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Contributions of peroxisome proliferator-activated receptor β/δ to skin health and disease

Abstract: Among the three peroxisome proliferator-activated receptor (PPAR) transcription factors, PPAR β/δ is the isotype with the broadest expression pattern. In fact, the expression of PPAR β/δ is ubiquitous, albeit at levels that are tightly regulated. Herein, we reviewed its multiple functions in skin health and disease. PPAR β/δ has pro-differentiating effects in keratinocytes, regulates sebocyte differentiation, and promotes hair follicle growth in healthy skin. Furthermore, we reviewed novel insights into the roles of PPAR β/δ in skin wound healing, especially in inhibiting apoptosis and in modulating keratinocyte proliferation and migration. Therefore, PPAR β/δ represents a research target for the understanding and treatment of inflammatory skin diseases, such as psoriasis and acne vulgaris. In addition, PPAR β/δ is a tumor growth modifier. Epidemiological studies have established that tumor progression may be exacerbated by chronic low-grade inflammation, a condition promoting the production of the lipids that act as modulators of PPAR β/δ activity. The action of PPAR β/δ in skin cancer is ambivalent, which might be explained by this receptor's putative highly context-specific behavior, which depends on a combination of factors ranging from receptor expression levels to co-regulator distribution, diversity and activity of the ligands produced, and other tissue-specific conditions. Given its diverse and crucial roles in many tissues and organs, PPAR β/δ will remain a major focus of future research.

Keywords: keratinocytes; non-melanoma skin cancer; nuclear receptors; psoriasis; wound healing.

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Introduction

The skin is the largest organ of the body with respect to weight and surface area. The outer layers of cells in the

skin compose the epidermis, which together with the dermis forms the cutis. In the latest stages of fetal development, the epidermis develops into a fully stratified and differentiated epithelium that undergoes continuous renewal after birth. In fact, the basal layer of this stratified tissue contains undifferentiated proliferating keratinocytes that undergo a differentiation program as they migrate from the basal to the uppermost layer of the epidermis. The basal layer is the source of skin cells throughout life, which are generated through asymmetrical division (1, 2). This cell layer is maintained through several autocrine and paracrine acting factors, which are important for epidermal homeostasis (3–5). The vectorial differentiation program of keratinocytes comprises the processing of lipids [sphingolipids, free fatty acids (FAs), and cholesterol] and the sequential expression of structural proteins (keratins, involucrin, loricrin, and filaggrin), which are each essential to the protective function of the epidermis. In fact, the outermost layer of the epidermis, the stratum corneum, consists of a layer of cross-linked proteins and lipids and dead keratinocytes (corneocytes), which form an efficient barrier against water loss, microorganism invasion, mechanical damage, and chemical poisoning. The stratum corneum is also where desquamation, the natural process by which dead outer skin cells are sloughed away and replaced, occurs (6). Epidermal homeostasis and the integrity of the barrier function depend on a well-tuned coordination of keratinocyte proliferation, differentiation, positioning in the tissue, and apoptosis.

As in other organs, lipid signaling contributes to skin health and disease. The nuclear receptors called peroxisome proliferator-activated receptors (PPARs) are key mediators of lipid signaling. They sense diverse lipophilic molecules that act as ligands and thereby link fluctuations in the levels and distribution of FAs, eicosanoids, and some phospholipids to differential gene expression (7, 8). There are three related PPAR isotypes that compose a small subfamily of nuclear receptors: PPAR α (NR1C1), PPAR β/δ (NR1C2), and PPAR γ (NR1C3) (9). The transcriptional activity of all three of them is mediated by PPAR:retinoid X receptor heterodimers that bind to specific DNA sequence elements termed peroxisome proliferator response elements. By doing so, they

regulate the expression of genes implicated in several important processes, such as lipid and carbohydrate metabolism, tissue repair, vascular biology, and sexual dimorphism, as well as in more general basic cellular processes, such as proliferation, differentiation, and migration (10, 11).

The expression patterns of PPAR α , PPAR β/δ , and PPAR γ are well described (10, 12, 13). Although presenting some specificity, they are also overlapping, with distinct expression levels. PPAR α , which has been well described in FA catabolism, is highly expressed in brown adipose tissue, heart, liver, kidney, and intestines, whereas PPAR β/δ is relatively abundant in the brain, skeletal muscle, gut, placenta, and skin. The third isotype, PPAR γ , is expressed as two isoforms, $\gamma 1$ and $\gamma 2$. PPAR $\gamma 1$ is 28 and 30 amino acids shorter at its N-terminus than PPAR $\gamma 2$ in rodents and humans, respectively. PPAR $\gamma 1$ also has a broader expression pattern, which includes immune cells, brain, gut, and endothelial cells, whereas PPAR $\gamma 2$ is found mainly in adipose tissues. This list is not exhaustive; a more detailed description of PPAR expression can be found in previous articles (10, 12–14).

In recent years, PPARs (and especially PPAR β/δ) have emerged as having multifaceted key roles in skin health and disease (Figure 1). Characterized natural PPAR β/δ ligands are unsaturated FAs, saturated FAs (weak), prostacyclin, 4-hydroxy-2-nonenal, 4-hydroxydodeca-(2E,6Z)-dial, and very low-density lipoprotein components (15). The present review summarizes our current knowledge of PPAR β/δ functions in rodent and human skin, including embryonic development, the maintenance of adult skin and skin appendages, and normal skin repair. We also discuss the implication of PPAR β/δ in skin pathologies,

such as hyperproliferative and inflammatory diseases, and cancer.

Skin development: cell differentiation, epidermis, and appendages

The three PPAR isotypes are expressed in rodent and human skin. Cell culture and *in vivo* approaches have demonstrated that PPAR β/δ has pro-differentiating effects in keratinocytes in normal and inflammatory conditions, regulates hair follicle growth, and promotes sebocyte differentiation.

In keratinocytes from adult human epidermis, PPAR β/δ is constitutively expressed and is present throughout all epidermal layers (16–18). In contrast, in the interfollicular epidermis of rodents, PPAR β/δ is expressed at relatively high levels during development and much less in the adult epidermis (19). In mouse keratinocytes, PPAR β/δ expression can be down-regulated by C/EBP α and C/EBP β through a mechanism that requires both the binding of C/EBP to the promoter of the PPAR β/δ gene and histone deacetylation. We propose that such interplay between PPAR β/δ and C/EBP transcription factors is crucial to the molecular control of the balance between differentiation and proliferation (20). Furthermore, keratinocyte fatty acid binding protein (K-FABP) is essential to the ability of PPAR β/δ to properly induce keratinocyte differentiation. Hence, the tissue-specific expression of particular FABPs may support the tissue-specific functions of PPARs, which raises the intriguing possibility that the same K-FABP–PPAR β/δ pair plays a similar role in mediating the differentiation of other cell types in which both factors are expressed concomitantly

