other driven by functional morphology; i.e., the composition of muscle fibre types. That condition highlights the difficulty in using gross anatomy of jaw muscle characters to interpret temporal bone arrangements among amniotes in general.

Keywords Cryptodira · Pleurodira · Character mapping · Musculus adductor mandibulae · Temporal skull region · Feeding · Testudinata · Trochlearis system · Fossil turtles

I. Werneburg (✉)
Fachbereich Geowissenschaften der Eberhard Karls Universität
Tübingen,
Hölderlinstraße 12,
72074 Tübingen, Germany
e-mail: i.werneburg@gmail.com

I. Werneburg
Paläontologisches Institut und Museum der Universität Zürich,
Karl Schmid-Strasse 4,
8006 Zürich, Switzerland

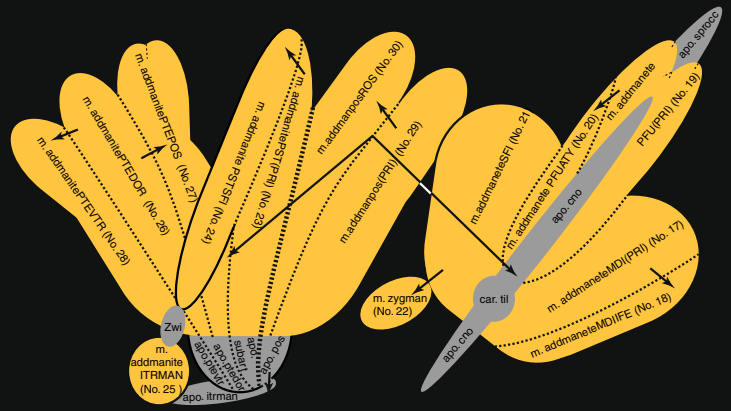
Introduction

The origin and interrelationship of turtles have been debated intensively (e.g., Rieppel 2008; Werneburg and Sánchez-Villagra 2009; Werneburg 2010; Scheyer et al. 2012) but, besides osteological and molecular data (e.g. Gaffney 1975; Gaffney et al. 1991; Joyce 2007; Lyson et al. 2010; Thomson and Shaffer 2010; Sterli 2010), only a few other character complexes have been used to reconstruct turtle phylogeny. I studied the jaw musculature as being a potential new source of phylogenetic information. A better understanding of soft tissue morphology (herein: musculature and tendons) may help in detecting relationships to skeletal transformations, functional adaptations, and evolutionary transitions of turtle and vertebrate heads in general.

The most prominent and most commonly studied muscular structures of vertebrates are those innervated by n. trigeminus (V), resulting in a great diversity of classificatory schemes (exemplified for turtles in Fig. 1). The trigeminal jaw musculature can be subdivided into three parts (Vetter 1878; Lubosch 1933, 1938a, b; Luther 1938; Rieppel 1981). First, the Constrictor primus (C_1) dorsalis homologue of Chondrichthyes, which is largely reduced in turtles [but see Fig. 2B/right, m. levator bulbi (No. 16) in *Dermochelys coriacea*, for numbers of muscular units see Fig. 1 and Werneburg (2011)] and spans between the palatal and the postorbital region of the cranium.

Second, the Constrictor primus lateralis homologue is represented by the jaw adductor musculature (Fig. 1), parts of which can be elongated extensively along the crista supraoccipitalis and crista squamosalis in several turtle groups (e.g., Kilius 1957; Gaffney 1975; Rieppel 1990). This extension and related changes in bite stresses were often assumed to be in the cause of the loss of various dermatocranial elements resulting in a caudal or/and a lateral emargination (e.g. Zdansky 1923–1925; Kilius 1957; reviewed

Lakjer 1926 (after Schumacher 1956c)							
M. adductor mandibulae							
M. add. mand. externus			M. add. mand. posterior			M. add. mand. internus	
M. supf.	M. med.	M. prof.				M. pseudotemporalis	M. pterygoideus
Ivo Poglayen-Neuwall (1953a) (after Schumacher 1956a)							
M. adductor mandibulae							
M. add. mand. externus			M. add. mand. posterior			M. add. mand. internus	
P. superficialis	P. media	P. profunda				P. pseudotemporalis	P. pterygoideus
Schumacher 1953/54							
M. adductor mandibularis							
M. adductor mandibularis externus			M. adductor mandibularis internus				
Portio superficialis	Portio medialis	Portio profunda	M. adductor mandibularis posterior (2-3 portions)		M. parietomandibularis (inconstant, 2 portions)	M. pterygoideus (2-3 portions)	
Schumacher (1954/55a)							
M. adductor mandibularis							
M. adductor mandibularis externus			M. adductor mandibularis internus				
M. zygomatico-mandibularis (only in "Trionyx")	Portio superficialis	Portio medialis	Portio profunda	M. adductor mandibularis posterior (caudalis)	M. adductor mandibularis posterior (rostralis)	M. parietomandibularis (inconstant)	M. pterygoideus
Schumacher (1954/55b)							
M. adductor mandibulae							
			M. add. mand. posterior		M. add. mand. anterior	M. add. mand. internus	M. pterygoideus
Schumacher (1956a)							
M. adductor mandibulae							
M. add. mand. externus			M. add. mand. internus				
M. zygomatico-mandibularis (only in trionychids)	P. superficialis	P. medialis	P. profunda	M. add. mand. posterior	M. add. mand. anterior		M. pterygoideus
M. parieto-mandibularis (inconstant)							
Schumacher (1956c)							
M. adductor mandibulae							
M. adductor mandibularis externus			M. adductor mandibularis internus				
M. zygomatico-mandibularis (only in trionychids)	P. supf.	P. med.	P. prof.	M. add. mand. post.	M. add. mand. ant.		P. pterygoideus
M. parieto-mandibularis (inconstant)							
Schumacher (1972)							
M. adductor mandibulae							
M. adductor mandibulae externus			M. adductor mandibulae internus				
M. zygomatico-mandibularis (trionychids)	Pars superf.	Pars media	Pars profunda	M. adductor mandibulae posterior	M. pseudotemporalis	M. intramandibularis	P. pterygoideus
Schumacher (1973)							
M. adductor mandibulae							
M. adductor mandibulae externus			M. adductor mandibulae internus				
M. zygomatico-mandibularis (only in trionychids)	Pars superficialis	Pars media	Pars profunda	M. adductor mandibulae posterior	M. pseudotemporalis	M. intramandibularis	M. pterygoideus
						<i>Cryptodires:</i>	Pars dorsalis
						<i>Pleurodires:</i>	Pars rostro-medialis
							Pars ventro-lateralis
							Pars ventro-posterior (<i>Podocnemis</i>)
Iordansky (1987, 1996)							
M. adductor mandibulae							
M. adductor mandibulae externus			M. adductor mandibulae internus				
M. zygomatico-mandibularis ("Trionyx")	pars postorbital		pars posterior	pars pseudotemporalis	pars digastricus: dorsal belly	pars pterygoideus	
	pars superior						
	pars inferior						
						dorsal belly (M. intramandibularis)	
Werneburg (2011)							



◀ **Fig. 1** Scheme listing different arrangements of m. adductor mandibulae structures (*Constrictor primus lateralis*) proposed for turtles in the literature. The abbreviations of words follow the style of Schumacher (1953/1954–1985). German words are translated. Note the “historically” shifting categorisation of m. adductor mandibulae externus posterior (“M. add. mand. posterior”) in relation to the other muscular structures; also, the parts of m. adductor mandibulae internus are categorised differently. *Below* The jaw musculature of turtles is presented following the lotus approach (modified from Werneburg 2011, see text for details)

by Werneburg 2012) and were correlated to a potential secondary loss of skull fenestration. The *Constrictor primus lateralis* consists of mm. adductor mandibulae externus et internus et posterior in turtles. The identity of the divisions of those muscles has often been discussed in such a way as to result in an inconsistent nomenclature in the literature (Fig. 1; Werneburg 2011). Lakjer (1926) developed a jaw muscle nomenclature for all Sauropsida based on the relative position of these muscles to the branches of the n. trigeminus (V) (e.g. see Holliday and Witmer 2007). Recently, Iordansky (1994) thoroughly proposed to categorise jaw muscles thoroughly based on the shape and differentiation of the tendinous framework that bears these muscles (see Werneburg 2010). On the one hand, the coronar aponeurosis, which attaches around the coronoid of the lower jaw, bears portions of m. adductor mandibulae externus (Fig. 2B) (No. 17–22). On the other hand, the m. adductor mandibulae internus (Fig. 1) and its portions (No. 23–28) insert into the subarticular aponeurosis, which attaches medially to the posterior part of the lower jaw. A separation of the pterygoidal aponeurosis from the subarticular tendon can occur in turtles [Fig. 1 (below)], bearing the pterygoideus parts (No. 26–28) of the m. adductor mandibulae internus complex. Often referred to as a part of the m. mandibulae internus in turtles (Fig. 2), and herein defined as a separated muscle (see Werneburg 2011), the m. adductor mandibulae posterior (No. 29–30) is situated between the mm. adductor mandibulae externus (No. 17–22) et internus (No. 23–28).

The third part of the n. trigeminus (V) innervated jaw musculature is the *Constrictor primus ventralis* homologue, which is represented by only three (No. 31–33) or four [see Werneburg (2011) for discussion on the identity of m. intra-mandibularis (No. 25)] muscular units that stretch between the dentaries in turtles.

Additional n. trigeminus (V) innervated muscular structures in turtles, not included in the present study, were previously observed in the nasal (No. 11–13) and ocular (No. 14–15) regions. The phylogenetic and developmental origin of those muscles is not entirely clear but may be derived from the *Constrictor primus dorsalis* and/or from cranial neural crest cell material (Werneburg 2011).

For turtles, there are at least nine different hypotheses for the arrangement of cryptodiran subgroups, and two different hypotheses for chelid (Pleurodira) phylogeny (e.g. summarised by Sánchez-Villagra et al. 2007; Scheyer 2007, 2009; Wilson and Sánchez-Villagra 2011). The pioneering work of

Gaffney and Meylan (1988) was based primarily on bone-characters resulting in a “basal” position of chelydrids (snapping turtles), incl. Platysternidae (big-headed turtles) within Cryptodira. Their morphological data supported a long-necked and a short-necked clade of chelid pleurodires. Other morphological works differ in the arrangement of Chelydridae, Platysternidae, and Chelonioidea (marine turtles) (Brinkman and Wu 1999; Joyce 2007; Werneburg and Sánchez-Villagra 2009). Since the late 1990s, genetic studies have consistently resulted in a stable chelid phylogeny consisting of distinct South American and Australasian clades (Shaffer et al. 2007; Krenz et al. 2005; Parham et al. 2006; Thomson and Shaffer 2010). All genetic studies also point to a “basal” position of Trionychia (soft-shelled turtles and *Carettochelys insculpta*) within Cryptodira. The relationship of all remaining cryptodire subgroups remained unclear; however, a molecular consensus is arising that Testudinoidea is the sister taxon to the remaining hard-shelled cryptodires with *Platysternon megacephalum* as a sister taxon to the Emydidae. Chelonioidea, Chelydridae, and *Dermatemys mawii* form the successive outgroup taxa of Kinosternidae.

Up to now, no phylogenetic study has considered the evolution of jaw musculature within turtles. Among other reasons discussed below, a diffuse nomenclature that lacked explicit primary homology assessments (de Pinna 1991) hindered phylogenetic inference. In a recent study, I reviewed all available literature references relating to the cranial musculature of turtles (Werneburg 2011). By atomising macroscopic structures into so-called muscular units, a traceable system on how to deal with muscular structures in phylogenetic research was developed. Having a plastic concept of muscular development and evolution in mind (Fig. 1 below; Werneburg 2011), the study presented here aims to test the phylogenetic signal of soft tissue characters in understanding turtle phylogeny and, more importantly, the correlation between the jaw musculature, the tendinous framework, and skull anatomy.

Materials and methods

Character definitions, coding strategies, and taxonomic sampling

I coded characters for those cranial muscles that are generally the most extensively studied among Tetrapoda, i.e. the n. trigeminus (V) innervated jaw musculature (No. 17–31) and the n. facialis (VII) innervated m. depressor mandibulae (No. 45). Information on other cranial muscles is available for only very few turtle species. Hence, I reduced the number of characters in order to have a broader sample of taxa, of which most characters are known.

The plastic nature of cranial musculature (Fig. 1, bottom; Werneburg 2011) is hard to code as discrete characters and

coding strategies that are used for bony structures are not applicable here. As such, several alternative character coding approaches could be developed, all of which have advantages and disadvantages (sensu Pleijel 1995). I decided on three alternative coding strategies, and estimated their influence on character distribution.

In coding-1, I present a largely phenetic/numerical approach coding mostly the absence (0) / presence (1) of muscle attachment to a bone (Appendices 1 and 2). Based on my own observations on *Emydura subglobosa* and on a comprehensive literature review (Werneburg 2011), I defined 202 characters (Appendix 1) for 42 turtle species (Table 1). The jaw muscles of all taxa were either described in detail and/or the depictions presented by the authors were sufficient to code character states clearly (Appendix 2, Fig. 2). The characters describe mainly the origin and insertion patterns of muscular units. Information on innervation pattern or fibre course would be valuable, but due to the restricted focus of several authors, such information is currently unavailable for most species.

In coding-2 (Appendices 3 and 4), I transferred the absence/presence-characters of coding-1 into an alternative shape, namely attachment of muscle X to bone A (0), to bone B (1), to bone C (2), etc. For all species, this coding strategy, with 74 characters, resulted in several multiple character states, which are not informative in phylogenetic analyses (Kornet and Turner 1999; Swofford 2003). As such, I present a third coding approach derived from coding-2, in which each composition of multiple character states of coding-2 was transferred into a new character state, coded as a letter (Appendix 4).

For the initial, coding approach (coding-1; Appendices 1 and 2), I decided on the following: where not explicitly stated, direct muscle fibre attachments were coded. Non-applicable character states were included in the character matrix (Appendix 2) only when a whole structure, such as a bone or a muscular unit, was missing. For turtles (not outgroup taxa), all possible attachments were coded. If one character state is coded for only one species, the character becomes parsimony uninformative in a phylogenetic reconstruction. However, I kept those uninformative characters in the matrix, firstly to retain as much information as possible to document the anatomy of terminal taxa and, secondly, to document that a muscle attaches to a particular site in contrast to none. In addition to a detailed character, e.g. “attaches laterally to element A (0) or not (1)”, I could have defined an absent/present character beforehand, e.g. “attaches to that element A (0) or not (1)”. This coding strategy would have (1) transferred the detailed characters to become non-independent, and (2) resulted in taxa that were coded as inapplicable for several characters. However, whether a muscle generally attaches to an element or not is often the only information available in the literature. By using the presented strategy, namely coding all possible attachments, I also indirectly present information on the general attachment to an element: if all possible attachment sites are coded as “not

Fig. 2 Examples of the diversity of jaw musculature in the major taxa of Testudines in lateral (*most images*) or dorsolateral (*f, i, k: right images*) view. Topology follows Shaffer (2009), which is the preferred topology herein. *Left pictures* each demonstrate the arrangement of mm. adductor mandibulae externus (No. 17–22) et depressor mandibulae (No. 45); the *right picture* for each clade demonstrates the arrangement of m. adductor mandibulae internus (No. 23–28) et posterior (No. 29–30). In *g* and *j*, the m. constrictor colli complex (No. 40–43, also indicated in *c*), m. intermandibularis (No. 31), and m. brachiomandibularis visceralis (No. 47) are also visible. *c* and *g* also show epaxial muscles. Images modified and redrawn from *a* Lakjer (1926: figures 151–153): *Amyda cartilaginea*, *b* Lakjer (1926: figures 51, 149–150): juvenil *Dermochelys coriacea*, *c* Rieppel (1990: figure 1A–B, E): *Chelydra serpentina*, *d* Poglayen-Neuwall (1953a: figures 1D, 2 G): *Kinosternon scorpioides*, *e* Lakjer (1926: figures 139, 228): *Chelonoidis denticulata*, *f* Poglayen-Neuwall (1953a: figures 2D, 3B): *Cuora amboniensis*, *g* Iordansky (1987: figures 1A, 1 F): *Trachemys terrapen*, *h* Schumacher (1954/55a: figure III2, 1954/55b: figure III2): *Platysternon megacephalum*, *i* Poglayen-Neuwall (1953a: figures 1B, 5A): *Mesoclemmys nasuta*, *j* Iordansky (1996: figures 2a, e): *Pelomedusa subrufa*, *k* *Podocnemis expansa*—Schumacher 1973: figure 9, Schumacher 1972: figure IX2): *Podocnemis expansa*. In *a* and *c*, the zygomatic arch is removed. In *b*, *h*, and *k* the postorbital and the temporal dermatocranial armour are removed

present”, the muscular unit generally does not attach to the element. If the structure attaches to an element, but the detailed attachment site is unknown, I coded the character as “?”. These coding strategies should be taken into consideration when interpreting character distribution on a phylogenetic tree.