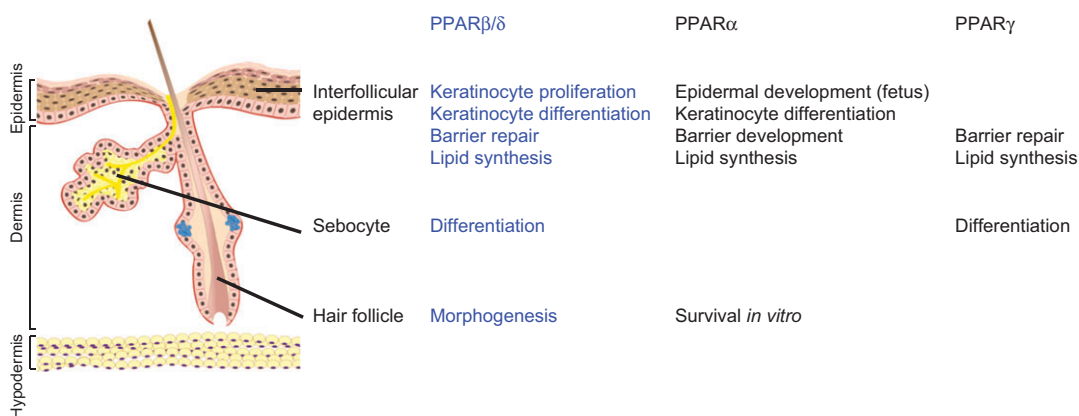


Figure 1 Functions of PPARs in the epidermis and epidermal appendages.

This schematic outlines the skin structures and functions of PPARs in the interfollicular epidermis, sebocytes, and hair follicles. Observations obtained after activation of PPARs by their specific ligands in three skin structures are summarized (see text for references).

(21). The pro-differentiating action of ligand-activated PPAR β/δ is keratinocyte-autonomous and mediated by an indirect mechanism that involves the transcriptional regulation of adipocytokine angiopoietin-like 4 (ANGPTL4) and the subsequent activation of protein kinase C and activator protein 1 (AP-1) transcription factors (22).

Hair follicle morphogenesis depends on a strictly controlled balance between keratinocyte proliferation, differentiation, and apoptosis. PPAR β/δ is expressed in the hair follicles of both fetal and adult skin. Furthermore, follicle morphogenesis involves complex bidirectional interactions between the keratinocytes and the underlying mesenchyme. These interactions involve hepatocyte growth factor (HGF), which is produced by dermal papilla cells during early hair follicle development. HGF stimulates cyclooxygenase-2 (COX-2) expression in keratinocytes, which results in the activation of PPAR β/δ and the subsequent stimulation of the protein kinase B α (PKB α)/Akt1 pathway at a specific stage of hair follicle development. However, in contrast to inflammatory cytokines that regulate both PPAR β/δ expression and activation, HGF is implicated only in PPAR β/δ activation. Interestingly, HGF is a versatile modulator of cell proliferation, migration, differentiation, and apoptosis. It has been implicated in mesenchyme-epithelium interactions in several tissues and has been attributed functions in the morphogenesis of other ectodermal structures, such as teeth and feathers, which may therefore also require PPAR β/δ activity (23).

In brief, activation of PPAR β/δ by COX-2-derived ligands protects hair peg keratinocytes against apoptosis, promoting normal hair follicle development (24).

Each PPAR isotype is expressed in the basal layer of the sebaceous gland, and ligand activation of each isotype increases lipogenesis (25). The gland comprises sebocytes that are epithelial cells, which accumulate neutral fat droplets during terminal differentiation. Sebocyte lysis releases lipids/sebum into the hair follicle canal, through which they reach the surface of the skin where they prevent drying of the skin and hair (26). Interestingly, PPAR β/δ seems to be important in the late stages of sebocyte differentiation (27), in contrast to its implication in the early phase of adipocyte differentiation in adipose tissues (28). The meaning of this stage-specific effect on the differentiation of two lipogenic cell types remains to be unveiled.

Skin lipid metabolism

As mentioned above, the permeability barrier resides in the stratum corneum and comprises lamellar membranes

enriched with extracellular neutral lipids surrounding the corneocytes (29, 30). These extracellular lipids differ from the lipids that constitute most biological membranes. Their total lipid mass comprises approximately 25% cholesterol, 50% ceramides, and 15% free FAs (31). Mice devoid of PPAR β/δ displayed delayed recovery of the permeability barrier, suggesting that this PPAR isotype has a physiologic function in permeability barrier homeostasis. This delay in barrier recovery was most likely due to the decreased synthesis and secretion of lamellar bodies, as well as reduced numbers of extracellular lamellar membranes in the stratum corneum (32).

Treatment of both cultured human keratinocytes and mouse epidermis with a selective PPAR β/δ ligand (GW1514) increased the accumulation of lipids that were predominantly triglycerides, but did not stimulate the conversion of keratinocytes to sebocytes or adipocytes. It is not yet clear why lipid accumulation is increased in these cultured keratinocytes or whether this *in vitro* observation has functional significance *in vivo* (33).

Skin wound healing

The formation of a new epithelium after a skin injury is part of a healing process that is often lifesaving. The initial stage of this process is an immediate inflammatory response to the injury. Next, keratinocytes migrate and proliferate to cover the wound bed in the *sensu stricto* reepithelialization event. The proliferation of fibroblasts from the dermis also contributes to the healing process, as does the production of novel blood vessels that irrigate the repaired skin. In rodents, PPAR β/δ expression is extremely low in the interfollicular adult epidermis, but is strongly reactivated and remains relatively high in keratinocytes at the edges of wounds as long as the entire repair process has not been completed (19). The injury-associated stimulation of PPAR β/δ is mediated by the activation of the stress-associated protein kinase pathway by inflammatory cytokines that are released after the injury, such as tumor necrosis factor- α (TNF- α) (Figure 2) (34). Concomitantly, the production of an endogenous ligand (or multiple ligands) is induced, which is a prerequisite for activation of the receptor in the keratinocytes (34). The expression of PPAR β/δ is maximal during the inflammatory phase and is then progressively reduced during reepithelialization to finally reach levels observed in the unwounded skin, once cicatrization has been achieved. The down-regulation of PPAR β/δ depends on the inhibition of AP-1 binding to the PPAR β/δ promoter, which

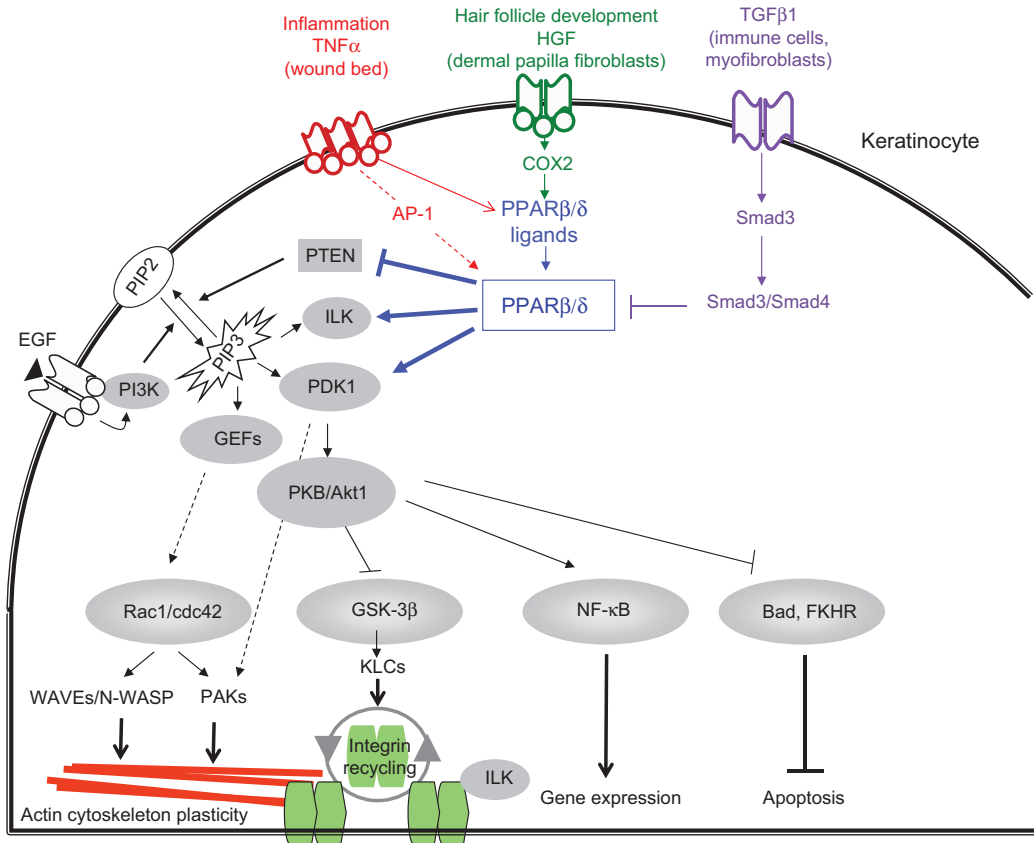


Figure 2 Roles of PPAR β/δ in skin wound healing.

The expression of the *Ppar* β/δ gene is stimulated by the AP-1 transcription factor complex, which is activated by the stress-associated protein kinase pathway triggered by pro-inflammatory cytokines such as TNF- α . TNF- α also induces the production of PPAR β/δ ligands in the injured epithelium. Ligands can also be produced during hair follicle development by COX-2-mediated HGF signaling. Activated PPAR β/δ down-regulates *Pten* expression and stimulates the genes that encode integrin-linked kinase (ILK) and 3-phosphoinositide-dependent kinase-1 (PDK1). The latter results in activation through the phosphorylation of PKB α /Akt1, which increases the inhibitory phosphorylation of BAD (BCL2-associated agonist of cell death), FKHR (forkhead transcription factor Foxo1), and GSK-3 β (glycogen synthase kinase 3 β), causing reduced apoptosis and increased gene expression and integrin recycling. As epithelialization proceeds, TNF- α -induced *Ppar* β/δ expression is progressively repressed by Smad3/Smad4 complex-mediated TGF β 1 signaling, which inhibits AP-1 binding to the *Ppar* β/δ promoter. Increased levels of PIP3 (phosphatidylinositol 3,4,5-trisphosphate) and PDK1 affect the cytoskeleton, which is implicated in directional sensing and cell migration.

is controlled by transforming growth factor β 1 (TGF β 1) (35). The completion of skin repair is delayed by 2–3 days in PPAR β/δ -deficient mice (19), which underscores the importance of this transcriptional regulator in wound healing. In fact, PPAR β/δ activates the phosphoinositide 3-kinase (PI3K)/PKB α /Akt1 (PI3K/PKB α /Akt1) pathway, which promotes keratinocyte survival at the wound edges (24). Furthermore, it favors keratinocyte adhesion and migration, which promotes reepithelialization (19, 36) (Figure 2).

To understand the functional interaction between PPAR β/δ and TGF β 1, topical application of recombinant TGF β 1 (TGF β 1 pathway gain of function) and Smad3 (mothers against decapentaplegic homolog 3) deficiency

(TGF β 1 pathway loss of function) were used (37). Manipulating TGF β 1 activity in the epidermis of the wounds induced prolonged PPAR β/δ expression and activity, which resulted in accelerated wound closure. This result suggested that PPAR β/δ expression and activity correlate with healing efficiency. Such knowledge about how TGF β 1 regulates PPAR β/δ during wound healing may help improve treatments for chronic wound disorders.

Moreover, gene expression profiling in wild type and PPAR β/δ -deficient primary keratinocyte cultures revealed similar amounts of genes stimulated or repressed by PPAR β/δ . Comparative analysis of expressed genes confirmed PPAR β/δ functions in cell proliferation, differentiation, migration, and adhesion (11). Of particular interest,

the most deregulated genes were genes previously implicated in cell proliferation, tumor growth, and angiogenesis (38–40). For instance, three of these genes, *Akt1*, *Cxcl12*, and *Src*, regulate the PI3K and mitogen-activated protein kinase (MAPK) pathways, both of which have been implicated in skin wound healing. In brief, PPAR β/δ deficiency in keratinocytes affected gene expression, which may explain the phenotypic changes observed during skin wound healing in PPAR β/δ -deficient mice (11).

The regeneration of the epithelium to close a wound depends on a contribution from the underlying dermal tissue, particularly fibroblasts and fibroblast-like cells, called fibrocytes, derived from the bone marrow (Figure 3) (41, 42). Regulation of the crosstalk between keratinocytes and fibroblasts is central to preventing either insufficient or excess wound repair. Notably, keratinocyte proliferation after an injury must be fine-tuned for reepithelialization (43). Interestingly, PPAR β/δ deficiency caused

epidermal hyperproliferation in early wound repair, upon hair plucking, or in response to a topical challenge with the phorbol ester 12-*O*-tetradecanoylphorbol-13-acetate (TPA) (19, 44). During wound healing, this keratinocyte proliferation is under the control of fibroblasts that produce mitogenic factors whose genes are under the control of AP-1, which is itself under the upstream regulation of the interleukin 1 (IL-1) released by keratinocytes at the site of injury (Figure 3). In parallel, activated PPAR β/δ in fibroblasts increases the production of the secretory IL-1 receptor antagonist (sIL-1Ra), resulting in the autocrine down-regulation of IL-1 signaling. As a consequence, the production of secreted mitogenic factors by the fibroblasts is reduced and the proliferation of keratinocytes is down-regulated (43). Together, these findings revealed a novel paracrine effect of PPAR β/δ on epithelial cell growth, which might also operate in other organs in addition to skin. Thus, PPAR β/δ controls keratinocyte differentiation,

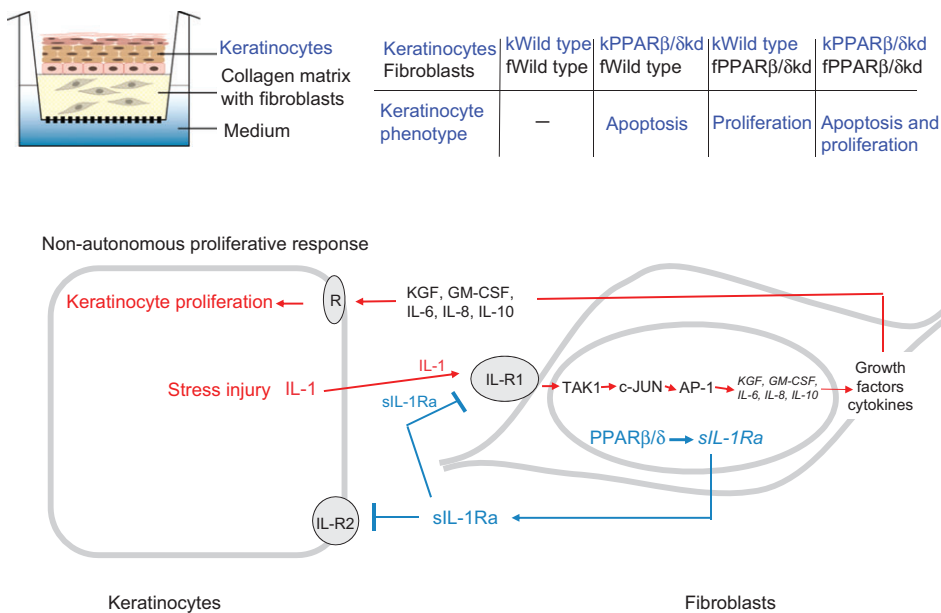


Figure 3 Non-cell-autonomous control of keratinocytes by PPAR β/δ .