For some species, I had information on different stages of development, including hatchlings, juveniles/subadults, and adults. To enable a certain amount of comparability, I coded the most adult condition of a character as documented in the literature.

The dorsad-leading part of the lower jaw is built mainly by the “coronoid area”. Possibly due to phylogenetic or ontogenetic variation, it may be represented by a coronoid process of the dentary or a coronoid bone. A clear definition of this “coronoid area” is mostly not documented in the literature; hence, I synonymised ‘coronoid’ and ‘coronoid process of the dentary’ in the character list as “coronoid” (Appendix 1).

Non-turtle taxa

Holliday and Witmer (2007: Tables 2, 3) presented a thorough and detailed summary of the jaw muscle attachments for three saurian clades—Lepidosauria, Crocodylia, and Neornithes—resulting from a literature review and their own observations. As the aim of my study was to observe the distribution of muscular characters within Testudines, I handled those three taxon summaries as terminal taxon descriptions. This decision leads, on the one hand, to a limitation in that the diversity in those clades, especially in lizards and birds, is not represented accurately, and the taxon summaries of Holliday and Witmer (2007), which are not based on a phylogenetic analysis, may therefore not necessarily represent the ground pattern of the respective groups. As these latter authors had a different aim

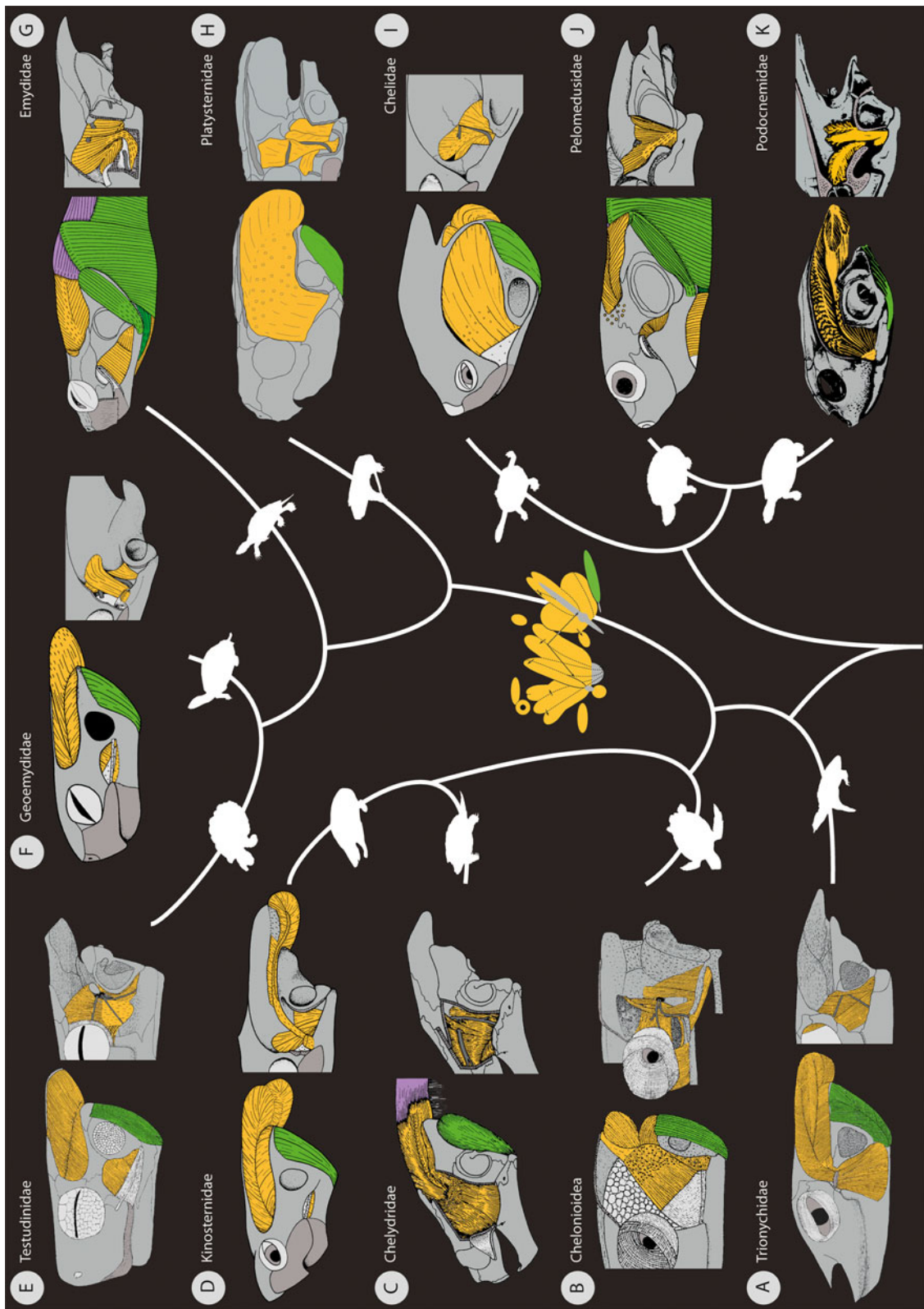


Table 1 Species analysed for phylogenetic analyses and the related references in chronological order. Taxonomic synonyms follow Fritz and Havaš (2007). For phylogenetic arrangements of the taxa in the preferred topology see Fig. 3

Species	References
Lepidosauria + Crocodylia + Neornithes	Mivart 1867, Adams 1919, Lakjer 1926, Anderson 1936, Hofer 1950, Ing. Poglayen-Neuwall 1953, 1954, Ivo Poglayen-Neuwall 1953b, Fisher and Goodman 1955 (<i>Grus americana</i>), Webb 1957 (<i>Struthio camelus</i>), Haas 1973, Schumacher 1973, Busbey III 1989, Müller and Weber 1998, Holliday and Witmer 2007, Jones et al. 2009
Testudines	
<i>Amyda cartilaginea</i>	Lakjer 1926: " <i>Trionyx cartilagineus</i> "
<i>Caretta caretta</i>	Ivo Poglayen-Neuwall 1953a, Schumacher 1953/54, Wyneken 2001, 2003
<i>Chelodina longicollis</i>	Kesteven 1942-45, Ivo Poglayen-Neuwall 1953a [not following his descriptions on m. adductor mandibulae externus Partes superficialis et medialis (No. 17, 21): see Werneburg 2011 for discussion], Schumacher 1955a, b
<i>Chelonia mydas</i>	Meckel 1828, Hoffmann 1890, Lubosch 1933: " <i>Chelone</i> ", Schumacher 1953/54, Iordansky 1987, 1996, Wyneken 2001
<i>Chelonoidis denticulata</i>	Wiedemann 1803 and Meckel 1828: " <i>Testudo</i> ", Lakjer 1926: " <i>Tistudo tabulata</i> ", Hoffmann 1890
<i>Chelonoidis nigra</i>	Schumacher 1953/54, Edgeworth 1935: " <i>Testudo elephantopus</i> "
<i>Chelus fimbriatus</i>	Hoffmann 1890: " <i>Chelys</i> ", Ivo Poglayen-Neuwall 1953a, 1966, Lemell et al. 2002, 2010
<i>Chelydra serpentina</i>	Stannius 1854, Adams 1919, Ivo Poglayen-Neuwall 1953a, Gaunt and Gans 1969, Rieppel 1990
<i>Chrysemys picta</i>	Edgeworth 1911: " <i>Chrysemis marginata</i> ", Ivo Poglayen-Neuwall 1953a, Ashley 1962, Scanlon 1982
<i>Clemmys guttata</i>	Ivo Poglayen-Neuwall 1953a
<i>Cuora amboinensis</i>	Ivo Poglayen-Neuwall 1953a
<i>Dermochelys coriacea</i>	Rathke 1848 and Hoffmann 1890: " <i>Sphargis</i> ", Burne 1905, Lakjer 1926, Ivo Poglayen-Neuwall 1953a, 1954, Schumacher 1972, Wyneken 2001, 2003
<i>Dogania subplana</i>	Rathke 1848, Schumacher 1955a, b
<i>Emydura macquarii</i>	Kesteven 1942-45: " <i>macquariae</i> ", Schumacher 1955a, b: " <i>krefftii</i> "
<i>Emydura subglobosa</i>	Werneburg 2011
<i>Emys orbicularis</i>	Bojanus 1819-21 and Meckel 1828 and Rathke 1848 and Stannius 1854, Owen 1866 and Hoffmann 1890: " <i>Emys europaea</i> ", Hoffmann 1890 also: " <i>Testudo europaea</i> ", Rathke 1848 also: " <i>Emys lutaria</i> ", Ivo Poglayen-Neuwall 1953a, Schumacher 1955a, b, Iordansky 1987, 1996
<i>Eretmochelys imbricata</i>	Rathke 1848 and Hoffmann 1890 and Lakjer 1926: " <i>Chelone imbricata</i> ", Wyneken 2001, 2003, Edgeworth 1935
<i>Graptemys geographica</i>	Schumacher 1955a, b
<i>Graptemys pseudogeographica</i>	Ivo Poglayen-Neuwall 1953a, Iordansky 1996: juvenil specimen
<i>Hydromedusa tectifera</i>	Schumacher 1955a, b
<i>Kinosternon leucostomum</i>	Ivo Poglayen-Neuwall 1953a
<i>Kinosternon scorpioides</i>	Rathke 1848 and Hoffmann 1890: " <i>Terrapene tricariata</i> ", Ivo Poglayen-Neuwall 1953a
<i>Lissemys punctata</i>	Rathke 1848: " <i>Trionyx granosus</i> ", Gnanamathu 1937: " <i>Emyda vittata</i> ", George & Shah 1955, Schumacher 1955a, b: " <i>Trionyx punctatus</i> "
<i>Macrochelys temminckii</i>	Schumacher 1955a, b
<i>Malaclemys terrapin littoralis</i>	Ivo Poglayen-Neuwall 1953a: " <i>Malaclemys pileata littoralis</i> "
<i>Mauremys caspica</i>	Hoffmann 1890, Ivo Poglayen-Neuwall 1953a and Iordansky 1987, 1996: " <i>Clemmys caspica</i> "
<i>Mauremys rivulata</i>	Schumacher 1955a, b: " <i>Clemmys caspica rivulata</i> "
<i>Mesoclemmys nasuta</i>	Ivo Poglayen-Neuwall 1953a: " <i>Batrachemys nasuta</i> "
<i>Pelodiscus sinensis</i>	Ogushi 1913 and Iordansky 1987, 1996: " <i>Trionyx</i> ", Ivo Poglayen-Neuwall 1953a and Schumacher 1955a, b: " <i>Amyda</i> "
<i>Pelomedusa subrufa</i>	Rathke 1848, Hoffmann 1890: " <i>Pentonyx capiensis</i> ", Schumacher 1955a, b, Iordansky 1996
<i>Pelusios niger</i>	Ivo Poglayen-Neuwall 1953a
<i>Pelusios sinuatus</i>	Schumacher 1953/54
<i>Pelusios subniger</i>	Schumacher 1953/54
<i>Platysternon megacephalum</i>	Schumacher 1955a, b
<i>Podocnemis expansa</i>	Schumacher 1953/54, Tvarožková 2006
<i>Podocnemis unifilis</i>	Schumacher 1953/54
<i>Sternotherus odoratus</i>	Ivo Poglayen-Neuwall 1953a, Stannius 1854: " <i>Staurotypus</i> "

Table 1 (continued)

Species	References
<i>Terrapene carolina</i>	Lakjer 1926: " <i>Cistudo</i> ", Ivo Poglayen-Neuwall 1953a: " <i>triunguis</i> " [not following his descriptions on m. adductor mandibulae externus Partes superficialis et medialis (No. 17, 21)]
<i>Testudo graeca</i>	Hoffmann 1890: <i>Testudo craeca</i> et " <i>Testudo mauretunica</i> ", Rathke 1848, Burkard 1902, Thomson 1932, Lubosch 1933, Ivo Poglayen-Neuwall 1953a, Hacker 1954, Hacker and Schumacher 1954, Schumacher 1955a, b
<i>Trachemys scripta</i>	Meckel 1828: " <i>Emys serrata</i> ", Ivo Poglayen-Neuwall 1953a: " <i>Pseudemys</i> "
<i>Trachemys terrapen</i>	Iordansky 1987, 1996: " <i>Pseudemys terrapen</i> "

in their study, they did not present the variability of characters among and within species in detail. The use of particular species of birds, lepidosaurs, and crocodiles may have been the better solution. However, I am not an expert in those groups and by using the thorough description by Holiday and Witmer I prevent misinterpretations in these taxa whose anatomy is a subject in itself. It would have been preferable to use outgroup taxa whose characters are based on a phylogenetic analysis, but producing this data was beyond the scope of this paper. All these considerations should be kept in mind when interpreting the output of the phylogenetic analyses presented here. Further studies should address saurian muscle characters in detail, which will require a detailed literature review and phylogenetic analyses.

For Lepidosauria, I extended the dataset of Holliday and Witmer (2007) with the information on *Sphenodon punctatus* (Rynchocephalia) presented by Jones et al. (2009), who also documented intraspecific variation. Other recent papers on the jaw muscles of lepidosaurs and other tetrapods were published by Daza et al. (2011) and Johnston (2011a, b).

For information not presented in the study of Holliday and Witmer (2007), I considered some other studies as summarised in Table 1. For muscle ratio-data (Appendix 1: characters 2–8), I compared published figures on selected representatives, namely *S. punctatus*, *Iguana iguana* (Lepidosauria), *Alligator mississippiensis* (Crocodylia), *Grus americana* and *Struthio camelus* (Neornithes).

For muscle-homologisations among sauropsids, I refer mainly to Holliday and Witmer (2007). I did not revise the literature cited by the authors, but their homologisation appears to be well founded. Nevertheless, particularly the homology of the m. adductor mandibulae internus Pars pterygoideus dorsalis/m. pterygoideus anterior remains contested by developmental studies and observations of the innervation pattern (Iordansky 1964, 2008, 2010; Tvarožková 2006). Moreover, current knowledge of the jaw musculature of tetrapods is very limited. There are several detailed studies on particular taxa (e.g. Poglayen-Neuwall 1953a; Iordansky 1987; Haas 2001, 2003a, b; Diogo and Wood 2011); however, they have not yet resulted in a cranial musculature based phylogenetic reconstruction of all tetrapods. I suggest critically rechecking the very influential categorisation of Lakjer (1926) (e.g. Lubosch 1933; Rieppel 1981, 1987; Holliday and Witmer

2007; Jones et al. 2009), who separated the n. trigeminus (V) innervated jaw musculature of sauropsids into three parts. Comparative developmental studies (Edgeworth 1935; Rieppel 1990; Tvarožková 2006) will need to be expanded.

For *Sphenodon punctatus*, I homologised the ‘m. Pterygoideus Typicus Middle Medial’ (m.PtTyMM) of Jones et al. (2009) to the herein presented m. adductor mandibulae internus Pars pterygoideus posterior (No. 27) due to similarities in the attachment sites and the relative positions of both structures to the surrounding tissues.