Organotypic cultures [OTCs; (79); see scheme] with primary human keratinocytes and fibroblasts were grown with control or PPAR β/δ knockdown keratinocytes (kWild type vs. kPPAR β/δ kd) and fibroblasts (fWild type vs. fPPAR β/δ kd). kPPAR β/δ kd presented increased apoptosis, whereas fPPAR β/δ kd potentiated keratinocyte proliferation. OTC with both kPPAR β/δ kd and fPPAR β/δ kd exhibited increased keratinocyte proliferation and apoptosis (43). Keratinocyte proliferation is regulated by a nonautonomous paracrine mechanism. IL-1 produced by keratinocytes activates c-Jun through IL-1 receptor type 1 (IL-1R1) and transforming growth factor β -activated kinase 1 (TAK1) in dermal fibroblasts. c-Jun is an obligate partner of the AP-1 transcription complex, which stimulates the expression of mitogenic factors [keratinocyte growth factor (KGF), granulocyte-macrophage colony-stimulating factor (GM-CSF), IL-6, IL-8, IL-10] secreted by the fibroblasts. In these cells, PPAR β/δ attenuates IL-1 signaling through the stimulation of sIL-1Ra production; the gene that encodes sIL-1Ra is a direct PPAR β/δ target. sIL-1Ra has little affinity for IL-1R2, which is highly expressed in keratinocytes. However, it binds with high affinity to IL-1R1 expressed by the fibroblasts. Therefore, sIL-1Ra acts as an autocrine antagonist of IL-1 signaling in fibroblasts. The binding of sIL-1Ra to IL-1R1 down-regulates IL-1-mediated signaling events and, consequently, the production of several AP-1-mediated mitogenic factors. This mechanism leads to reduced keratinocyte proliferation. Therefore, the absence of PPAR β/δ in fibroblasts results in increased keratinocyte proliferation (43).

survival, and migration, as well as keratinocyte proliferation by cell-autonomous and nonautonomous pathways.

Skin diseases

Psoriasis

Psoriasis is an inflammatory skin disorder in which epidermal hyperproliferation and abnormal keratinocyte differentiation are major characteristics. It affects 1–2% of the population in the USA and is often debilitating. PPAR β/δ is increased in psoriatic areas, most likely through inflammatory signals as previously described after a skin injury (45). A transgenic murine model allowing PPAR β/δ activation in the epidermis sustained an inflammatory skin disease similar to psoriasis, with keratinocyte hyperproliferation, dendritic cell accumulation, and endothelial activation (45). This response required the activation of the Th17 subset of T cells, known to be central to psoriasis. Key transcriptional programs of psoriasis, such as cholesterol synthesis and IL-1-related signaling, were replicated in this model, suggesting that PPAR β/δ might regulate these changes in psoriasis. Furthermore, phosphorylation of signal transducer and activator of transcription 3 (STAT3) was enhanced by PPAR β/δ , whereas inhibition of STAT3 phosphorylation blocked disease development, implicating PPAR β/δ in this process (45). Interestingly, PPAR β/δ was also expressed in activated human T cells isolated from psoriatic skin lesions, where it is induced by stimulation with type 1 interferon (IFN). This study suggested that the induction of PPAR β/δ by type 1 IFN contributes to the prolongation of activated T cells in psoriasis-affected skin (46). Using the same murine model as above, it was shown that three selective PPAR β/δ antagonists formulated for topical application to the skin demonstrated efficacy in reducing psoriasis-like changes triggered by PPAR β/δ activation. One of these compounds, an irreversible PPAR β/δ antagonist (GSK3787), retained efficacy when applied topically only three times per week. Collectively, these observations suggested the clinical usefulness of topical inhibition of PPAR β/δ to treat psoriasis, which deserves further exploration (47). In this experimental model of psoriasis triggered by PPAR β/δ overexpression, one can argue that inhibiting PPAR β/δ logically suppresses the origin of the disease. Thus far, PPAR β/δ antagonists have not been evaluated on psoriasis models not induced by experimental PPAR β/δ overexpression, and antipsoriasis drugs have not been applied to the PPAR β/δ -induced disease model. In contrast to these encouraging results obtained with

PPAR β/δ antagonists, a pilot study with psoriatic patients showed that the PPAR β/δ agonist tetradecylthioacetic acid did not exhibit beneficial effects when topically applied on plaque psoriasis (48). This finding comes as no surprise, given the rather psoriasis-promoting characteristics of PPAR β/δ described above.

Atopic dermatitis

Atopic dermatitis (AD) is an inflammatory, chronically relapsing, noncontagious, and pruritic dermatosis that is linked to alterations of the stratum corneum (49, 50). Little is known about the putative link between PPAR β/δ and AD. In the oxazolone-induced AD-like model in hairless mice, topical application of a PPAR β/δ activator (GW0742) reversed clinical dermatosis, improved barrier function, and increased stratum corneum hydration. These results suggest that topical applications of certain activators/ligands of PPAR β/δ may be worth testing as AD treatment in human subjects (50). Thus, in contrast to psoriasis, for which PPAR β/δ antagonists were beneficial in experimental models, AD treatment might benefit from PPAR β/δ agonists. However, it is important to note that there are no human data so far and the data discussed above are from an experimental mouse model that shows only a few similarities with human AD.

Acne

Acne is a common skin condition characterized by spots that range from mild to inflamed pus-filled pustules and cysts. When long-lasting, it can lead to scarring. Sebum secretion can be considered the major factor responsible for acne, even if seborrhoea *per se* is not a sufficient condition for its development. Current studies concentrate on the factors that regulate sebum composition and secretion. Differences in the type, amount, and arrangement of FAs in the sebum lipids have been reported in acne patients. Unsurprisingly, a number of lipid mediators that have been shown to be PPAR ligands impact sebocyte lipid synthesis and metabolism (51). In fact, increased sebum levels combined with follicular hyperkeratinization are hallmarks of acne vulgaris, and PPAR β/δ was found overexpressed in inflammatory and non-inflammatory acne vulgaris (52). Given that the suppression of sebum secretion correlates with reduced acne, the manipulation of PPAR activity in this condition is worthy of additional exploration.

Among PPAR activators, the PPAR β/δ ligand L-165.041, compared with PPAR α and PPAR γ ligands, was the most

efficient at inhibiting basal and staurosporine-induced apoptosis in the human sebocyte cell line SZ95 *in vitro*. L-165.041 activated PKB/Akt and p44/42 MAPK, two kinases involved in antiapoptosis activity and proliferation, respectively. The inhibition of these kinases reversed the effect of L-165.041, implicating these pathways in the observed antiapoptotic effect. It was concluded that PPAR β/δ activation might have beneficial effects on acne vulgaris by inhibiting the release of lipids associated with sebocyte apoptosis (53).

Skin cancer

Non-melanoma skin cancer (NMSC)

PPAR β/δ regulates cellular mechanisms that have been recognized as pro- or anticancer, such as cell survival, migration, and differentiation. Therefore, whether PPAR β/δ supports or suppresses tumor formation is still being debated (54, 55). Similarly, the implication of PPAR β/δ in NMSC is much discussed (54, 56).

The role of PPAR β/δ in cancer development has remained elusive, most likely because its promoting or repressing effects on the above-mentioned processes are fine-tuned in a context-dependent manner, which thus far has allowed PPAR β/δ to escape a simplistic view of its functions in tumor biology. Two general models have emerged from these studies that attempt to characterize the ambivalent action of PPAR β/δ in tumorigenesis, possibly reflected in the variation in genetic defects among tumors. They have been presented in some detail recently (56).

In the first model, PPAR β/δ expression is mediated by inflammatory signals through the stress-associated protein kinase pathway, in concert with the notion that low-grade inflammation is often associated with tumor initiation and progression. Alternatively, increased PPAR β/δ expression is supported by APC- β -catenin-dependent signaling, which is enhanced in several tumors, especially colorectal carcinomas. An increase in PPAR β/δ level is insufficient for the activation of target genes, which depends on ligand activation of the receptor. Although endogenous ligands that activate PPAR β/δ during tumorigenesis have not yet been identified, they are produced and are likely diverse. Once activated, one effect of PPAR β/δ is the down-regulation of phosphatase and tensin homolog (PTEN), which results in increased AKT phosphorylation, which in turn inhibits apoptosis and up-regulates vascular endothelial growth factor expression, which might promote tumor growth. Furthermore, ANGPTL4, encoded in a previously identified

PPAR target gene (57, 58), has been implicated in cancer progression, although evidence that PPARs up-regulate ANGPTL4 directly in tumor cells is still lacking (59). Moreover, at least in colorectal cancer, PPAR β/δ might mediate the prostaglandin E2-induced expression of ANGPTL4 under hypoxic conditions, which are often observed in the tumor environment (60). Importantly, targeted deletion of PPAR β/δ in colonic epithelial cells inhibited azomethane-induced colonic tumorigenesis (61), and PPAR β/δ is a k-Ras target gene that is up-regulated by the Raf/MEK/ERK pathway in cultured intestinal epithelial cells (62). However, in another study, germline deletion of PPAR β/δ increased azomethane colonic carcinogenesis (63). Taken together, these two studies might indicate paracrine effects of PPAR β/δ . Other factors that might have contributed to such contrasting results will be discussed below. Interestingly, analysis of human skin biopsies of premalignant and malignant skin carcinoma documented increased PPAR β/δ expression, which correlated with both increased COX-2 expression, which has been implicated in skin carcinoma development, and increased microvessel density (64–66). Collectively, these studies have implicated PPAR β/δ in skin tumor formation and progression.

In the second model, activated PPAR β/δ promotes terminal epithelial cell differentiation, and therefore cell cycle withdrawal. Furthermore, its anti-inflammatory action might contribute to decreased tumorigenesis (56). Several studies, all from the same group, show that loss of PPAR β/δ function increases chemically induced skin carcinogenesis (67–70). Along the same line, combining ligand activation of PPAR β/δ with inhibition of COX-2 activity increased the efficacy of preventing chemically induced skin tumorigenesis over that of either intervention alone (70). This effect of PPAR β/δ was thought to be mediated by the inhibition of keratinocyte proliferation, mainly through the activity of protein kinase C α . However, in transgenic mice that overexpress protein kinase C α , 7,12-dimethylbenz[*a*]anthracene (DMBA)/TPA treatment had no effect on skin tumor susceptibility. Moreover, the induction of skin cancer by DMBA/TPA involves different molecular mechanisms than skin cancer associated with chronic UV exposure, which is largely the more frequent form in humans (71, 72). Notably, tumor initiation with a unique topical dose of the genotoxic carcinogen DMBA primarily induces *Ha-Ras*-activating mutations, whose frequency only reaches 10–20% in human NMSCs (73–75). In this context, many upstream signaling pathways involved in skin cancer establishment that might be regulated by PPAR β/δ could be blinded by mutations such as those affecting *Ha-Ras*, thereby masking the impact of

this nuclear receptor on skin cancer in the DMBA/TPA model.

In summary, the opposite pro-differentiation and pro-carcinogenic roles of PPAR β/δ described above may be due to different environmental stimuli, possibly impacting ligand production, and/or different genetic contexts, which could influence PPAR β/δ activities and functions. Furthermore, different mouse genetic models targeting the PPAR β/δ DNA binding domain with a complete loss of PPAR β/δ function or the transactivating domain that might result in a dominant-negative PPAR β/δ were used. All of the above-mentioned variables are likely to affect the expression pattern of numerous PPAR β/δ target genes. Obviously, more work is needed to uncover the determinants of the ambivalent role of PPAR β/δ in tumor development.

Melanoma

Similar to NMSC, the incidence of malignant melanoma is also steadily increasing (76). PPAR β/δ expression has been observed in human melanoma biopsies. Low-dose pharmacological ligand activation of PPAR β/δ in cultured murine and human melanoma cells inhibited cell growth *in vitro*. At the molecular level, this inhibition is due to the direct binding of PPAR β/δ to the Wilms' tumor suppressor (WT1) promoter, which represses its activity, and consequently the growth-stimulating effects of WT1 on these melanoma cells (77).

Expert opinion and outlook

Exploration of the functions of PPAR β/δ remains of highest interest because of its multifaceted roles in major cellular and physiological functions. At the molecular level, PPAR β/δ operates as a ligand-dependent transcription factor that stimulates or represses gene expression. The diversity of functions of this receptor, which range from the control of basic cellular processes to the regulation of complex metabolic functions, represents a major challenge for present and future studies (15). Herein, we have discussed how PPAR β/δ occupies the crossroads of multiple functions in skin biology. From this discussion, PPAR β/δ emerged as a key factor in skin homeostasis and repair. In addition, its implication in several skin diseases identifies it as a target to take into account in the development of novel therapeutic strategies. The similarity of processes involved in tissue repair

and tumor development (78) (Table 1), in all of which PPAR β/δ is implicated, underscores how many facets of PPAR β/δ biology remain hidden. Is the difference, if any, between cell proliferation in tissue repair and cell proliferation in tumorigenesis situated in parallel with the PPAR β/δ repressor and activator activity of these processes? The ambivalent roles of this receptor may explain its apparently discordant activities reported in the literature. The debate is not closed, and most likely reflects the difficulty of accurately describing the highly context-specific behaviors of PPAR β/δ , which depend on a combination of factors, including the level of receptor expression, co-regulator distribution, activity of ligand-producing pathways, and others. In particular, a thorough investigation of paracrine signals among the PPAR β/δ -dependent effects on carcinogenesis is still lacking. Thus, a full understanding of context-specific PPAR β/δ activity – in the skin and elsewhere – remains a burning necessity. Hopefully, the availability of diverse new high-throughput technologies allowing global analysis of the interactions between the components of complex biological systems will assist in a step-by-step resolution of today's unsolved problems.