Character mappings

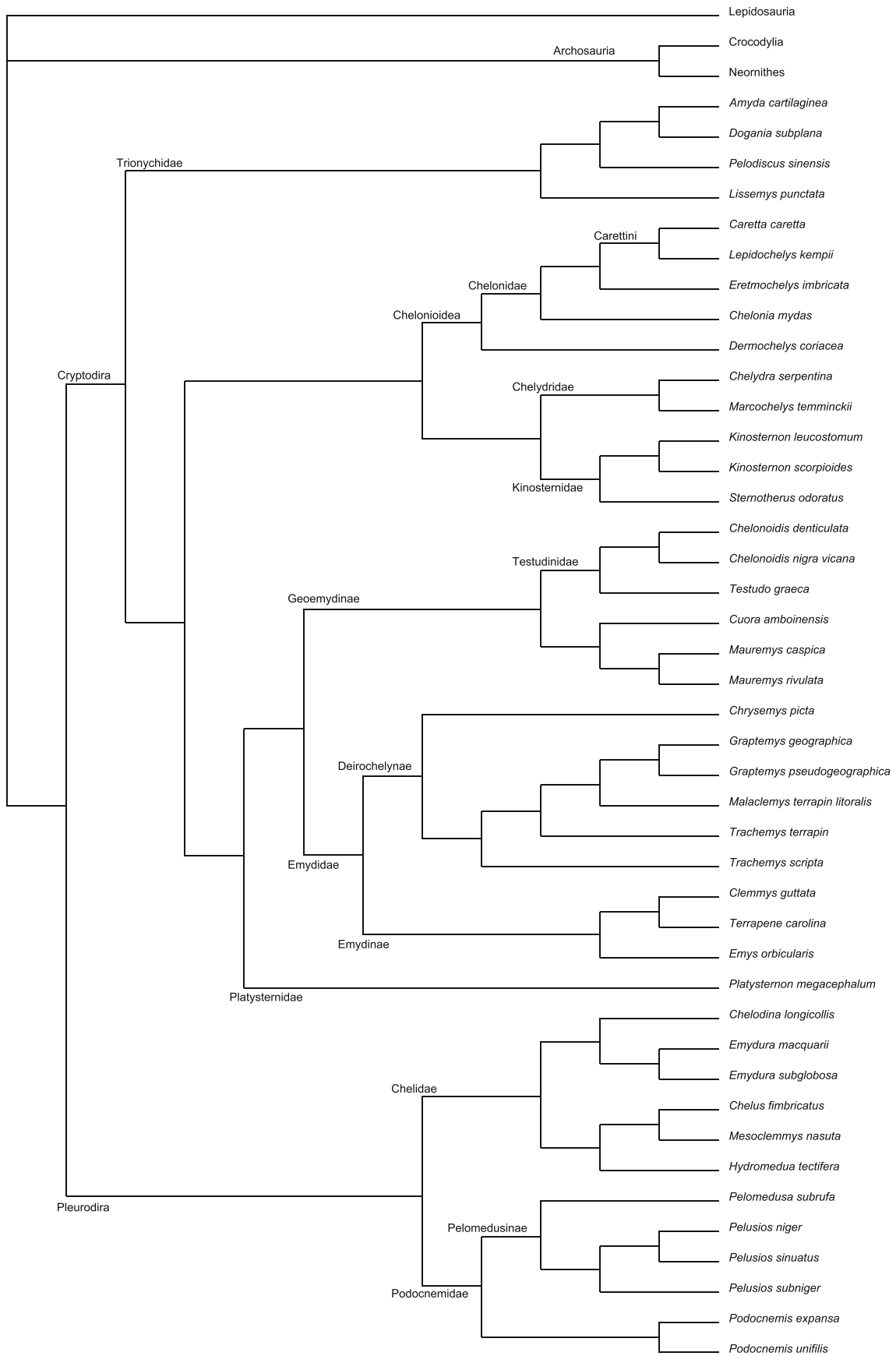
Using PAUP* (Swofford 2003), I plotted the coded soft tissue characters of my study onto a composed topology (Fig. 3) drawn with Mesquite 2.72 (Maddison and Maddison 2007). The topology derived from Shaffer (2009) for major turtle taxon relationships and Iverson et al. (2007) for the arrangements of “genera” and species (Fig. 3). I calculated Acctran and Deltran optimisations. As outgroup to Testudines, I defined [Lepidosauria + (Crocodylia + Neornithes)].

Results and discussion

Musculature and phylogeny

Relatively few studies (e.g. Winterbottom 1974; Estes et al. 1988; Hoyos 1990; Abdala and Moro 1996; Haas 2003a, b; Diogo 2004, 2007; Diogo et al. 2008a, b; Hertwig 2008; Werneburg 2009, Johnston 2011a) have extensively integrated soft tissue characters to reconstruct phylogenies. Among the reasons for this, these causes seem relevant (e.g. Hertwig 2008; Diogo and Abdala 2010; Werneburg 2011): (1) Inadequate, incomparable nomenclature resulting in terminological confusions in the literature; (2) uncertainties about homologies; (3) the challenge of gathering a broad set of information based on detailed manual dissections and histological serial sections; (4) an assumed, never tested, high degree of variability, (5) incomparability (non-homology) of muscular structures among clades.

It is generally believed that musculature is strongly exposed to adaptations such as locomotion, feeding behaviour,



◀ **Fig. 3** Composed, preferred topology of this study. Taxon arrangement follows the propositions of Shaffer (2009) and Iverson et al. (2007). A basal position of Trionychia within Cryptodira, as well as the sister group relationships of (Platysternidae + Testudinoidea), (Cheloniodea + (Chelydridae + Kinosternidae)) and (South American chelids + Australian chelids) are suggested

and ecophysiological demands (molluscivory vs. piscivory vs herbivory), resulting in several homoplastic character states in evolution. To a certain extent, this may be true, but hard tissue material underlies adaptive processes in development and evolution too, whilst muscles may keep a conservative condition (Costelli 1973: cited after Abdala and Moro 2003; Diogo and Abdala 2010). Moreover, the general shape of a muscular structure may stay constant within a major taxon, while the attachments—as mostly coded in the presented study—may be highly variable (Poglayen-Neuwall 1954), and vice versa.

One has to consider the taxonomic level when referring to the phylogenetic signal of any character including molecular, hard, and soft tissue material (e.g. Abdala and Moro 2003; Hertwig 2008). Only total evidence analyses, combining different kinds of data sets and using character congruence in the search, will be able to detect the best-fitting hypothesis for all of the available characters (e.g. Eernisse and Kluge 1993; Frost et al. 2006; Hertwig 2008; Asher et al. 2010). The presented contribution of jaw muscle characters of turtles did not aim to present a new phylogenetic hypothesis for the interrelationship of this group, and PAUP* reconstructions failed to recover any of the commonly accepted clades of turtle subgroups. As such, a character mapping on a given topology was performed (Fig. 3, Table 2).

The characters defined in the present study are gathered mainly from a literature review; hence, the descriptions of the respective authors had to be interpreted, and character states could be defined based only on the accuracy of those studies. For the initial coding, coding-1, I coded mostly simple binary characters.

In the future, the set of characters presented here could be combined with osteological and molecular characters to test hypotheses of turtle interrelationships and to estimate the phylogenetic signal and the relationship of different character sets on different levels of phylogeny. One may argue against such a combining method, as bone and muscle complexes could be non-independent. Muscular structures attach to defined areas of the bones and could influence their anatomy. On the other hand, skeletal shapes influence the extent of muscular structures. However, this problem usually exists when defining any morphological complex in a cladistic sense. Bones contact other bones, which, for this reason, cannot attach to another bone, etc.

Future studies could thoroughly define bone characters in the adductor chamber region with respect to the soft tissue characters defined herein. I refrained from simply combining my muscle data set with published data sets on cranial bone

characters (e.g. Gaffney et al. 1991; Joyce 2007). Regarding the details defined for soft tissue anatomy herein, comparably detailed bone characters, such as the ratios of bony contacts in the lower jaw or small tuberosities and “rippled” areas, which may be correlated to attachment sites (e.g. the articular), need to be defined. Moreover, a future study could focus on internal bone structures such as Sharpey’s fibres. With such a detailed approach, internal muscular characters could also indirectly be inferred (see also discussion below).

Each of the studies dealing with a set of muscle characters observed by the authors themselves (Abdala and Moro 1996, 2003; Hoyos 1990, 1998, 1999; Haas 2001, 2003a, b; Diogo 2008; Hertwig 2008) is *internally* consistent (scientifically and terminologically) and leads to traceable results. However, this has so far hindered a combined analysis for several vertebrate groups. The muscle nomenclature and the table of homology for turtles by Werneburg (2011) presented the first critical summary of a broad set of literature references and will enable broader analyses of comparative muscle anatomy in the future. I hope that the approach presented herein, using a plastic model of muscular structures (Fig. 1: below), will be useful when interpreting intra- and interspecific and evolutionary patterns of morphological structures of vertebrates in general.

Using atomised characters for phylogenetic reconstructions

Besides the character mapping, phylogenetic reconstructions were performed using PAUP*, resulting in different topologies (not shown). Those topologies deviate strongly from more plausible topologies and commonly accepted clades. Different considerations are required to adequately interpret the unsuccessful results of these phylogenetic reconstructions:

- (1) The more plausible results could be wrong. This, however, is less likely as many clades are confirmed both by osteological and molecular data;
- (2) The codings of characters presented herein might not be adequate for phylogenetic reconstructions. As pointed out by Pleijel (1995), Wiens (2000), and many others, the definition of character states indeed has an important influence on the output of a phylogenetic analysis. As reviewed by those authors, and many others, none of the coding strategies (e.g. atomised/multiple characters, absent/present characters, ordered characters) can be favored either on first principles or based on existing empirical evidence, so they recommend that the author of each analysis decides which strategy is best to use. The focus of the present paper is descriptive rather than phylogenetic. The illustration of the distribution of characters among turtle phylogeny presents hypotheses of primary homology and treatments, which are explicit and represent the first comprehensive study using muscle characters in turtle phylogenetic research. Future workers could revise

Table 2 Characters supporting Testudines, Pleurodira, and Cryptodira in the composed topology (Fig. 3) as inferred by the three alternative character codings. # Coding strategy; 1, 2, 3 coding strategy -1, -2, -3; *A* Acctran optimisation; *Ch* character; *D* Deltran, *C* consensus of Acctran and Deltran, *O* optimisation method

#	O	Ch	Character changes
Testudines			
1-3	C	1	M. levator bulbi (No. 16). Structure is present (1) ==> absent (0)
		3	Ratio of m. adductor mandibulae externus portions (No. 17-21). The origin site of Pars profundus (No. 19) extends more caudad than the origin site of Pars medialis (No. 17): other way around (0) -> yes (1)
1	C	10	M. adductor mandibulae externus Pars medialis (No. 17). Origin of extends to the rostral face of the quadrate: no (0) -> yes (1)
		33	M. adductor mandibulae externus Pars profundus (No. 19). Origin extends to the supraoccipitale: no (0) ==> yes (1)
		83	Coronar aponeurosis inserts to the dentary: no (0) -> yes (1)
		90	M. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23). Origin extends to the processus descendens parietalis: no (0) ==> yes (1)
		113	M. adductor mandibulae internus Pars pseudotemporalis superficialis (No. 24). Insertion extends around the insertion of the coronar aponeurosis (coronar aponeurosis, cartilago transiliens, surangular, and / or coronoid process): yes (1) ==> no (0)
		144	M. adductor mandibulae internus Pars pterygoideus ventralis (No. 28). Insertion extends to the angular: yes (1) ==> no (0)
		148	M. adductor mandibulae internus Pars pterygoideus ventralis (No. 28). Insertion extends to the pterygoid tendon: no (0) ==> yes (1)
		154	M. adductor mandibulae posterior (Pars principalis) (No. 29). Origin extends to the prootic: no (0) ==> broadly to the prootic (1)
		159	M. adductor mandibulae posterior (Pars principalis) (No. 29). Insertion extends to the angular: yes (1) ==> no (0)
A		26	M. adductor mandibulae externus Pars medialis (No. 17). Insertion extends to the posterior face of the coronar aponeurosis: no (0) -> yes (1)
		51	M. adductor mandibulae externus Pars superficialis (No. 21). Origin extends to the medial face of the quadratum: no (0) -> yes (1)
		73	M. adductor mandibulae externus Pars superficialis (No. 21). Insertion extends to the lateral face of the coronar aponeurosis: no (0) -> yes (1)
		79	Cartilago transiliens within coronar aponeurosis is absent (0) -> is present and has an articulation with: the pterygoid (1)
		111	M. adductor mandibulae internus Pars pseudotemporalis superficialis (No. 24). Insertion extends to the Zwischensehne [with this, also a m. intramandibularis (No. 25) is present]: no (0) -> yes (1)
		152	M. adductor mandibulae posterior (Pars principalis) (No. 29). Origin extends to the quadrate: broadly at anterior surface (0) -> restricted to the medial aspect of the anterior surface (1)
		157	M. adductor mandibulae posterior (Pars principalis) (No. 29). Insertion extends to the prearticular: no (0) -> yes (1)
		182	M. intermandibularis (No. 31) covers anterior region of constrictor colli complex (No. 40-43) superficially: no, homogen transition (2) -> muscles do not contact (only connected in area of median raphe (3)
		200	M. depressor mandibulae (No. 45). Insertion extends via a tendon to the retroarticular process: no (0) -> yes (1)
2-3	C	12	M. adductor mandibulae externus Pars medialis (No. 17). Insertion extends to the surangular (1) -> coronar aponeurosis (3)
		19	M. adductor mandibulae externus Pars profundus (No. 19). Insertion extends to the coronoid (0) -> coronar aponeurosis (1)
		24	M. adductor mandibulae externus Pars superficialis (No. 21). Origin extends to the lateral (0) -> medial (1) face of the squamosal
		41	M. adductor mandibulae internus Pars pseudotemporalis superficialis (No. 24). Insertion extends around the insertion of the coronar aponeurosis (coronar aponeurosis, cartilago transiliens, surangular, and / or coronoid process) (2) ==> to the Zwischensehne [with this, also a m. intramandibularis (No. 25) is present] (0)
2	C	14	M. adductor mandibulae externus Pars medialis (No. 17). Insertion extends to the dorsal face (0) -> lateral face (1) of the surangular
		17	M. adductor mandibulae externus Pars profundus (No. 19). Origin extends to the parietal (0) ==> supraoccipital (1)
		25	M. adductor mandibulae externus Pars superficialis (No. 21). Insertion extends to the surangular (2) ==> coronar aponeurosis (3)
		34	M. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23). Origin extends to the parietal (not processus descendens part) (0) -> processus descendens parietalis (1)
		35	M. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23). Insertion extends to the coronoid (7) ==> subarticular aponeurosis (0)
		51	M. adductor mandibulae internus Pars pterygoideus ventralis (No. 28). Insertion extends to the angular (2) ==> pterygoid tendon (6)
A		67	M. intermandibularis (No. 31) covers anterior region of constrictor colli complex (No. 40-43) superficially: no, homogen transition (2) -> muscles do not contact (only connected in area of median raphe (3)
3	C	17	M. adductor mandibulae externus Pars profundus (No. 19). Origin extends to the parietal (0) -> parietal and supraoccipital (A)
		23	M. adductor mandibulae externus Pars superficialis (No. 21). Origin extends to the dorsal and lateral (B) -> medial and anterior (D) face of the quadrate
		25	M. adductor mandibulae externus Pars superficialis (No. 21). Insertion extends to the dentary, coronoid, and surangular (A) ==> coronar aponeurosis (3).
		31	Coronar aponeurosis inserts to the dorsal (0) -> to the dorsal, medial, and lateral (B) face of the coronoid (process)
A		22	M. adductor mandibulae externus Pars superficialis (No. 21). Origin extends to the quadratum and quadratojugal (A) -> squamosal (2)

Table 2 (continued)