Highlights

- PPAR β/δ is involved in the regulation of key processes in skin biology in fetal and postnatal life.
- PPAR β/δ promotes keratinocyte and sebocyte differentiation and is implicated in hair follicle development.
- The inflammatory response to skin wounds enhances PPAR β/δ expression and triggers the production of PPAR β/δ ligands. Activated PPAR β/δ accelerates wound healing.
- During reepithelialization of the wound bed, PPAR β/δ enhances keratinocyte survival, directional

Wound healing	Cancer
Controlled proliferation	Sustained proliferation
Controlled cell migration (no invasion, no metastasis, no epithelial-mesenchymal transition)	Invasion metastasis (epithelial-mesenchymal transition)
Transient resistance to apoptosis	Resistance to cell death
Angiogenesis and lymphangiogenesis	Angiogenesis and lymphangiogenesis

Table 1 Comparison of the main processes of wound healing and cancer that implicate PPAR β/δ .

- sensing, and migration through cell-autonomous mechanisms.
- During wound healing, keratinocyte proliferation is attenuated through the PPAR β/δ -dependent production of sIL-1Ra by skin fibroblasts, which represses the IL-1 signaling that stimulates mitogenic cytokine production and secretion by these cells.
 - PPAR β/δ expression is increased in psoriasis, and animal model observations indicate the potential usefulness of its topical inhibition to treat this disease.
 - The role of PPAR β/δ in NMSC is ambivalent, and its role in tumor promotion or suppression appears to be context-specific and requires further investigation.

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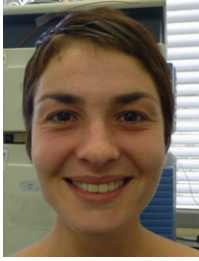
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References

1. Fuchs E. Finding one's niche in the skin. *Cell Stem Cell* 2009; 4: 499–502.
2. Beck B, Blanpain C. [Mechanisms regulating epidermal stem cells](#). *EMBO J* 2012; 31: 2067–75.
3. auf dem Keller U, Krampert M, Kumin A, Braun S, Werner S. Keratinocyte growth factor: effects on keratinocytes and mechanisms of action. *Eur J Cell Biol* 2004; 83: 607–12.
4. Steiling H, Werner S. [Fibroblast growth factors: key players in epithelial morphogenesis, repair and cytoprotection](#). *Curr Opin Biotechnol* 2003; 14: 533–7.
5. Beenken A, Mohammadi M. [The FGF family: biology, pathophysiology and therapy](#). *Nat Rev Drug Discov* 2009; 8: 235–53.
6. Suter MM, Schulze K, Bergman W, Welle M, Roosje P, Muller EJ. The keratinocyte in epidermal renewal and defence. *Vet Dermatol* 2009; 20: 515–32.
7. Desvergne B, Wahli W. Peroxisome proliferator-activated receptors: nuclear control of metabolism. *Endocr Rev* 1999; 20: 649–88.
8. Michalik L, Wahli W. Peroxisome proliferator-activated receptors (PPARs) in skin health, repair and disease. *Biochim Biophys Acta* 2007; 1771: 991–8.
9. Laudet V, Auwerx J, Gustafsson JA, Wahli W. A unified nomenclature system for the nuclear receptor superfamily. *Cell* 1999; 97: 161–3.
10. Michalik L, Auwerx J, Berger JP, Chatterjee VK, Glass CK, Gonzalez FJ, Grimaldi PA, Kadowaki T, Lazar MA, O'Rahilly S, Palmer CN, Plutzky J, Reddy JK, Spiegelman BM, Staels B, Wahli W. International Union of Pharmacology. LXI. Peroxisome proliferator-activated receptors. *Pharmacol Rev* 2006; 58: 726–41.
11. Montagner A, Rando G, Degueurce G, Leuenberger N, Michalik L, Wahli W. New insights into the role of PPARs. *Prostaglandins Leukot Essent Fatty Acids* 2011; 85: 235–43.
12. Braissant O, Foufelle F, Scotto C, Dauca M, Wahli W. Differential expression of peroxisome proliferator-activated receptors (PPARs): tissue distribution of PPAR- α , - β , and - γ in the adult rat. *Endocrinology* 1996; 137: 354–66.
13. Abbott BD. Review of the expression of peroxisome proliferator-activated receptors alpha (PPAR α), beta (PPAR β), and gamma (PPAR γ) in rodent and human development. *Reprod Toxicol* 2009; 27: 246–57.
14. Giroir EE, Hollingshead HE, He P, Zhu B, Perdev GH, Peters JM. Quantitative expression patterns of peroxisome proliferator-activated receptor- β/δ (PPAR β/δ) protein in mice. *Biochem Biophys Res Commun* 2008; 371: 456–61.
15. Wahli W, Michalik L. [PPARs at the crossroads of lipid signaling and inflammation](#). *Trends Endocrinol Metab* 2012; 23: 351–63.
16. Matsuura H, Adachi H, Smart RC, Xu X, Arata J, Jetten AM. Correlation between expression of peroxisome proliferator-activated receptor β and squamous differentiation in epidermal and tracheobronchial epithelial cells. *Mol Cell Endocrinol* 1999; 147: 85–92.
17. Rivier M, Safonova I, Lebrun P, Griffiths CE, Ailhaud G, Michel S. [Differential expression of peroxisome proliferator-activated receptor subtypes during the differentiation of human keratinocytes](#). *J Invest Dermatol* 1998; 111: 1116–21.
18. Westergaard M, Henningsen J, Svendsen ML, Johansen C, Jensen UB, Schroder HD, Kratchmarova I, Berge RK, Iversen L, Bolund L, Kragballe K, Kristiansen K. [Modulation of keratinocyte gene expression and differentiation by PPAR-selective ligands and tetradecylthioacetic acid](#). *J Invest Dermatol* 2001; 116: 702–12.
19. Michalik L, Desvergne B, Tan NS, Basu-Modak S, Escher P, Rieusset J, Peters JM, Kaya G, Gonzalez FJ, Zakany J, Metzger D, Chambon P, Duboule D, Wahli W. Impaired skin wound healing in peroxisome proliferator-activated receptor (PPAR) α and PPAR β mutant mice. *J Cell Biol* 2001; 154: 799–814.
20. Di-Poi N, Desvergne B, Michalik L, Wahli W. Transcriptional repression of peroxisome proliferator-activated receptor β/δ in murine keratinocytes by CCAAT/enhancer-binding proteins. *J Biol Chem* 2005; 280: 38700–10.
21. Tan NS, Shaw NS, Vinckenbosch N, Liu P, Yasmin R, Desvergne B, Wahli W, Noy N. Selective cooperation between fatty acid