#	O	Ch	Character changes
			27 M. adductor mandibulae externus Pars superficialis (No. 21). Insertion extends to the dorsal and lateral (A) → lateral (1)
			67 M. intermandibularis (No. 31) covers anterior region of constrictor colli complex (No. 40-43) superficially: yes (1) → muscles do not contact (only connected in area of median raphe) (3)
Cryptodira			
1	C	53	M. adductor mandibulae externus Pars superficialis (No. 21). Origin extends to the anterior face of the quadratum: no (0) ⇒ yes (1)
		54	M. adductor mandibulae externus Pars superficialis (No. 21). Origin extends to the quadratojugal: no (0) ⇒ yes (1)
	A	61	M. adductor mandibulae externus Pars superficialis (No. 21). Origin extends to the postorbital: no (0) → yes (1)
		79	Cartilago transiliens within coronar aponeurosis is present and has an articulation with: the pterygoid (1) → with the quadratum / prootic (2)
		115	M. adductor mandibulae internus Pars intramandibularis (No. 25). Insertion extends to the bones of fossa (canalis) primordialialis: no (0) → yes (1)
	D	79	Cartilago transiliens within coronar aponeurosis is absent (0) → with the quadratum / prootic (2)
2-3	A	29	Cartilago transiliens within coronar aponeurosis has an articulation with: the pterygoid (0) → with the quadratum / prootic (1)
		37	M. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23). Insertion extends to the dorsal (0) → lateral (2) face of the cartilago Meckeli
		39	M. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23). Insertion extends to the dorsal (0) → ventral (1) face of the coronoid
		57	M. adductor mandibulae posterior (Pars principalis) (No. 29). The posterior aponeurosis (also “Sehnenspiegel”) inserts to prearticular (1) → articular (2)
		61	Subarticular aponeurosis. Insertion extends to the dorsal (0) → lateral (1) face of the cartilago Meckeli
2	C	45	M. adductor mandibulae internus Pars pterygoideus dorsalis (No. 26). Muscle fibres insert (next to insertion to the subarticular aponeurosis) to the articular (0) ⇒ prearticular (1)
3	C	43	M. adductor mandibulae internus Pars intramandibularis (No. 25). Insertion extends with a direct attachment to the bones of fossa (canalis) primordialialis (0) → inserts also by a tendon to that fossa (A)
Pleurodira			
1	C	42	M. adductor mandibulae externus Pars profundus (No. 19). Insertion extends to the dorsal face of the coronar aponeurosis: no (0) → yes (1)
		181	M. intermandibularis (No. 31). Fibre course of the posterior part of the muscle around 45 ° in posteromedial direction: no (0) ⇒ yes (1)
	A	23	M. adductor mandibulae externus Pars medialis (No. 17). Insertion extends to the lateral face of the surangular: no (0) → yes (1)
		25	M. adductor mandibulae externus Pars medialis (No. 17). Insertion extends to the lateral face of the coronar aponeurosis: no (0) → yes (1)
		85	Coronar aponeurosis inserts to the prearticular: no (0) → yes (1)
		136	M. adductor mandibulae internus Pars pterygoideus posterior (No. 27). Origin extends to the lateral face and/or to the dorsal face of the pterygoid: no (0) → yes (1)
	D	73	M. adductor mandibulae externus Pars superficialis (No. 21). Insertion extends to the lateral face of the coronar aponeurosis: no (0) → yes (1)
		79	Cartilago transiliens within coronar aponeurosis is absent (0) → is present and has an articulation with: the pterygoid (1)
		182	M. intermandibularis (No. 31) covers anterior region of constrictor colli complex (No. 40-43) superficially: no, homogen transition (2) → yes (1)
2-3	C	66	M. intermandibularis (No. 31). Fibre course of the posterior part of the muscle around 45 ° in posteromedial direction: no (0) ⇒ yes (1)
	A	48	M. adductor mandibulae internus Pars pterygoideus posterior (No. 27). Origin extends to the ventral face of or/and to the posterior face (0) → lateral face and/or to the dorsal face of the pterygoid (1)
2	C	27	M. adductor mandibulae externus Pars superficialis (No. 21). Insertion extends to the dorsal (0) ⇒ lateral (1) face of the surangular
		60	Subarticular aponeurosis. Insertion extends to the prearticular (2) → articular (3)
	A	30	Coronar aponeurosis inserts to the coronoid (process) (0) → dentary (1)
		43	M. adductor mandibulae internus Pars intramandibularis (No. 25). Insertion extends with a direct attachment to the bones of fossa (canalis) primordialialis (0) → inserts also by a tendon to that fossa (1)
	D	67	M. intermandibularis (No. 31) covers anterior region of constrictor colli complex (No. 40-43) superficially: no, homogen transition (2) → yes (1)
3	C	28	M. adductor mandibulae externus Pars superficialis (No. 21). Insertion extends to the dorsal (3) → lateral and ventral (D) face of the coronar aponeurosis.

Table 2 (continued)

#	O	Ch	Character changes
A	20	M. adductor mandibulae externus Pars profundus (No. 19).	Insertion extends to the medial (0) → posterior face (4) of the coronar aponeurosis
	22	M. adductor mandibulae externus Pars superficialis (No. 21).	Origin extends to the squamosal (2) → quadratum and squamosal (Q)
D	22	M. adductor mandibulae externus Pars superficialis (No. 21).	Origin extends to the quadratum and quadratojugal (A) → quadratum and squamosal (Q)
	27	M. adductor mandibulae externus Pars superficialis (No. 21).	Insertion extends to the dorsal and lateral (A) → lateral (1) face of the surangular

the matrices. My work may serve as a case study with which to understand and estimate how phylogenetically conserved or homoplastic this kind of character complex is at different levels of taxonomic hierarchy.

I present three alternative coding strategies, which are derived from each other. More sophisticated and pure cladistic character codings may result in more plausible results. Some of the multiple state characters coded herein could certainly be ordered. I refrained from that strategy to reduce ad hoc hypotheses of polarity. In addition, I rely mostly on a literature review. If I had dissected the specimens myself, a more integrative, less atomised/numerical character coding could have been produced, derived from a deeper impression of the actual material.

- (3) There might be much homoplasy in the characters defined for phylogenetic reconstructions. Among other functions, all characters are related to feeding and breathing and may be assumed to be influenced strongly by functional adaptations. But what characters are not in biology? Some taxa were ‘correctly’ reconstructed, which indicates that some characters defined herein are powerful enough to resolve at least different levels of the reptile phylogeny (e.g. Archosauria and Testudines as monophyla; groupings of some turtle “genera”; see Werneburg 2010).
- (4) The primary homology statements of the characters could be incorrect, i.e. they do not survive the test of homology in a phylogenetic reconstruction and are hence not secondary homologies (De Pinna 1991). Most characters coded herein refer simply to the attachment of a muscular structure to a bone. This kind of character atomisation might be misleading for phylogenetic reconstructions as the actual attachment of a muscular structure to a particular bone might be less important than a general attachment of that structure to a broader spatial appearance such as the otic region or the postorbital bony bridge in general. But this is a highly subjective approach, which, when coded by different researchers, will result in different phylogenetic reconstructions again.

As mentioned above, the cranial musculature behaves as a plastic structure in evolution and development (Fig. 1, Werneburg 2011), which is hard to code in a cladistic sense.

Using Remane’s (1950) approach of homologisation, I discussed the identities of muscular units extensively (Werneburg 2011) based on criteria such as *general* attachments, fibre courses, ontogeny, and innervation patterns. Given the certain validity of that homologisation (e.g. detailed innervations pattern are known for only a few species), one needs to consider the plasticity of muscle attachments. The material source of a muscle attachment can vary along the whole muscular unit and, depending on the timing and spatial interrelationship of the primordial muscle/bone contact, the actual fibres forming the attachments may not be homologous among species although derived from a homologous muscular unit. The example of jaw muscle attachments presented herein is a case study for the different levels of homology that need to be considered for phylogenetic reconstructions.

Considerations on character mapping

The character lists and matrices of the three different coding approaches can be found in the Appendix. For the clades Testudines, Cryptodira, and Pleurodira, Acctran and Deltran, optimisations and the consensi of both for all coding strategies can be found in Table 2.

Pattern cladists dealing with muscle characters “just” run their analyses and listed the resulting derived characters for each node in the resulting phylogenies (e.g. Abdala and Moro 1996). Those trees do not resample any other topologies, which show overlappings between osteological- or molecular-based clades. The characters listed in those papers are rarely cited or discussed in an evolutionary context.

It is a common strategy to plot data, of which the phylogenetic signal is hard to estimate (e.g. Winkler 2006; Scheyer 2009), onto a topology and to discuss those characters as given apomorphies due to the lack of more precise phylogenetic support. However, Assis and Rieppel (2011) criticised “the mapping of morphological characters onto nodes of a molecular tree [as this strategy] results in an empirically empty procedure for synapomorphy discovery”. This is because morphological characters are not incorporated into the molecular data matrix in those cases. As such, I should refrain from using the term “apomorphy” in the context of my analysis and rather use a term like “diagnostic character”. Nevertheless,

including 202 or only 74 morphological characters into a matrix of several tens of thousand molecular characters would not change the arrangement of the resulting topology drastically, as such and in actual fact, one finds apomorphies by character mapping.

In plotting anatomical characters onto a more reasonable topology (in which more overlappings of clades derived from different datasets exist), a sound discussion is possible in a traditional, morphological matter. By just following a cladistic approach, no sensible biological discussion is possible with the kind of data presented herein. I am “just” interested in character distribution on given topologies and its possible meaning for the evolution of the adductor chamber of turtles.

The character mappings of the three alternative codings enable different views on character distribution (Table 2). Each mapping reveals different perspectives on how and in which contexts the characters evolved. Certainly, the binary characters of coding 1 present only the gain or loss of an attachment in a phylogenetic framework. The multiple character states of coding 2 result in several equivocal character distributions. In coding-2 and -3, the descriptive information for the pure anatomical distribution of characters is lost.

The position of turtles within Sauropsida

I refrain from discussing the output of the phylogenetic analysis for non-turtle taxa in detail as they form the outgroup to Testudines entirely. As mentioned above, I used the data of Holliday and Witmer (2007) to score these groups, which represents a summary of a literature review rather than the result of a cladistic analysis. Although this may have a crucial influence on the character distribution, the strikingly different muscle anatomy of the major saurian taxa (Lakjer 1926; Poglayen-Neuwall 1953, 1954; Poglayen-Neuwall 1953b; Fisher and Goodman 1955; Webb 1957; Iordansky 1964, 2000; Haas 1973; Schumacher 1973; Jones et al. 2009) is well reflected in the output of my character mapping.

The high number of characters supporting Testudines in the character mapping (Table 2) indicates major rearrangements of the n. trigeminus (V) innervated jaw musculature within Sauropsida—resulting in a Lepidosauria/Archosauria and a Testudines morphotype, both of which could possibly reflect a sister group relationship of those taxa. Based on my exclusively neontological data, I cannot test whether Testudines evolved within one of the fossil “Anapsida”-clades, on the stemline of Sauria, or even on the stemline of Diapsida (sensu Werneburg and Sánchez-Villagra 2009). A position of turtles within Diapsida on the other hand must also have come along with crucial rearrangements of the jaw musculature, the origin of which cannot be detected with the characters defined herein.

The arrangement of jaw muscle characters as observed in the presented study does not help addressing the question of

the origin of temporal bone arrangements in turtles, as the ground pattern of jaw muscle *attachments* to the temporal region of Testudines did not reveal a strongly divergating shape when compared to other sauropsids. Assuming the Lepidosauria/Archosauria jaw muscle morphotype to be correlated to the diapsid skull condition, turtles either evolved within “Anapsida” or completely rearranged a diapsid dermatocranial armour in concert with a changed jaw muscle arrangement (and vice versa).

Following the preferred topology for turtle interrelationship (Fig. 3, for characters see Table 2), character 83 (coding-1) and character 31 (coding-3) for Testudines may be of particular relevance when interpreting the origin of turtles: the coronar aponeurosis gains an insertion to the dentary and the attachment to the coronoid is expanded. These character changes could have evolved based on a complete rearrangement of the whole m. adductor mandibulae externus complex in turtles when compared to that of the saurian taxa, implying a changed angle of bite forces and possibly a complete rearrangement of muscle precursors in early development of the reptiles. This would support my former hypothesis of a non-homology of the *portions* of m. adductor mandibulae externus among Sauropsida (Werneburg 2011). In concert to this, many characters of coding-2 and -3, such as a changed origin of m. adductor mandibulae externus Pars profundus (No. 19) (character 17) or the changed origin of Pars superficialis (No. 21) on the quadrate (character 24), can also be listed.

The muscular characters as defined herein are not intended to reconstruct the origin and interrelationship of the major turtle taxa using PAUP* (Swofford 2003). At a lower taxonomic level, e.g. “genera”, those characters may be of phylogenetic relevance. Gross muscle shape, not extensively coded herein could, on the other hand, be of particular relevance for turtle interrelationship (see Kiliyas 1957). The discussion of the present study on the phylogenetic signal of the jaw musculature (see above) should not be applied as a general feature to other vertebrate taxa and muscles, as, when compared to other vertebrate groups, turtles show a high degree of cranial diversity, which evolved relatively recently (e.g. Zdansky 1923–1925; Gaffney 1979; Joyce 2007; Werneburg 2012). Among other vertebrate taxa, muscle anatomy contains much phylogenetic signal (e.g. review by Diogo and Abdala 2010; Diogo and Wood 2011).

Lyson et al. (2010) used only the “basal” most fossil turtle species known to reconstruct their amniote phylogeny, resulting in a position of turtles deep within Parareptilia and hence outside of the Sauria-clade. If the latter authors had integrated extant turtle taxa, a high degree of phylogenetic noise would be imaginable, resulting in a differently reconstructed position of turtles within amniotes. This is again because emarginations are a secondary phenomenon among extant turtles involving a high degree of derived jaw muscle characters (Table 2). Future studies using jaw muscle

characters to reconstruct tetrapod phylogeny should be careful when either using turtles with emarginations or using marine turtles with their mostly complete temporal armour, including related secondary jaw muscle arrangements, which was acquired secondarily (Jones et al. 2012).

The evidence from fossils to interpret jaw muscle evolution in turtles

In general, Testudines are well defined by several derived characters that characterise them as a monophyletic clade (Mickoleit 2004). To what degree most of those characters can be discussed as autapomorphic for all Testudinata [Testudines (Pancryptodira + Panpleurodira) + stem Testudines; terminology sensu Joyce et al. 2004] is uncertain, as fossil preservation is often limited and as soft tissue characters obviously cannot be studied in extinct taxa. This is also true for extinct and stem taxa of groups of lower taxonomic level (Gaffney 1979). However, for the presented study, the recent discovery of the most “basal” known stem turtle *Odontochelys semitestacea* (Li et al., 2008: 497) is important: “*The skull of Odontochelys shows an elongate pre-orbital region forming a pointed snout [...]. Small and peg-like, pointed teeth are present on premaxilla, maxilla, dentary, pterygoid and vomer [...]. The jugal shows a rounded posteroventral margin, indicating a weak ventral excavation of the cheek. The skull table is not excavated posteriorly. The temporal region is not fenestrated. Distinct basiptyergoid processes form an open basicranial articulation with the pterygoids. The pterygoid shows a distinct transverse process that may have separated a subtemporal fenestra from a sub-orbital fossa [...]. The quadrate is weakly concave posteriorly; a cavum tympani is absent.*” To what degree *O. semitestacea* shows a derived or juvenile condition (Li et al. 2008; Scheyer et al. 2012) cannot be discussed adequately herein. The presumably second most “basal” turtle is *Proganochelys quenstedti* (Gaffney 1990), which, together with (and as a sister taxon to) the remaining Testudinata, is sister to *O. semitestacea* (Li et al. 2008; Lyson et al. 2010). Following the study of Sterli and de la Fuente (2010), the following evolutionary changes have to be considered for the adductor chamber in turtles:

(1) An akinetic skull enabled by a suturing of the basitrabecular (basiptyergoid) process and the pterygoid first evolved within—and not in the ground pattern—of Testudinata. The plesiomorphic condition of a basicranial kinesis still can be found in the basal-most turtles *P. quenstedti* and *O. semitestacea*. An additional plesiomorphic condition for both species (at least for *P. quenstedti*) may be the condition that the opisthotic is tightly sutured to the squamosal.

(2) Testudines and some advanced stem Testudines (*Meiolania platyceps*, *Mongolochelys efremovi*, *Kallokibotion bajazidi*) [together simplified discussed as ‘advanced testudinates’ herein; not to be confused with crown Testudines] show a complete reduction of the basitrabecular process. They also show the tendency to close the interpterygoid vacuity completely, a closure of the foramen nervi trigemini by the processus inferior parietalis, various kinds of temporal emarginations, caudad extensions of the crista supraoccipitalis, and the presence of an otic, or pterygoid trochlearis system (see Sterli and Joyce 2007). Sterli and de la Fuente (2010) argued for a strong evolutionary association of several of those characters in order to evolve a more effective bite performance in correlation with a growing “volume” of the adductor muscles.

Considering these anatomical features, muscle characters regarding the attachment sites of the adductor musculature (Table 2) are possibly only valuable for ‘advanced testudinates’. The caudad extension of m. adductor mandibulae externus Pars profundus (No. 19) over the Pars medialis (No. 17) (all codings: character 3) and the origin of the former on the supraoccipital (coding 1: character 33; coding 2+3: character 17) may be explained as relating to the emargination and/or caudad extension of the crista supraoccipitalis (Kiliyas 1957).

The secondary lateral braincase wall of turtles, formed by a laminar epiptyergoid and/or an extended processus descendens parietalis, first occurs in Testudines. In the more basal Testudinata, the epiptyergoid is quite small and the processus descendens parietalis rostrally does not extend over the anterior border of the prootic (Sterli and de la Fuente 2010). Consequently, the extension of m. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23) to the broad processus descendens parietalis (coding 1: character 90; coding 2: character 34) should be interpreted as a derived character of Testudines.

As mentioned above, a cavum tympani is absent in *O. semitestacea* (Li et al. 2008). Although slightly concave in all early Testudinata, Sterli and de la Fuente (2010) determined that a particularly enlarged otic region first occurs just below the appearance of the Testudines clade. Muscle characters regarding the newly gained origination sites of the mm. adductor mandibulae externus (No. 17, 21) et posterior (No. 29) portions to the quadrate or the prootic may be correlated to the acquisition of the enlarged otic region within Testudinata (coding 1: characters 51, 154; coding 3: character 23). Also the attachments of m. adductor mandibulae externus Pars superficialis (No. 21) to the squamosal (coding 2-3: character 24), the latter of which has a changed relative position in the cranium, may be correlated to this feature. However, detailed observations of this skull region among Reptilia are needed to possibly detect autapomorphic arrangements of related bones already in the ground pattern of Testudinata.

Several characters for turtles resulting from my character mapping concern the insertion sites of adductor muscles to the tendons and the lower jaw (Table 2). Compared to the remainder of the skull, no crucial evolutionary changes occurred in the arrangement of the posterior lower jaw bones of taxa on the stem lineage of Testudines (see Joyce 2007). This could, on the one hand, speak in favour of the hypothesis that related muscle characters have been conserved since the first occurrence of Testudinata. However, first, *O. semitestacea* still had teeth on the dentary and bones related to the palate region (Li et al. 2008) and, as mentioned above, a basicranial kinesis of the turtle skull is still assumed to have been present in *O. semitestacea* and *P. quenstedti*. Both features may require different muscle arrangements to those reconstructed in my analysis. Moreover, it has often been mentioned in the literature that the trochlearis system of extant turtles might enable particular bite performances (e.g. Schumacher 1956b), which are correlated to topographical changes in the attachment of jaw musculature. It was also reported that the coronar aponeurosis (Schumacher's works: "external tendon"; Iordansky 1996) of extant turtles, which is correlated to the trochlearis system herein, differs markedly when compared to other reptiles. It extends far caudally, forms an internal cartilaginous or bony sesamoid structure (cartilago/os transiliens: Schumacher 1956b; Ray 1959), and may contain several sheets (Werneburg 2011).

As most jaw muscle characters defined herein seem to be dependent on the presence of the unique trochlearis system, the information they provide for the phylogenetic origin of Testudinata within Amniota may be completely blurred. The trochlearis system as a whole includes the coronar aponeurosis containing one cartilago transiliens and one skeletal element. The skeletal elements, processus pterygoidei in pleurodires respectively processus otici in cryptodires, are built of non-homologous structures (Joyce 2007), whereas the coronar aponeurosis is homologous among all turtle clades (Iordansky 1994). Given Joyce's (2007: Fig. 19) hypothesis of a double articulation of the coronar aponeurosis in the last common ancestor of Pleurodira and Cryptodira, the two cartilagine transiliens certainly resulting, not mentioned by the author, must be interpreted as non-homologous structures between both extant taxa. This double articulation of the coronar aponeurosis could have resulted in a different arrangement of jaw muscle structures in the last Testudines-ancestor when compared to the last common ancestor of Testudinata. Before the two distinct extant trochlearis mechanisms evolved, a previous rearrangement of jaw muscles may hence have occurred in concert with the acquisition of a simultaneous pterygoid and otic related articulation. However, as will be shown below, gross anatomy of jaw muscles does not differ extensively among both extant turtle groups. This could indicate that a howsoever looking trochlearis system of the last common ancestor of Testudines already resulted in that particular jaw muscle anatomy, which was detected for

extant turtles herein. However, this also could indicate that the double articulation already did not have much influence on jaw muscle gross anatomy. In this context, the absence of muscle data for early amniotes, which exhibited the plesiomorphic anapsid morphotype, makes it difficult to generally declare the turtle jaw muscle anatomy as being derived. And, regardless of the elongated supraoccipital crest, it actually could resample most of the plesiomorphic condition, whereas saurian taxa could represent the more derived, temporal bar related jaw muscle arrangements.

The trochlearis systems and jaw muscle anatomy in extant taxa

Compared to Testudines, the two major extant turtle clades, Cryptodira and Pleurodira, are not obviously supported by jaw muscle characters (Table 2). It was a particular interest of this study to examine whether the attachments of the Partes pterygoidei of m. adductor mandibulae internus (No. 26–28) correspond to the distinguishing architecture of the jaw adductor chamber. In Pleurodira, the m. adductor mandibulae externus (No. 17–21) is bent around the processus trochlearis pterygoidei, whereas in Cryptodira the external jaw closing muscle is bent around the processus trochlearis otici (Schumacher 1956b; Gaffney and Meylan 1988).

Schumacher (1973) categorised the pterygoid-associated structures of the m. adductor mandibulae internus differently in pleurodires and cryptodires (Fig. 1). However, based on the criteria of homology presented by Werneburg (2011), and contrary to Schumacher (1973), all muscular units of both taxa can be clearly homologised, including three Partes pterygoidei (No. 26–28) in both taxa (Fig. 1). In the Acctran optimisation of the preferred topology, Pleurodira show an autapomorphic shift of m. adductor mandibulae internus Pars pterygoideus posterior (No. 27) to the lateral and/or to the dorsal face of the pterygoid (coding 1: character 136). In the same topology, a consensus character for Cryptodira is the expansion of the origin of m. adductor mandibulae externus Pars superficialis (No. 21) to the anterior face of the quadrate (coding 1: character 53; coding 2–3: character 48). Both characters may, with caution, be correlated with the differing trochlearis mechanisms in both taxa.

The m. intermandibularis (No. 31) of Pleurodira shows a rotation of its posterior fibres to a posteromedial direction when compared to the plesiomorphic condition (coding 1: character 181 = coding 2+3: character 66). In addition, the Deltran optimisation of the preferred topology suggests that the m. intermandibularis (No. 31) in Pleurodira changed from a continuous transition with the m. constrictor colli complex (No. 40–43) to a superficial covering of the latter muscle anteriorly (coding 1: character 182 = coding 2, character 67). I hypothesise that both characters regarding the m. intermandibularis (No. 31) anatomy might be correlated to the autapomorphic trochlearis mechanism in Pleurodira. The trochlearis system in that taxon is situated more anteroventrally when compared to

cryptodires and, as such, it may mechanically be linked with *m. intermandibularis* (No. 31). This muscle is situated nearby the trochlearis system and may need to stand particular forces in that taxon during jaw muscle activity.

The otic related trochlearis mechanism in Cryptodira, on the other hand, may have no effect on the anatomy of this mouth floor muscle—possibly balanced by the insertion of *m. intramandibularis* (No. 25) into the fossa primordialialis (Deltran for Cryptodira; coding 1: character 115 = coding 3: character 43). However, possibly independent, the *m. intramandibularis* (No. 25) reportedly occurs only in a few cryptodire taxa (Iordansky 2008; Werneburg 2011) and the distribution of this character depends strongly on the underlying phylogenetic topology. Joyce (2007) mentioned the loss of the sphenial bone without any reversals to be found in the lower jaw of Cryptodira and pelomedusoids. Hence, the resulting broader access to the primordial fossa may be correlated to the insertion of a *m. intramandibularis* (No. 25) in Cryptodira. However, to what degree trochlearis anatomy can be correlated to the arrangement of *m. intramandibularis* (No. 25) in Testudines should be tested by mechanical studies in the future.

As mentioned above, Cryptodira is characterised by an expansion of the origin of *m. adductor mandibulae externus Pars superficialis* (No. 21) to the anterior face of the quadrate, which could be correlated to the otic related trochlearis system in this taxon. The functional implications of the joint formed by *processus trochlearis otici* and *cartilago/os transiliens* may have an impact on the orientation of the ventral fibre arrangements in the dorsolaterally situated *Pars superficialis* (No. 21) towards an area ventrally to the course of the coronar aponeurosis.

Two additional points need to be mentioned that highlight the preliminary nature of my interpretations. (1) The proportions of the pterygoid portions (No. 26–28) were not coded herein due to insufficient data in the literature, but may contain some information to characterise both clades separately or even to interpret a possible, transitional, form as hypothesised by Joyce (2007) (see also Joyce and Sterli 2011). The proportions of the *m. adductor mandibulae externus* portions as coded herein (No. 17–21, characters 2–8), however, did not contribute to this discussion. (2) As shown by Hertwig (2005), the distribution of muscle fibre types within muscular structures of the jaw apparatus may contain an enormous information content regarding functional adaptations in vertebrates. Although the muscular units related to the trochlearis systems do not contain much phylogenetic information in their gross anatomy, as coded herein, the distribution of muscle fibres inside the muscular structure may clearly distinguish both turtle taxa. Future studies should check my hypothesis that the muscle fibre distribution (e.g. slow-twitch oxidative type-I-fibres vs fast-twitch glycolytic type-II-fibres) differs particularly in the portions of the external jaw adductor (No. 17–21/22). Enabled by the different extent of their attachment areas

(presence of *processus trochlearis pterygoidei*), the pterygoideus portions might differ only in their volume and fibre content as long as they have the same function. I maintain the same interpretive reservation regarding the functional morphology of all the muscular structures discussed in this paper.

Conclusions

Using the atomised definition of muscular units, traceable soft tissue characters were coded for phylogenetic analysis (character mapping). The concept of a plastic understanding of muscle anatomy and development (“lotus approach” of Werneburg 2011) served as the framework to understand the evolutionary changes of jaw muscle characters for the interpretation of muscle-bone interrelationship and their phylogenetic signal. Jaw muscle characters as defined in the presented study contain a high degree of phylogenetic noise on higher taxonomic level; however, at a “generic” level, characters may be informative.

Jaw muscle anatomy—as coded in the presented study—does not contribute to the debate on turtle origin within Tetrapoda, and phylogenetic reconstructions with jaw muscle characters using PAUP* (Swofford 2003) did not resolve the interrelationship of the major turtle taxa. However, a character mapping was performed for a combined topology (Shaffer 2009; Iverson et al. 2007) of turtle interrelationship. Herein, Chelidae (Pleurodira) are subdivided into an Australasian and a South American clade. Trionychia forms the sister taxon to all remaining cryptodires. *Platysternon megacephalum* forms the sister taxon to Testudinoidea; marine turtles, snapping turtles, and kinosternids form a monophyletic group.

Some clear correlations could be detected concerning the interrelationship of muscle attachment and skull anatomy; however, several muscle characters could not be correlated directly to skull shape. Internal anatomical features not documented herein, such as fibre orientation or muscle-fibre-type distribution, may contain information that could be correlated more directly to skull shape.

Using extant turtle taxa in broad-scale analysis of amniote phylogeny could result in a high degree of phylogenetic noise, as emarginations and related muscle characters evolved relatively recently. Although they have an anapsid morphotype, marine turtles do not represent the plesiomorphic skull and jaw muscle conditions of stem Testudines or of a possible anapsid ancestors.

The trochlearis system, which diagnoses the major extant turtle taxa (Pleurodira and Cryptodira) is hardly reflected by jaw muscle characters as defined herein. The general shape of jaw muscles remains constant among both groups. Fibre orientation of *m. intermandibularis* (No. 31) may, however, reflect the pleurodiran trochlearis system. In the future,

mechanical studies may be useful to understand differences between the jaw mechanics among turtle taxa.

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Appendix 1. Characters list 1 (phenetic approach; coding-1)

The character list corresponds to Werneburg (2010).

M. levator bulbi (No. 16)

- 1 Structure is absent (0), present (1)

Ratio of m. adductor mandibulae externus portions (No. 17–21)

The origin site of Pars ...

- 2 profundus (No. 19) extends more caudad than the origin site of Pars superficialis (No. 21): other way around (0), yes (1), +/- same (2).
 3 profundus (No. 19) extends more caudad than the origin site of Pars medialis (No. 17): other way around (0), yes (1), +/- same (2).
 4 medialis (No. 17) extends more caudad than the origin site of superficialis (No. 21): other way around (0), yes (1), +/- same (2).
 5 profundus (No. 19) extends more rostrad than the origin site of superficialis (No. 21): other way around (0), yes (1), +/- same (2).
 6 6 profundus (No.19) extends more rostrad than the origin site of medialis (No. 17): other way around (0), yes (1), +/- same (2).
 7 medialis (No. 17) extends more rostrad than the origin site of superficialis (No. 21): other way around (0), yes (1), +/- same (2).

Covering (from lateral view)

- 8 Pars superficialis (No. 21) covers Pars medialis laterally: not or only a bit (0), mostly or completely (1).

M. adductor mandibulae externus Pars medialis (No. 17+18)

- 9 Structure is clearly separated from Pars superficialis (No. 21): no (0), yes (1).

M. adductor mandibulae externus Pars medialis (No. 17)

Origin of extends ...

- 10 to the rostral face of the quadrate: no (0), yes (1).
 11 to the dorsal face of the quadrate: no (0), yes (1).
 12 to the capsule of the jaw joint: no (0), yes (1).
 13 to the squamosal: no (0), yes (1).
 14 to the postorbital: no (0), yes (1).
 15 to the quadratojugal: no (0), yes (1).
 16 to the opisthotic: no (0), yes (1).
 17 to the prootic: no (0), yes (1).
 18 to the supraoccipital: no (0), yes (1).
 19 to the parietal: no (0), yes (1).

Insertion extends ...

- 20 to the coronoid: no (0), yes, to the lateral border (1), yes, to the posterior border (2).
 21 to the dorsal face of the surangular: no (0), yes (1).
 22 to the medial face of the surangular: no (0), yes (1).
 23 to the lateral face of the surangular: no (0), yes (1).
 24 to the dentary: no (0), yes (1).
 25 to the lateral face of the coronar aponeurosis: no (0), yes (1).
 26 to the posterior face of the coronar aponeurosis: no (0), yes (1).
 27 to the ventral face of the coronar aponeurosis: no (0), yes (1).
 28 to the medial face of the coronar aponeurosis: no (0), yes (1).
 29 to the anterior face of the coronar aponeurosis: no (0), yes (1).
 30 to the dorsal face of the coronar aponeurosis: no (0), yes (1).

M. adductor mandibulae externus Pars medialis inferior (No. 18)

- 31 Structure is absent (0), present (1).

M. adductor mandibulae externus Pars profundus (No. 19)

Origin extends ...

- 32 to the parietal: no (0), yes (1).
 33 to the supraoccipitale: no (0), yes (1).
 34 via a tendinous framework to the supraoccipital: no (0), yes (1).
 35 to the postorbital: no (0), yes (1).
 36 to the opisthotic: no (0), yes (1).
 37 to the prootic: no (0), yes (1).
 38 to the exoccipital: no (0), yes (1).
 39 to the squamosal: no (0), yes (1).

Insertion extends ...

- 40 to the coronoid: no (0), yes (1).

- 41 to the medial face of the coronar aponeurosis: no (0), yes (1).
 42 to the dorsal face of the coronar aponeurosis: no (0), yes (1).
 43 to the ventral face of the coronar aponeurosis: no (0), yes (1).
 44 to the lateral face of the coronar aponeurosis: no (0), yes (1).
 45 to the posterior face of the coronar aponeurosis: no (0), yes (1).
 46 to the anterior face of the coronar aponeurosis: no (0), yes (1).
 47 to the rictal plate: no (0), yes (1).
 48 to the dentary: no (0), yes (1).

M. adductor mandibulae externus Pars profundus atypica (No. 20)

- 49 Structure is absent (0), present (1).

M. adductor mandibulae externus Pars superficialis (No. 21)

Origin extends ...