- binding proteins and peroxisome proliferator-activated receptors in regulating transcription. *Mol Cell Biol* 2002; 22: 5114–27.
22. Pal M, Tan MJ, Huang RL, Goh YY, Wang XL, Tang MB, Tan NS. Angiopoietin-like 4 regulates epidermal differentiation. *PLoS One* 2011; 6: e25377.
 23. Stenn KS, Paus R. Controls of hair follicle cycling. *Physiol Rev* 2001; 81: 449–94.
 24. Di-Poi N, Ng CY, Tan NS, Yang Z, Hemmings BA, Desvergne B, Michalik L, Wahli W. Epithelium-mesenchyme interactions control the activity of peroxisome proliferator-activated receptor β/δ during hair follicle development. *Mol Cell Biol* 2005; 25: 1696–712.
 25. Trivedi NR, Cong Z, Nelson AM, Albert AJ, Rosamilia LL, Sivarajah S, Gilliland KL, Liu W, Mauger DT, Gabbay RA, Thiboutot DM. Peroxisome proliferator-activated receptors increase human sebum production. *J Invest Dermatol* 2006; 126: 2002–9.
 26. House JS, Zhu S, Ranjan R, Linder K, Smart RC. C/EBP α and C/EBP β are required for Sebocyte differentiation and stratified squamous differentiation in adult mouse skin. *PLoS One* 2010; 5: e9837.
 27. Rosenfield RL, Kentsis A, Deplewski D, Ciletti N. Rat preputial sebocyte differentiation involves peroxisome proliferator-activated receptors. *J Invest Dermatol* 1999; 112: 226–32.
 28. Bastie C, Holst D, Gaillard D, Jehl-Pietri C, Grimaldi PA. Expression of peroxisome proliferator-activated receptor PPAR δ promotes induction of PPAR γ and adipocyte differentiation in 3T3C2 fibroblasts. *J Biol Chem* 1999; 274: 21920–5.
 29. Elias PM. Stratum corneum defensive functions: an integrated view. *J Invest Dermatol* 2005; 125: 183–200.
 30. Feingold KR, Denda M. Regulation of permeability barrier homeostasis. *Clin Dermatol* 2012; 30: 263–8.
 31. Feingold KR. Thematic review series: skin lipids. The role of epidermal lipids in cutaneous permeability barrier homeostasis. *J Lipid Res* 2007; 48: 2531–46.
 32. Man MQ, Barish GD, Schmutz M, Crumrine D, Barak Y, Chang S, Jiang Y, Evans RM, Elias PM, Feingold KR. Deficiency of PPAR β/δ in the epidermis results in defective cutaneous permeability barrier homeostasis and increased inflammation. *J Invest Dermatol* 2008; 128: 370–7.
 33. Schmutz M, Haqq CM, Cairns WJ, Holder JC, Dorsam S, Chang S, Lau P, Fowler AJ, Chuang G, Moser AH, Brown BE, Mao-Qiang M, Uchida Y, Schoonjans K, Auwerx J, Chambon P, Willson TM, Elias PM, Feingold KR. Peroxisome proliferator-activated receptor (PPAR)- β/δ stimulates differentiation and lipid accumulation in keratinocytes. *J Invest Dermatol* 2004; 122: 971–83.
 34. Tan NS, Michalik L, Noy N, Yasmin R, Pacot C, Heim M, Fluhmann B, Desvergne B, Wahli W. Critical roles of PPAR β/δ in keratinocyte response to inflammation. *Genes Dev* 2001; 15: 3263–77.
 35. Tan NS, Michalik L, Di-Poi N, Ng CY, Mermod N, Roberts AB, Desvergne B, Wahli W. Essential role of Smad3 in the inhibition of inflammation-induced PPAR β/δ expression. *EMBO J* 2004; 23: 4211–21.
 36. Tan NS, Icre G, Montagner A, Bordier-ten-Heggeler B, Wahli W, Michalik L. The nuclear hormone receptor peroxisome proliferator-activated receptor β/δ potentiates cell chemotaxis, polarization, and migration. *Mol Cell Biol* 2007; 27: 7161–75.
 37. Tan NS, Michalik L, Desvergne B, Wahli W. Genetic- or transforming growth factor- β 1-induced changes in epidermal peroxisome proliferator-activated receptor β/δ expression dictate wound repair kinetics. *J Biol Chem* 2005; 280: 18163–70.
 38. Frame MC. Src in cancer: deregulation and consequences for cell behaviour. *Biochim Biophys Acta* 2002; 1602: 114–30.
 39. Chu CY, Cha ST, Lin WC, Lu PH, Tan CT, Chang CC, Lin BR, Jee SH, Kuo ML. Stromal cell-derived factor-1 α (SDF-1 α /CXCL12)-enhanced angiogenesis of human basal cell carcinoma cells involves ERK1/2-NF-kappaB/interleukin-6 pathway. *Carcinogenesis* 2009; 30: 205–13.
 40. Hafner C, Landthaler M, Vogt T. Activation of the PI3K/AKT signalling pathway in non-melanoma skin cancer is not mediated by oncogenic PIK3CA and AKT1 hotspot mutations. *Exp Dermatol* 2010; 19: e222–7.
 41. Grieb G, Steffens G, Pallua N, Bernhagen J, Bucala R. Circulating fibrocytes – biology and mechanisms in wound healing and scar formation. *Int Rev Cell Mol Biol* 2011; 291: 1–19.
 42. Gurtner GC, Werner S, Barrandon Y, Longaker MT. Wound repair and regeneration. *Nature* 2008; 453: 314–21.
 43. Chong HC, Tan MJ, Philippe V, Tan SH, Tan CK, Ku CW, Goh YY, Wahli W, Michalik L, Tan NS. Regulation of epithelial-mesenchymal IL-1 signaling by PPAR β/δ is essential for skin homeostasis and wound healing. *J Cell Biol* 2009; 184: 817–31.
 44. Peters JM, Lee SS, Li W, Ward JM, Gavrilova O, Everett C, Reitman ML, Hudson LD, Gonzalez FJ. Growth, adipose, brain, and skin alterations resulting from targeted disruption of the mouse peroxisome proliferator-activated receptor β (δ). *Mol Cell Biol* 2000; 20: 5119–28.
 45. Romanowska M, Reilly L, Palmer CN, Gustafsson MC, Foerster J. Activation of PPAR β/δ causes a psoriasis-like skin disease in vivo. *PLoS One* 2010; 5: e9701.
 46. al Yacoub N, Romanowska M, Krauss S, Schweiger S, Foerster J. PPAR δ is a type 1 IFN target gene and inhibits apoptosis in T cells. *J Invest Dermatol* 2008; 128: 1940–9.
 47. Hack K, Reilly L, Palmer C, Read KD, Norval S, Kime R, Booth K, Foerster J. Skin-targeted inhibition of PPAR β/δ by selective antagonists to treat PPAR β/δ -mediated psoriasis-like skin disease in vivo. *PLoS One* 2012; 7: e37097.
 48. Kuenzli S, Saurat JH. Effect of topical PPAR β/δ and PPAR γ agonists on plaque psoriasis. A pilot study. *Dermatology* 2003; 206: 252–6.
 49. De Benedetto A, Agnihothri R, McGirt LY, Bankova LG, Beck LA. Atopic dermatitis: a disease caused by innate immune defects? *J Invest Dermatol* 2009; 129: 14–30.
 50. Hatano Y, Man MQ, Uchida Y, Crumrine D, Mauro TM, Feingold KR, Elias PM, Holleran WM. Murine atopic dermatitis responds to peroxisome proliferator-activated receptors α and β/δ (but not γ) and liver X receptor activators. *J Allergy Clin Immunol* 2010; 125: 160–9 e1–5.
 51. Ottaviani M, Camera E, Picardo M. Lipid mediators in acne. *Mediators Inflamm* 2010; 2010: pii: 858176, 6 pages.
 52. Elmongy NN, Shaker O. Expression of peroxisome proliferator activator receptor β/δ (PPAR β/δ) in acne vulgaris. *Eur J Dermatol* 2012; 22: 42–5.
 53. Schuster M, Zouboulis CC, Ochsendorf F, Muller J, Thaci D, Bernd A, Kaufmann R, Kippenberger S. Peroxisome proliferator-activated receptor activators protect sebocytes from apoptosis: a new treatment modality for acne? *Br J Dermatol* 2011; 164: 182–6.