- 50 to the dorsal face of the quadratum: no (0), yes (1).
 51 to the medial face of the quadratum: no (0), yes (1).
 52 to the lateral face of the quadratum: no (0), yes (1).
 53 to the anterior face of the quadratum: no (0), yes (1).
 54 to the quadratojugal: no (0), yes (1).
 55 to the lateral face of the squamosal: no (0), yes (1).
 56 to the medial face of the squamosal: no (0), yes (1).
 57 to the anterior face of the squamosal: no (0), yes (1).
 58 to the dorsal face of the squamosal: no (0), yes (1).
 59 to the ventral face of the squamosal: no (0), yes (1).
 60 to the posterior face of the squamosal: no (0), yes (1).
 61 to the postorbital: no (0), yes (1).
 62 to the jugal: no (0), yes (1).
 63 to the parietal: no (0), yes (1).
 64 to the opisthotic: no (0), yes (1).
 65 to the posterior face of the quadrate / basioccipital: no (0), yes (1).
 66 to the prootic: no (0), yes (1).

Insertion extends ...

- 67 to the dentary: no (0), yes (1).
 68 to the lateral face of the coronoid: no (0), yes (1).
 69 to the posterior face of to the coronoid: no (0), yes (1).
 70 to the dorsal face of the surangular: no (0), yes (1).
 71 to the lateral face of the surangular: no (0), yes (1).
 72 to the ventral face of the surangular: no (0), yes (1).
 73 to the lateral face of the coronar aponeurosis: no (0), yes (1).
 74 to the anterior face of the coronar aponeurosis: no (0), yes (1).

- 75 to the posterior face of the coronar aponeurosis: no (0), yes (1).
 76 to the dorsal face of the coronar aponeurosis: no (0), yes (1).
 77 to the ventral face of the coronar aponeurosis: no (0), yes (1).
 78 to the rictal plate: no (0), yes (1).

Cartilago transiliens within coronar aponeurosis ...

- 79 is absent (0), is present and has an articulation with: the pterygoid (1), with the quadratum / prootic (2).

Coronar aponeurosis inserts ...

- 80 to the dorsal face of the coronoid (process): no (0), yes (1).
 81 to the medial face of the coronoid (process): no (0), yes (1).
 82 to the lateral face of the coronoid (process): no (0), yes (1).
 83 to the dentary: no (0), yes (1).
 84 to the surangular: no (0) yes (1).
 85 to the prearticular: no (0), yes (1).
 86 to the rictal plate: no (0), yes (1).

M. zygomaticomandibularis (No. 22)

- 87 Structure is absent (0), present (1).

M. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23)

Position in relation to the ramus maxillaris nervi trigemini (V₂)

- 88 lateral (0), medial (1), posterolaterally pierced by nerve (2)

Origin extends ...

- 89 to the parietal (not processus descendens part): no (0), yes (1).
 90 to the processus descendens parietalis: no (0), yes (1).
 91 to the epipterygoid (no epipterygoid in Pleurodira and *Dermochelys coriacea* sensu Joyce 2007): no (0), yes (1).
 92 to the prootic: no (0), yes (1).
 93 to the postorbital: no (0), yes (1).
 94 to the frontal: no (0), yes (1).
 95 to the interorbital septum: no (0), yes (1).

Insertion extends ...

- 96 to the subarticular aponeurosis: no (0), to the subarticular aponeurosis without m. adductor mandibulae internus Partes pterygoidei (No. 26-28) (at least connected to their tendon) (1), to the subarticular aponeurosis together with Partes pterygoidei (No. 26-28) (2).
 97 to the dorsal face of the cartilago Meckeli: no (0), yes (1).
 98 to the medial face of the cartilago Meckeli: no (0), yes (1).
 99 to the lateral face of the cartilago Meckeli: no (0), yes (1).

- 100 to the ventral face of the cartilago Meckeli: no (0), yes (1).
 101 to the surangular: no (0), yes (1).
 102 to the dorsal face of the prearticular: no (0), yes (1).
 103 to the medial face of the prearticular: no (0), yes (1).
 104 to the Zwischensehne [with this also a m. intramandibularis (No. 25) is present]: no (0), yes (1).
 105 to the dental: no (0), yes (1).
 106 to the articular: no (0), yes (1).
 107 to the dorsal face of the coronoid: no (0), yes (1).
 108 to the ventral face of the coronoid: no (0), yes (1).
 109 to the angular: no (0), yes (1).

M. adductor mandibulae internus Pars pseudotemporalis superficialis (No. 24)

Origin extends ...

- 110 to the parietal: no (0), yes (1).

Insertion extends ...

- 111 to the Zwischensehne [with this, also a m. intramandibularis (No. 25) is present]: no (0), yes (1).
 112 to the subarticular aponeurosis: no (0), yes (1).
 113 around the insertion of the coronar aponeurosis (coronar aponeurosis, cartilago transiliens, surangular, and / or coronoid process): no (0), yes (1).

M. adductor mandibulae internus Pars intramandibularis (No. 25)

Insertion extends ...

- 114 to the lateral face of the cartilago Meckeli: no (0), yes (1).
 115 to the bones of fossa (canalis) primordialialis: no (0), yes (1).
 116 inserts also by a tendon to f the fossa (canalis) primordialialis: no (0), yes (1).
 117 in a very long rostro-caudad extension: no (0), yes (1).

M. adductor mandibulae internus Pars pterygoideus dorsalis (No. 26)

Origin extends (next to the dorsal face of the pterygoid)

...

- 118 to the palatine: no (0), yes (1).
 119 to the processus descendens parietalis: no (0), yes (1).
 120 to the septum interorbitale: no (0), yes (1).
 121 to the postorbital: no (0), yes (1).
 122 to the jugal and maxilla: no (0), yes (1).
 123 to the frontal: no (0), yes (1).

Muscle fibres insert (next to insertion to the subarticular aponeurosis) ...

- 124 to the articular: no (0), yes (1).
 125 to the prearticular: no (0), yes (1).
 126 to the cartilago Meckeli: no (0), yes (1).

- 127 to the capsule of the jaw joint: no (0), yes (1).
 128 to the surangular: no (0), yes (1).
 129 to the angular: no (0), yes (1).
 130 to the coronoid: no (0), yes (1).
 131 to the pterygoid tendon: no (0), yes (1).
 132 via pterygoid tendon: to the internal tendon (as apo.pteDOR) (0) / separated insertion to the prearticular/canalis primordialialis (as apo.pteVEN) (1)
 133 to the rictal plate: no (0), yes (1).

M. adductor mandibulae internus Pars pterygoideus posterior (No. 27)

Origin extends ...

- 134 to the quadrate: no (0), yes (1).
 135 to the ventral face of or/and to the posterior face of the pterygoid: no (0), yes (1).
 136 to the lateral face and/or to the dorsal face of the pterygoid: no (0), yes (1).
 137 to the palatine: no (0), yes (1).

Insertion extends ...

- 138 to the articular (incl. retroarticular process): no (0), yes (1).
 139 to the surangular: no (0), yes (1).
 140 to prearticular: no (0), yes (1).

M. adductor mandibulae internus Pars pterygoideus ventralis (No. 28)

Origin extends (next to pterygoid) ...

- 141 to the palatine: no (0), yes (1).

Insertion extends ...

- 142 to the surangular: no (0), yes (1).
 143 to the coronoid: no (0), yes (1).
 144 to the angular: no (0), yes (1).
 145 to the prearticular: no (0), yes (1).
 146 to the articular: no (0), yes (1).
 147 to the subarticular aponeurosis: no (0), yes (1).
 148 to the pterygoid tendon: no (0), yes (1).
 149 to the rictal plate: no (0), yes (1).
 150 to the capsule of the jaw joint: no (0), yes (1).

M. adductor mandibulae posterior (Pars principalis) (No. 29)

Origin extends ...

- 151 Has two muscle heads in the origin area: no (0), yes (1).
 152 to the quadrate: broadly at anterior surface (0), restricted to the medial aspect of the anterior surface (1), ventrally (2)
 153 to the pterygoid: no (0), yes (1)
 154 to the prootic: no (0), broadly to the prootic (1), restricted to the lateral aspect of the prootic (2).

- 155 to the lateral face of the parietal: no (0), yes (1).
 156 to the quadratojugal: no (0), yes (1).

Insertion extends ...

- 157 to the prearticular: no (0), yes (1).
 158 to the articular: no (0), yes (1).
 159 to the angular: no (0), yes (1).
 160 to the surangular: no (0), yes (1).
 161 to the cartilago Meckeli: no (0), yes (1).
 162 to the coronoid: no (0), yes (1).
 163 to the coronar aponeurosis: no (0), yes (1).
 164 via posterior aponeurosis (also “Schnenspiegel”) to surangular: no (0), yes (1).
 165 via posterior aponeurosis to prearticular: no (0), yes (1).
 166 via posterior aponeurosis to articular: no (0), yes (1).
 167 via posterior aponeurosis to coronoid: no (0), yes (1).

M. adductor mandibulae posterior Pars rostralis (No. 29)

Origin extends (next to the anterior face of the prootic) ...

- 168 to the parietal: no (0), yes (1).

Insertion extends...

- 169 to the subarticular aponeurosis: no (0), yes (1).
 170 to the coronoid: no (0), yes (1).

Subarticular aponeurosis

Insertion extends ...

- 171 to the dorsal face of the cartilago Meckeli: no (0), yes (1).
 172 to the lateral face of the cartilago Meckeli: no (0), yes (1).
 173 to the coronoid (process): no (0), yes (1).
 174 to the dorsal face of the prearticular: no (0), yes (1).
 175 to the medial face of the prearticular: no (0), yes (1).
 176 to the articular: no (0), yes (1).
 177 to the surangular: no (0), yes (1).

M. intermandibularis (No. 31) ...

- 178 reaches the symphysis of the dentals: no (0), yes (1).
 179 a posterior trigonium is formed between m. intermandibularis and constrictor colli complex (No. 40-43): no (0), yes (1).
 180 contralateral muscles fuse in: a thin median raphe (0), a broad aponeurosis anteriorly (1), a broad aponeurosis posteriorly (2).
 181 fibre course of the posterior part of the muscle around 45° in posteromedial direction: no (0), yes (1).

- 182 covers anterior region of constrictor colli complex (No. 40-43) superficially: other way around (0), yes (1), no, homogen transition (2), muscles do not contact (only connected in area of median raphe (3), connected by connective tissue (4).

M. depressor mandibulae (No. 45) ...

- 183 has a bipinnated shape: no (0), yes (1).

Origin extends ...

- 184 originates with two muscle heads: no (0), yes (1).
 185 to the posterior face of the squamosal: no (0), yes (1).
 186 to the ventral face of the squamosal: no (0), yes (1).
 187 to the lateral face of the squamosal: no (0), yes (1).
 188 to the dorsal face of the squamosal: no (0), yes (1).
 189 to the posterior face of the quadrate: no (0), yes (1).
 190 to the lateral face of the quadrate: no (0), yes (1).
 191 to the medial face of the quadrate: no (0), yes (1).
 192 to the quadratojugal: no (0), yes (1).
 193 to the opisthotic: no (0), yes (1).
 194 to the jugal: no (0), yes (1).
 195 to the cornu branchiale-II: no (0), yes (1).

Insertion extends ...

- 196 to the articular: no (0), yes (1).
 197 to the angular: no (0), yes (1).
 198 to the surangular: no (0), yes (1).
 199 via tendon to the articular body: no (0), yes (1).
 200 via tendon to the retroarticular process: no (0), yes (1).
 201 via tendon to the angular: no (0), yes (1).
 202 via tendon to the surangular: no (0), yes (1).

Appendix 2. Character matrix 1 (phenetic approach; coding-1)

Character matrix, including all information gathered from literature and own observations, including not applicable data. The character coding corresponds to Werneburg (2010) with few modifications in characters 9, 80, 89, 96, 104, 114-117, 125, and 198 for *Caretta caretta* and *Lepidochelys kempii* following new data by Jones et al. (2012). Multiple characters states for all characters A=(0,1), B=(0,2), C=(1,2), D =(1,3), E=(2,3).

Lepidosauria

11210A0110 00100000A0 AA00?000AA 010000A011 AA0A000000 0000111010 1A0000AA0A 10?????00A
 ??0A0001A0 11000????? 1000001011 001????A01 000A0000AA ??0??????? 00A101?000 020000A110
 000?????--- ??????01A0 0E00??????? ?????1????? ??

Crocodylia

1000112111 0000000000 1010000000 0100000000 ??????0000 0111000000 0000000001 00000000??
 ??????0100 00000?????0 0000000000 001?????101 0001000010 ??0----- 010100?000 0000000110
 000?????--- ??????0012 0200??????? ??????1????? 00

Neornithes
 10A011C010 001000000? ???000000 0100000011 ?????0001 0100100000 00000011?1 1?????00?
 ??????0100 00000????? ???0000110 001----100 000?????? ??00000000 1????????? 000000????
 ???????--- ??????0A10 0100?????? ??????1???? ??

Amyda cartilaginea
 0110111011 1??0101000 1010000000 1110000001 11111??00? ??0010100 0000000001 010000000?
 ??????1111 00??020000 000000000- -----000 0000000000 --0----- 0000101-00 1101001100
 00000000--- 0001100?? ?210??10?? ?0??0??000 00

Caretta caretta
 0110210111 00A0A0000A 0000AA0AAA 0110000000 011000000A A0A10AA000 1110000?? ?A0000021
 1111000111 0000A00010 1001000001 100111011A 0001111000 1A01101100 A000110110 A001001000
 000AAA0--- ??????0?? ?2011000?? ?A00000011 00

Chelodina longicollis
 021011?001 00A0000000 11100A0A0A 0110110100 000010A010 10A00A0AAA 0001000000 001AA00A11
 A11AAA0101 -?0002???? 0000AA??- -----AA0 000A0000A0 --00010010 -----?--- 010CA01100
 0AA0000--- ?1110A111 1?0AA0A000 0010010000 00

Chelonia mydas
 011????110 0000000002 ???1010000 0110000000 0110000001 0001010000 1110000?? ?210000021
 1111000101 0000000010 110000000- -----11A 0001111000 1101101100 0000110101 1001000000
 0011110--- ??????0?? ?201100000 0000000011 00

Chelonoidis denticulata
 0110210011 1000000000 ?210????? ?21?????0 1100000001 0001010000 1000000001 ?100000021
 ??01000111 1100010000 000000000- -----11A 0000000000 110----- 1?????0?? 0?????1000
 0010000--- 0001000010 11000110?? ?0??0??000 00

Chelonoidis nigra
 0210210010 000000000? ???0110000 0110000000 0001000000 1001010000 1001000100 101000002?
 ??????0?01 0100020000 1??000000- -----11A 0000100100 0-0----- 0000000100 0100000100
 0000010--- 0000010?? ?201000011 0000011001 00

Chelus fimbriatus
 0010020010 1010-11001 1010111111 0010000100 0101000000 0000001000 0010000000 101000001?
 ??01??0100 -010020000 0000000001 010----111 0000000000 100----- ??????0?? 0012000100
 0010000--- 0000010111 A????????? ??????0?? ??

Chelydra serpentina
 0111010100 001001000? 0000110100 0110000000 1000100000 A0AA001A00 1AA000100? ??1A000A21
 10100A0201 0A1?A20000 000000000- -----1?0 0000000000 --0----- 0000001-00 A101001100
 0000000--- 000?00110 000010AA00 00000010000 00

Chrysemys picta
 02100100?0 0010000000 ???1110000 0110000000 1000100000 1011000000 010000???? ??????????
 ??????0101 0000020000 000100000? ??0101?00 0000000000 --0----- ???????-?? 0101000000
 0001?10--- ??????0100 03010?00?? ?0?00?0000 00

Clemmys guttata
 011?????0 0010000000 0010100000 0110000000 1000100000 1011000000 1100001000 001100002?
 ??????0201 000002???? ?????????? ???-???00 0000000000 --0----- ???????-?? 010100????
 ???????--- ??????0010 00??001000 00000?0?? ??

Cuora amboinensis
 01122101?0 0010010100 0000000100 0110000000 1000100000 1011000000 110000???? ??????????
 1100000201 000002???? ?????????? -----?00 0000000000 0-0----- ???????-?? 0101000000
 1000000--- ??????00?? ?20001?000 00??0??000 00

Dermochelys coriacea
 1121000011 10101A0002 0100AA0AA0 0A1000000A 0AA0AA0000 1011010000 1110000001 ?111000000
 0000000000 -10001???? 01?000000? ???-???A00 0000110000 110----- 1000000100 AA010A0001
 0000000??? 00000001A0 0E0A1000?? ?A000111?? ??

Dogania subplana
 0110110??0 010000000C 1011000000 0111111001 1101001000 1011010000 0101000110 0010100021
 1111101?11 0011020000 000000000- -----100 1100000001 --0----- -----?--- 0?????0000
 0010000--- 0011110?? ?2??0????0 0000010001 00

Emydura macquarii
 0210210000 0000000002 1?00000000 0110100000 0110000000 100-010011 1011110000 0000011011
 1111100?11 -011010000 01000?000- -----100 0001100000 110----- 0000000100 0102001100
 000???--- ??21??0?? ?210001000 0000010000 00

Emydura subglobosa
 0210210010 0000000000 0011000000 0110000001 0110000001 000-010100 0010100000 0010001011
 1100000101 -110010000 000000000- -----100 0000000000 110----- ??????0?? 0102100001
 1000000--- 0000010110 1D00101100 000000001? ??

Emys orbicularis
 0110121000 001001000A 00A0110A00 01100A0000 AAAAA00001 A0A1000000 1100000A0? ??1A000021
 1111100201 0000020010 011000??- -----100 0000001000 --0----- 1000000-01 010A00?A00
 0001010--- ??????0A10 0300101000 00000?0?? ??

Eretmochelys imbricata
 0110210111 1000100000 ?210?1???? 0110000000 0000100000 0001010000 1000001001 ?100000021
 00010?0111 0000010000 000000000- -----101 0000100000 --0----- 1?????-?? 1101001100
 0010000--- 0001000110 0?00100000 00??0??000 00

Graptemys geographica
 0110210011 0000000000 ???0110000 0111010000 1000100000 1000010000 0000000000 0010100021
 1111100?01 000002???? 000000000- -----100 0000001000 --0----- 1000000-01 0102000000
 0000010--- 0101100?? ?2?00001000 0000000001 00

Graptemys pseudogeographica
 0?????????1 001000000? ?????????? 0110000000 0000100000 1011000000 010000???? ?211??????
 ??????0101 0000020000 000100000? ??0101?00 0000000000 --0----- ???????-?? 010100????
 ???????--- ??????0?? ?????????? ??????0?? ??

Hydromedusa tectifera
 0210111001 0000000000 0010110000 0110110000 ?????????0 1000010001 0000000000 00?????011
 1111100111 -011021000 000000000- -----010 1000100000 --0----- -----?--- 0101-01001
 0010000--- 1001100?? ?200111001 0010000001 00

Kinosternon leucostomum
0?????????1 0010000110 0000001001 0111000000 1110100000 1011010100 1100001001 1011000021
11000?0201 000002???? ????0????? -----?00 0000000000 --1----- ????-?? 0101000100
0000010--- ??????00?? ?????????? ??????0?? ??

Kinosternon scorpioides
0110110??1 0010000110 0000001001 0111000000 1110100000 1011010100 1100001001 101100002?
?????0201 000002???? ????0????? -----?00 0000000000 --1----- ????-?? 0101000100
0000010--- ??????00?? ??????1000 00?0?0000 00

Lepidochelys kempii
0110??0?01 0000000000 0100000000 0110000000 1000000000 0011000100 110101???? ????000021
11????0101 0000000000 000100000? ???1110101 0000100000 1?0??????? 1000000100 1102001100
0000000??? ??????01?0 02??111000 00000?0?? 00

Lissemys punctata
0110??0?11 0000000000 0100000000 0110000000 1000000000 0011000101 110101???? ????000021
11????0111 0000020000 000?00000- -----101 0000000000 1-0----- 1000000100 1102001100
0000000--- ??????01?0 02??111000 00000?0?? 00

Macrochelys temminckii
01101100?1 010000000C 1011000000 0111111001 1101001000 1010010000 0001000110 0010100021
11AAA01111 0011020000 000000000- -----A00 A10000000A --0----- ??????-?? 0000001100
00A0001001 0011110110 ?310001100 0000010001 A0

Malaclemys terrapin litoralis
01102101?1 0000000000 ????0010000 0010011000 1011000000 1000010000 0000000??0 101000012?
1111100?01 0111020000 0000000101 100101?010 1000000000 --0----- -----?--- 0100001100
0000000--- 0011110000 0201100101 0001011000 00

Mauremys caspica
011??????1 0010000000 0011111000 0110000000 1001100000 1011000000 1010001000 0011000021
?????0201 000002???? ????0????? ???-???00 0000000000 --0----- ??????-?? 0101000000
000??10--- ??????00?? ??????1000 00?0?0000 00

Mauremys rivulata
0110??01?1 0010000000 0010100000 0110000000 1000100000 1011000000 1100000000 0011000021
110?0?0201 000002???? ????0????? ???-???00 0000000000 --0----- ??????-?? 010200????
?????--- 11010100?? ????01?0?? ?0?0?0000 00

Mesoclemmys nasuta
0?????1?1 0000000000 ????0110000 0110010000 1110000000 0001010000 010000000? ??????12?
1111100?01 0000021010 001000100? ???-???110 0000100100 0-0----- 0001010100 0102001100
0010000--- 0000010?? ????0001000 00000?0?? ??

Pelodiscus sinensis
0010?10101 00?00?0000 0000001000 0110000000 0100100000 101-?????? 001101??1 ????1?011
??00010201 -000020000 010000000- -----?00 0000000000 --0----- ??????-?? 010100?1?1
00?0000--- ??????01?0 00000?1000 00?0?00?? ??

Pelomedusa subrufa
0110110011 01100A000C 1011A0000A 0111AAA001 11A1A0AA0 10AA0A0000 0A0A00AAA? ?1AA00021
11AAA012A1 000A020000 000000000- -----AA0 AA00000001 0-00010100 1010000-00 0A01001100
0000000--- 0011110?? ?1A011000 0000010101 01

Pelusios niger
0210020001 0100000000 0010110000 0110000001 0101000000 1001010000 1100000100 001000101?
1110100101 -000010000 010000000- -----010 1011100001 110----- 1010000110 010110?100
1010000--- 100101001? 0300011000 0010010001 00

Pelusios sinuatus
0210210001 00?00?0000 0000?01?00 0110000000 0100100010 1011010?01 01010?0000 0000000111
?????10101 -000020000 000001000- -----?00 0000000000 --0----- ??????-?? 1101A00000
0000100--- ??????0110 13????????? ?????????? ??

Pelusios subniger
0210210001 000000000? ????0110000 0110001000 1000000000 1001010000 1010000100 1010001011
1111000101 -100020000 0??000000- -----?00 0000000000 --0----- 0000110-00 0001000001
000??0--- ??????0?? ????01010000 10100100?? ??

Platysternon megacephalum
0?????????1 000000000? ????0110000 0110001000 1000000000 1001010000 1010000100 1010001011
1111000?01 -100020000 0??000000- -----?00 0000000000 --0----- 0000110-00 0001000001
000??0--- ??????0?? ????01010000 10100100?? ??

Podocnemis expansa
0110010011 0000000000 0000111000 0110000000 1110000000 0000010000 1000000100 0000100021
1111100101 0000020000 0000000001 100101?100 0000001001 --0----- 1010000-01 0001001000
0000100--- 0011111?? ????0100010 00000000?1 ?0

Podocnemis unifilis
0110010011 00A000000? ????0110000 0111000001 1010000001 0000010000 0001000100 0010001011
1111000101 -100020000 000000000- -----?00 0000000000 --00100101 0100101-00 0001000100
0000000010 0000011010 1101010000 0011000001 00

Sternotherus odoratus
0?????????1 000000000? ????0110000 0111000000 1010000001 0000010000 0001000100 001000101?
1111000?01 -100020000 000000000- -----?00 0000000000 --10100101 0100101-00 0001000100
0000000010 0000010?? ?3?1010000 0011000001 00

Terrapene carolina
0?????????1 0010000110 0000001001 0111000000 111000000? ???1000000 0000000000 0011000121
?????0110 0000020000 000000000? ???-???00 0000100000 --1----- ??????-?? 0101000100
0000010--- ??????00?? ?????????? ?????????? ??

Testudo graeca
0110210011 0010100001 ??101????? 0110100000 1100000010 A0A00A0A0 000000000A ?AAA000A20
??01??0111 10000B0000 010000??- -----10A 0000000000 --A----- ??????-?? 0A01001101
00A111?--- ??1?00?? ????0?11?? ?0?0?0000 00

Trachemys scripta
0210210011 00A0100000 A??0100000 0110000000 1001100001 A01A010000 0A0000A001 10AA00002?
11110000A1 0A00020000 000000000- -----1A0 0000000000 1100000000 0000001000 010A00?100
00A0000--- 0001010010 1400000011 0000011000 00

Trachemys terrapen

```

0?????????1 0010000000 ???0110000 0110000000 1000100000 1011000000 110000???? ?11000021
???????0?01 0000020000 0101000001 1001?0??00 0000000000 --00000000 ??????-?? 0101000000
00?1?10--- ??????0A10 03????????? ?????????? ??

```

Appendix 3. Characters list 2 (multiple character approach; coding-2 and -3)

Characters are derived from Appendix 1. Numbers in square brackets indicate the corresponding characters in Appendix 1.

1-9 Characters 1-9 correspond to characters 1-9 of Appendix 1.

M. adductor mandibulae externus Pars medialis (No. 17)

- 10 Origin extends to the quadrate (0), capsule of the jaw joint (1), squamosal (2), postorbital (3), quadratojugal (4), opisthotic (5), prootic (6), supraoccipital (7), parietal (8). [10-19]
- 11 Origin extends the rostral face (0) or to the dorsal (1) of the quadrate. [10-11]
- 12 Insertion extends to the coronoid (0), surangular (1), dentary (2), coronar aponeurosis (3). [20-30]
- 13 Insertion extends to the lateral border (0), to the posterior border (1) of the coronoid. [20/1,20/2]
- 14 Insertion extends to the dorsal face (0), medial face (1), lateral face (1) of the surangular. [21-23]
- 15 Insertion extends to the lateral face (0), posterior face (1), ventral face (2), medial face (3), anterior face (4), dorsal face (5) of the coronar aponeurosis. [25-30]

M. adductor mandibulae externus Pars medialis inferior (No. 18)

16. Structure is absent (0), present (1). [31]

M. adductor mandibulae externus Pars profundus (No. 19)

17. Origin extends to the parietal (0), supraoccipital (1), postorbital (2), opisthotic (3), prootic (4), exoccipital (5), squamosal (6). [32-39]
18. Originates directly (0), via a tendinous framework (1) from the supraoccipital. [33-34]
19. Insertion extends to the coronoid (0), coronar aponeurosis (1), rictal plate (2), the dentary (3). [40-48]
20. Insertion extends to the medial face (0), dorsal face (1), ventral face (2), lateral face (3), posterior face (4), anterior face (5) of the coronar aponeurosis. [41-46]

M. adductor mandibulae externus Pars profundus atypica (No. 20)

21. Structure is absent (0), present (1). [49]

M. adductor mandibulae externus Pars superficialis (No. 21)

22. Origin extends to the quadratum (0), quadratojugal (1), squamosal (2), postorbital (3), jugal (4), parietal (5), opisthotic (6), prootic (7). [50-66]
23. Origin extends to the dorsal (0), medial (1), lateral (2), anterior (3) face of the quadrate, or to the posterior face of the quadrate and/or basioccipital (4). [50-53, 65]
24. Origin extends to the lateral (0), medial (1), anterior (2), dorsal (3), ventral (4), posterior (5) face of the squamosal. [55-60]
25. Insertion extends to the dentary (0), coronoid (1), surangular (2), coronar aponeurosis (3), rictal plate (4). [67-78]
26. Insertion extends to the lateral (0), posterior (1) face of the coronoid. [68-69]
27. Insertion extends to the dorsal (0), lateral (1), ventral (2) face of the surangular. [70-72]
28. Insertion extends to the lateral (0), anterior (1), posterior (2), dorsal (3), ventral (4) face of the coronar aponeurosis. [73-77]

Cartilago transiliens within coronar aponeurosis ...

29. has an articulation with: the pterygoid (0), with the quadratum / prootic (1). [79]

Coronar aponeurosis inserts ...

30. to the coronoid (process) (0), dentary (1), surangular (2), prearticular (3), rictal plate (4). [80-86]
31. to the dorsal (0), medial (1), lateral (2) face of the coronoid (process). [80-82]

M. zygomatocmandibularis (No. 22)

32. Structure is absent (0), present (1). [87]

M. adductor mandibulae internus Pars pseudotemporalis principalis (No. 23)

33. Position in relation to the ramus maxillaris nervi trigemini (V₂): lateral (0), medial (1), posterolaterally pierced by the nerve (2). [88]
34. Origin extends to the parietal (not processus descendens part) (0), processus descendens parietalis (1), epipterygoid (no epipterygoid in Pleurodira and Dermochelys coriacea sensu Joyce 2007) (2), prootic (3), postorbital (4), frontal (5), interorbital septum (6). [89-95]

35. Insertion extends to the subarticular aponeurosis (0), cartilago Meckeli (1), surangular (2), prearticular (3), Zwischensehne with this also a m. intramandibularis (No. 25) is present] (4), dental (5), articular (6), coronoid (7), angular (8). [96-109]
36. Insertion extends to the subarticular aponeurosis without m. adductor mandibulae internus Partes pterygoidei (No. 26-28) (at least connected to their tendon) (0), to the subarticular aponeurosis together with Partes pterygoidei (No. 26-28) (1). [96/1, 96/2]
37. Insertion extends to the dorsal face (0), medial (1), lateral (2), ventral (3) face of the cartilago Meckeli. [97-100]
38. Insertion extends to the dorsal (0), medial (1) face of the prearticular. [102-103]
39. Insertion extends to the dorsal (0), ventral (1) face of the coronoid. [107-108]

M. adductor mandibulae internus Pars pseudotemporalis superficialis (No. 24)

40. Origin extends to the parietal: no (0), yes (1). [110]
41. Insertion extends to the Zwischensehne [with this, also a m. intramandibularis (No. 25) is present] (0), subarticular aponeurosis (1), around the insertion of the coronar aponeurosis (coronar aponeurosis, cartilago transiliens, surangular, and / or coronoid process) (2). [111-113]

M. adductor mandibulae internus Pars intramandibularis (No. 25)

42. Insertion extends to the lateral face of the cartilago Meckeli (0), to the fossa (canalis) primordialialis (1), in a very long rostro-caudad extension (2). [114-117]
43. Insertion extends with a direct attachment to the bones of fossa (canalis) primordialialis (0), by a tendon to that fossa (1). [115-116]

M. adductor mandibulae internus Pars pterygoideus dorsalis (No. 26)

Origin extends (next to the dorsal face of the pterygoid) ...

44. Origin extends to the palatine (0), processus descendens parietalis (1), septum interorbitale (2), postorbital (3), jugal and maxilla (4), the frontal (5). [118-123]

Muscle fibres insert (next to insertion to the subarticular aponeurosis) ...