54. Michalik L, Wahli W. PPARs mediate lipid signaling in inflammation and cancer. *PPAR Res* 2008; 2008: 134059.
55. Youssef J, Badr M. Peroxisome proliferator-activated receptors and cancer: challenges and opportunities. *Br J Pharmacol* 2011; 164: 68–82.
56. Peters JM, Shah YM, Gonzalez FJ. The role of peroxisome proliferator-activated receptors in carcinogenesis and chemoprevention. *Nat Rev Cancer* 2012; 12: 181–95.
57. Mandard S, Zandbergen F, Tan NS, Escher P, Patsouris D, Koenig W, Kleemann R, Bakker A, Veenman F, Wahli W, Muller M, Kersten S. The direct peroxisome proliferator-activated receptor target fasting-induced adipose factor (FIAF/PGAR/ANGPTL4) is present in blood plasma as a truncated protein that is increased by fenofibrate treatment. *J Biol Chem* 2004; 279: 34411–20.
58. Zhu P, Tan MJ, Huang RL, Tan CK, Chong HC, Pal M, Lam CR, Boukamp P, Pan JY, Tan SH, Kersten S, Li HY, Ding JL, Tan NS. Angiopoietin-like 4 protein elevates the pro-survival intracellular O₂:H₂O₂ ratio and confers anoikis resistance to tumors. *Cancer Cell* 2011; 19: 401–15.
59. Tan MJ, Teo Z, Sng MK, Zhu P, Tan NS. Emerging roles of angiopoietin-like 4 in human cancer. *Mol Cancer Res* 2012; 10: 677–88.
60. Kim SH, Park YY, Kim SW, Lee JS, Wang D, DuBois RN. ANGPTL4 induction by prostaglandin E2 under hypoxic conditions promotes colorectal cancer progression. *Cancer Res* 2011; 71: 7010–20.
61. Zuo X, Peng Z, Moussalli MJ, Morris JS, Broaddus RR, Fischer SM, Shureiqi I. [Targeted genetic disruption of peroxisome proliferator-activated receptor- \$\delta\$ and colonic tumorigenesis.](#) *J Natl Cancer Inst* 2009; 101: 762–7.
62. Shao J, Sheng H, DuBois RN. Peroxisome proliferator-activated receptors modulate K-Ras-mediated transformation of intestinal epithelial cells. *Cancer Res* 2002; 62: 3282–8.
63. Harman FS, Nicol CJ, Marin HE, Ward JM, Gonzalez FJ, Peters JM. [Peroxisome proliferator-activated receptor- \$\delta\$ attenuates colon carcinogenesis.](#) *Nat Med* 2004; 10: 481–3.
64. Nijsten T, Geluyckens E, Colpaert C, Lambert J. [Peroxisome proliferator-activated receptors in squamous cell carcinoma and its precursors.](#) *J Cutan Pathol* 2005; 32: 340–7.
65. Sertznig P, Seifert M, Tilgen W, Reichrath J. Peroxisome proliferator-activated receptors (PPARs) and the human skin: importance of PPARs in skin physiology and dermatologic diseases. *Am J Clin Dermatol* 2008; 9: 15–31.
66. Trifan OC, Hla T. Cyclooxygenase-2 modulates cellular growth and promotes tumorigenesis. *J Cell Mol Med* 2003; 7: 207–22.
67. Bility MT, Devlin-Durante MK, Blazanian N, Glick AB, Ward JM, Kang BH, Kennett MJ, Gonzalez FJ, Peters JM. Ligand activation of peroxisome proliferator-activated receptor β/δ (PPAR β/δ) inhibits chemically induced skin tumorigenesis. *Carcinogenesis* 2008; 29: 2406–14.
68. Kim DJ, Akiyama TE, Harman FS, Burns AM, Shan W, Ward JM, Kennett MJ, Gonzalez FJ, Peters JM. Peroxisome proliferator-activated receptor β (δ)-dependent regulation of ubiquitin C expression contributes to attenuation of skin carcinogenesis. *J Biol Chem* 2004; 279: 23719–27.
69. Kim DJ, Prabhu KS, Gonzalez FJ, Peters JM. Inhibition of chemically induced skin carcinogenesis by sulindac is independent of peroxisome proliferator-activated receptor- β/δ (PPAR β/δ). *Carcinogenesis* 2006; 27: 1105–12.
70. Zhu B, Bai R, Kennett MJ, Kang BH, Gonzalez FJ, Peters JM. Chemoprevention of chemically induced skin tumorigenesis by ligand activation of peroxisome proliferator-activated receptor- β/δ and inhibition of cyclooxygenase 2. *Mol Cancer Ther* 2010; 9: 3267–77.
71. Boukamp P. [Non-melanoma skin cancer: what drives tumor development and progression?](#) *Carcinogenesis* 2005; 26: 1657–67.
72. Matsumura Y, Ananthaswamy HN. Short-term and long-term cellular and molecular events following UV irradiation of skin: implications for molecular medicine. *Expert Rev Mol Med* 2002; 4: 1–22.
73. Lieu FM, Yamanishi K, Konishi K, Kishimoto S, Yasuno H. [Low incidence of Ha-ras oncogene mutations in human epidermal tumors.](#) *Cancer Lett* 1991; 59: 231–5.
74. Pierceall WE, Goldberg LH, Tainsky MA, Mukhopadhyay T, Ananthaswamy HN. [Ras gene mutation and amplification in human nonmelanoma skin cancers.](#) *Mol Carcinog* 1991; 4: 196–202.
75. Campbell C, Quinn AG, Rees JL. Codon 12 Harvey-ras mutations are rare events in non-melanoma human skin cancer. *Br J Dermatol* 1993; 128: 111–4.
76. Berwick M, Wiggins C. [The current epidemiology of cutaneous malignant melanoma.](#) *Front Biosci* 2006; 11: 1244–54.
77. Michiels JF, Perrin C, Leccia N, Massi D, Grimaldi P, Wagner N. PPAR β activation inhibits melanoma cell proliferation involving repression of the Wilms' tumour suppressor WT1. *Pflugers Arch* 2010; 459: 689–703.
78. Schafer M, Werner S. [Cancer as an overhealing wound: an old hypothesis revisited.](#) *Nat Rev Mol Cell Biol* 2008; 9: 628–38.
79. Maas-Szabowski N, Stark HJ, Fusenig NE. Keratinocyte growth regulation in defined organotypic cultures through IL-1-induced keratinocyte growth factor expression in resting fibroblasts. *J Invest Dermatol* 2000; 114: 1075–84.



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