45. to the articular (0), prearticular (1), cartilago Meckeli (2), capsule of the jaw joint (3), surangular (4), angular

(5), coronoid (6), pterygoid tendon (7), rictal plate (8). [124-131, 133]

46. Inserts via pterygoid tendon: to the internal tendon (as apo.pteDOR) (0) / separated insertion to the prearticular/canalis primordialialis (as apo.pteVEN) (1). [132]

M. adductor mandibulae internus Pars pterygoideus posterior (No. 27)

47. Origin extends to the quadrate (0), pterygoid (1), palatine (2). [134-137]
48. Origin extends to the ventral face of or/and to the posterior face (0), lateral face and/or to the dorsal face of the pterygoid (1). [135-136]
49. Insertion extends to the articular (incl. retroarticular process) (0), surangular (1), prearticular (2). [138-140]

M. adductor mandibulae internus Pars pterygoideus ventralis (No. 28)

50. Origin extends only to the pterygoid (0), also to the palatine (1). [141]
51. Insertion extends to the surangular (0), coronoid (1), angular (2), prearticular (3), articular (4), subarticular aponeurosis (5), pterygoid tendon (6), rictal plate (7), capsule of the jaw joint (8). [142-150]

M. adductor mandibulae posterior (Pars principalis) (No. 29)

52. Has two muscle heads in the origin area: no (0), yes (1). [151]
53. Origin extends to the quadrate (0), pterygoid (1), prootic (2), lateral face of the parietal (3), quadratojugal (4). [152-156]
54. Origin extends to the quadrate: broadly at anterior surface (0), restricted to the medial aspect of the anterior surface (1), ventrally (2). [152]
55. Origin extends broadly to the prootic (0), restricted to the lateral aspect of the prootic (1). [154/1, 154/2]
56. Insertion extends to the prearticular (0), articular (1), angular (2), surangular (3), cartilago Meckeli (4), coronoid (5), coronar aponeurosis (6), inserts via posterior aponeurosis (also "Sehnenspiegel") (7). [157-167]
57. The posterior aponeurosis (also "Sehnenspiegel") inserts to surangular (0), prearticular (1), articular (2), coronoid (3). [164-167]

M. adductor mandibulae posterior Pars rostralis (No. 29)

58. Origin extends to the anterior face of the prootic (0), also to the parietal (1). [168]
59. Insertion extends to the subarticular aponeurosis (0), coronoid (1). [169-170]

Subarticular aponeurosis

60. Insertion extends to the cartilago Meckeli (0), coronoid (process) (1), prearticular (2), articular (3), surangular (4). [171-177]
61. Insertion extends to the dorsal (0), lateral (1) face of the cartilago Meckeli. [171-172]
62. Insertion extends to the dorsal (0), medial (1) face of the prearticular. [174-175]

M. intermandibularis (No. 31) ...

63. reaches the symphysis of the dentals: no (0), yes (1). [178]
64. a posterior trigonium is formed between m. intermandibularis and constrictor colli complex (No. 40-43): no (0), yes (1). [179]
65. contralateral muscles fuse in: a thin median raphe (0), a broad aponeurosis anteriorly (1), a broad aponeurosis posteriorly (2). [180]
66. fibre course of the posterior part of the muscle around 45° in posteromedial direction: no (0), yes (1). [181]
67. covers anterior region of constrictor colli complex (No. 40-43) superficially: other way around (0), yes (1), no, homogen transition (2), muscles do not contact (only connected in area of median raphe (3), connected by connective tissue (4). [182]

M. depressor mandibulae (No. 45) ...

68. has a bipinnated shape: no (0), yes (1). [183]
69. originates with two muscle heads: no (0), yes (1). [184]
70. Origin extends to the squamosal (0), quadrate (1), quadratojugal (2), opisthotic (3), jugal (4), cornu branchiale-II (5). [185-195]
71. Origin extends to the posterior (0), ventral (1), lateral (2), dorsal (3) face of the squamosal. [185-188]
72. Origin extends to the posterior (0), lateral (1), medial (2) face of the quadrate. [189-191]
73. Insertion extends direct to the articular (0), angular (1), or surangular (2), or via a tendon to the lower jaw (3). [196-202]
74. Insertion extends via a tendon to the articular body (0), retroarticular process (1), angular (2), surangular (3). [199-202]

Appendix 4. Characters matrix 2 (for coding-2 and -3)

Multiple character states are shown as a letters. For coding-2, all multiple states (numbers) were used. For coding-3, the letters were used. Multiple characters are coded as follows (bold number = character, letter = character with multiple states, number in brackets = multiple character states coded by the letter): **3** A=(0,1); **6** A=(0,1); **7** A=(1,2); **10** A=(0,1); B=(0,1,2,5),

C=(0,2), D=(0,2,4), E=(0,2,5,6), F=(0,2,7,8), G=(0,4), H=(0,4,6), K=(2,4), M=(2,4,5), N=(2,5), P=(2,6), Q=(2,8); **11** A=(0,1); **12** A=(0,1), B=(0,1,2), C=(0,1,3), D=(0,3), E=(1,3), F=(2,3); **13** A=(0,1); **14** A=(0,1); B=(0,1,2), C=(0,2); **15** A=(0,1), B=(0,1,2), C=(0,1,2,3,4,5), D=(0,1,3), E=(0,1,3,4), F=(0,1,3,4,5), G=(0,5), H=(1,3,5), K=(2,5), M=(4,5); **17** A=(0,1), B=(0,1,2), C=(0,1,2,3), D=(0,1,2,3,4), E=(0,1,2,3,5), F=(0,1,3), G=(0,1,4), H=(0,4,6), K=(0,6), M=(1,3,4), N=(1,5); **18** A=(0,1); **19** A=(0,1), B=(0,1,2), C=(0,1,2,3), D=(1,2); **20** A=(0,1), B=(0,1,2), C=(0,1,2,3,4), D=(0,1,2,3,4,5), E=(0,1,2,4), F=(0,1,3), G=(0,1,5), H=(0,2), K=(0,2,3), M=(0,3,4), N=(0,4), P=(1,2), Q=(1,2,4,5), R=(1,3), S=(1,4); **22** A=(0,1), B=(0,1,2), C=(0,1,2,3), D=(0,1,2,3,4), E=(0,1,2,3,4,5), F=(0,1,2,3,4,6,7), G=(0,1,2,3,5), H=(0,1,2,4), K=(0,1,2,4,6), M=(0,1,3,4), N=(0,1,3,5), P=(0,1,4), Q=(0,2), R=(0,2,3,4,5), S=(0,2,3,5,6,7), T=(0,2,5), U=(0,2,6), V=(0,3,4), W=(0,5,6,7), X=(1,2,3), Y=(1,2,4), Z=(2,3), a=(2,3,4), b=(2,5); **23** A=(0,1,3), B=(0,2), C=(0,4), D=(1,3), E=(1,4), F=(2,3); **24** A=(0,1,2,4), B=(1,2), C=(1,3), D=(1,3,4,5), E=(1,4,5), F=(1,5), G=(2,3); **25** A=(0,1,2), B=(0,1,3), C=(0,2), D=(0,2,3), E=(0,3), F=(0,3,4), G=(1,2), H=(1,2,3), K=(1,3), M=(2,3), N=(2,4), P=(3,4); **26** A=(0,1); **27** A=(0,1), B=(0,2); **28** A=(0,1), B=(0,1,2), C=(0,2), D=(0,4), E=(3,4); **30** A=(0,1,2), B=(0,1,2,3), C=(0,1,2,3,4), D=(0,1,3), E=(0,1,4), F=(0,2); **31** A=(0,1), B=(0,1,2), C=(1,2); **34** A=(0,1), B=(0,1,2), C=(0,1,2,3), D=(0,1,3), E=(0,1,4,5), F=(0,1,5), G=(0,1,6), H=(0,2,3), K=(1,3), M=(1,3,4), N=(1,3,4,5), P=(1,3,4,6); **35** A=(0,1), B=(0,1,3), C=(0,1,3,7), D=(0,2), E=(0,3), F=(0,3,4), G=(0,4), H=(0,5,6), K=(0,6), M=(0,7), N=(1,2,3), P=(1,2,4), Q=(2,7,8), R=(7,8); **37** A=(0,2); **38** A=(0,1); **42** A=(0,1), B=(1,2); **43** A=(0,1); **44** A=(0,1); B=(0,1,2), C=(0,1,3,4), D=(0,2), E=(0,3,4), F=(1,3), G=(1,5); **45** A=(0,1,2,3,7), B=(0,1,2,6,7), C=(0,1,7), D=(0,5), E=(0,5,6), F=(1,2,7), G=(1,4), H=(1,7), K=(1,8), M=(3,6); **46** A=(0,1); **47** A=(0,1,2); **49** A=(0,2); **50** A=(0,1); **51** A=(0,2), B=(0,3,5), C=(1,2,4), D=(1,6,7), E=(1,8), F=(2,4,6), G=(3,4), H=(3,4,6,7), K=(3,4,6,8), M=(3,5); **52** A=(0,1), **54** A=(0,1), **55** A=(0,1); **56** A=(0,1), B=(0,1,2), C=(0,1,3,6,7), D=(0,1,5,6), E=(0,1,6,7), F=(0,3,6), G=(0,7), H=(1,2), K=(1,3), M=(1,4,6), N=(1,6), P=(1,7), Q=(3,4), R=(6,7); **57** A=(0,1,2), B=(0,2); **60** A=(0,2), B=(0,2,3), C=(1,2,3), D=(1,2,3,4), E=(1,2,4), F=(2,3), G=(3,4); **61** A=(0,1); **62** A=(0,1); **63** A=(0,1); **64** A=(0,1); **66** A=(0,1); **67** A=(1,3), B=(2,3); **69** A=(0,1); **70** A=(0,1), B=(0,1,3), C=(0,1,4), D=(0,2), E=(0,3), F=(0,3,4); **71** A=(0,1,2), B=(0,2), C=(0,2,3), D=(0,3), E=(1,2), F=(2,3), **72** A=(0,1); **73** A=(0,1), B=(0,3), C=(0,1,2), D=(0,1,3), E=(0,2,3), F=(0,3); **74.** A=(0,1), B=(1,2), C=(1,3)

Lepidosauria									
11210A011Q	-E?AM0H-AF	0A-AA-A?x?F	001HP---01	2??DE????0	C002-B?--?	??1A00B005	????		
Crocodylia									
1000112110	01?C-00--?	0AG-2-0-??	?01-----0	2??DD?---0	A000-H?--?	??0120200?	?0-		
Neornithes									
10A011A012	--??-0K-0?	0QB0A-A?x?	?01-Q---10	2?-0??---1	?000-??--?	??A100100?	?0?		
Amyda cartilaginea									
011011101H	A1?C-1A0AD	02-C2-B-x?	?11A01----	-----0	M1010A---2	-A?????100	2?--		
Caretta caretta									
011021011G	0D0-F0A01P	0EAB3??01A	B01GN-2-1	0AABAAA00A	HA000GA--?	?????02010	0?3A		
Chelodina longicollis									
021011?00C	0E-BHOR0D4	1UDDP--B0C	B011H1--?-	---AD-111-	-001AD---E	?A1111?0AE	B-0-		
Chelonia mydas									
011?????11-	-C1210A01P	0E013??01A	B011M-20--	---BA1A-00	K1000RA--?	??????010	0-3A		
Chelonoidis denticulata									
0110210010	A1-2?0101A	0C012-B-1F	001C00----	---B71-0-1	-0???0----	-001011000	E?--		
Chelonoidis nigra									
021021001-	-3??A0A013	0C11H-101?	?0?KD1----	---BG----0	6001-P2--3	--?????011	-AD1		
Chelus fimbriatus									
001002001E	1C0CC0N01R	0b-2M-1002	?01401---1	1--B70---?	-0001N---3	--111A????	?-?		
Chelydra serpentina									
011101010P	-3?-D0A010	0EDGF-?A1E	A02P01----	---0-----0	5A010A----	--11000000	C-0-		
Chrysemys picta									
02100100?2	-F??A0A010	0HD-?????1?	?011G1---?	?B0-----?	-00107B--?	?-1000301-	-?--		
Clemmys guttata									
011??????2	-E-200A010	0MD-E--A1?	?02101--??	?-----?	-0010??--?	?-01000?0	2-??		
Cuora amboinensis									
01122101?N	-3--30A010	0VD-?????10	C02101--?-	-----?	-00104---?	?-0?????000	1---		
Dermochelys coriacea									
112100001M	AC11E0A0AQ	0RD1M-BAx-	-00300-0-?	?--0F1---1	6A0A03-??-	--1A00B0AD	0?C?		
Dogania subplana									
0110110?21	-BAC-0DABF	0K11KA-C1B	B1?E01----	---E6-----	-0???6---C	-A??????0-	?-B1		
Emydura macquarii									
021021000-	-A10-0B01P	0SEE3--E0B	B0?EE0-0--	---0C1---0	60011A?---	?0?????100	2-0-		
Emydura subglobosa									
021021001-	-1-2-0A0AP	0TCC3--D01	B01M00----	---071---?	-0011Q---3	--1101A000	C-00		
Emys orbicularis									
011012100N	-C02D0F01C	0MA-K0?A1B	B021B12A?-	---03---1	800101B---	??A1003000	B--?		
Eretmochelys imbricata									
011021011G	AE-210A014	0--1C-A-1F	001A00----	---D1---1	-1010A---2	?01100?000	0---		
Graptemys geographica									
0110210010	03??A0FA1N	0Q113--C1C	B0?101----	---03---1	8001172--A	1A??????00	2-31		
Graptemys pseudogeographica									
0???????C	0-???0A014	0PD-3??A1?	?011G1---?	?20-----?	-0010??--?	???????????	?-?		
Hydromedusa tectifera									
0210111000	03-2A0C01?	0Q1F---?0B	B01EA10---	---F4-----	-0010F---A	0A?????00B	A131		
Kinosternon leucostomum									
0???????C	03--K0AA1E	0DDCD-AA10	B02101--??	----8----?	-0010P2--?	??0???????	?-?		
Kinosternon scorpioides									
0110110??C	03--K0AA1E	0DDCD-AA1?	?02101--?-	----8----?	-0010P2--?	??0?????000	2---		
Lepidocheilus kempii									
0110??0?00	0----0A010	0F33-??-10	B01141----	-AA0H----1	81011A---?	??1?002??0	A---		
Lissemys punctata									
01101100?A	00A2-0DABF	0UD13A-C1B	B11E01----	---E6----?	-000-E301C	-A110?3100	F-BB		
Macrochelys temminckii									
01102101?0	03??10M01K	0B11P?101B	B0?NM1--11	0A1F-----	-001-A---C	-A0000201C	D1A-		
Malaclemys terrapin litoralis									
011??????C	03-2B0A01M	0ND-E--A1?	?02101--??	?-----?	-001072--?	??0?????000	B---		
Mauremys caspica									
0110??01?C	03-200A01N	0MD-3--A10	B02101--??	?-----?	-0011??--B	A00?????000	B?--		
Mauremys rivulata									
0???????1?0	03??A0F01B	0Y1-4-??1B	B0?1C1A10?	?--AG----0	F0011A---3	--???????00	2--?		
Mesoclemmys nasuta									
0010?10100	03--20A01S	0WD?M?0304	?021E1-0--	-----?	-0010K---?	??1?000000	2--?		
Pelodiscus sinensis									
011011001B	0DA2G0DACC	0KD1BA?B1B	B12F01----	---C6-1101	100A0A---C	-A?????1A0	E-EC		
Pelomedusa subrufa									
021002000A	03-2-0A0AR	0D11K0-D0D	B011E0-0--	---GB1---1	D001AM---B	0001?0300E	E-B1		
Pelusios niger									
0210210000	03---0A014	1KD14---04	?011K1----	-----?	-101071--?	??11013???	???		
Pelusios sinuatus									
0210210000	03??A0G010	0G11G01D0A	B01K01----	-----0	G00003---?	???????01B	120?		

<i>Pelusios subniger</i>	0?????????0	03??A0G010	0G11G01D0A	B0?K01----	-----0	G00003---?	?????????1B	120?
<i>Platysternon megalcephalum</i>	0110010010	03--B0A01B	0Z-1K0-21B	B01101---1	0A10M---1	E0000G1--D	-A?????00A	0031
<i>Podocnemis expansa</i>	011001001C	03??A0AAA	0Q0110-D0A	B01K01----	-----10A0	B00001-00G	--0101101F	1-31
<i>Podocnemis unifilis</i>	0?????????0	03??A0AA1H	0Q0110-D0A	B0?K01----	----8-10A0	B00001-003	--?????3?1F	1-31
<i>Sternotherus odoratus</i>	0?????????F	03--K0AA1B	01--4--A1?	?01001---?	?---K---?	-0011P2--?	??0???????	???
<i>Terrapene carolina</i>	011021001D	0D0200B01A	1QDCN-BA1F	001BE1-0?-	---D8----?	-00AACA--?	?00?????000	F?--
<i>Testudo graeca</i>	021021001D	03-000A01M	0HA1D-AA1A	C00D01----	---A71---0	50011N---F	-001014001	-AA-
<i>Trachemys scripta</i>	0?????????C	03??A0A01N	0MD--??A1?	?0?1F1-0-1	00-----?	-00117B--?	??A1003???	???
<i>Trachemys terrapen</i>	011021?00C	03???0B01G	0Q11---A10	B02A01---1	00-----?	-0???-????	??010130?A	10??

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