

Basic science for clinicians series

Treating inflammation in atherosclerotic cardiovascular disease: emerging therapies

Roland Klingenberg^{1,2*} and Göran K. Hansson²

¹Department of Cardiology, University Hospital Zurich, CH-8091 Zurich, Switzerland; and ²Karolinska Institutet, Center for Molecular Medicine, Department of Medicine, Karolinska University Hospital, 17176 Stockholm, Sweden

Received 5 August 2009; revised 4 September 2009; accepted 9 October 2009; online publish-ahead-of-print 30 October 2009

Atherosclerosis constitutes the underlying disease to the clinical manifestations of myocardial infarction, stroke, and gangrene. Despite the success of statins, prevention of clinical events of atherosclerosis remains a major challenge in current-day cardiology. Research into the inflammatory nature of atherosclerosis has led to improved mechanistic understanding of its pathogenesis and to the identification of novel therapeutic targets discussed in this review. Recent genetic and epidemiological data document shared pathologies of chronic inflammatory diseases and atherosclerosis. Anti-inflammatory treatment regimens used in these diseases, including tumor necrosis factor- α blockade, IL-1 receptor antagonism, and leukotriene blockade may be beneficial also in patients with coronary artery disease. Enhancing inherent atheroprotective immunity by expansion of regulatory T cells may emerge as a future therapeutic strategy. Immunization strategies directed against atherosclerosis-related antigens such as epitopes within the low-density lipoprotein particle have been extensively studied in animal models and may enter the clinical stage. Success of these novel therapies will be critically dependent on the adequate identification of patients and choice of appropriate clinical endpoints.

Keywords Atherosclerosis • Inflammation • Immunity • Therapies • Acute coronary syndromes

Background

Cardiovascular disease represents the main cause of death and morbidity in the western world and is projected to be the number one killer globally by 2020.¹ Cardiovascular medicine faces a need for effective prevention of myocardial infarction, stroke, and gangrene, which constitute the clinical manifestations of atherosclerosis. Currently, 70% of clinical events cannot be prevented with available drug therapy including statins² and at least 10% of coronary events occur in apparently healthy individuals in the absence of major traditional risk factors.³ Better therapeutic opportunities are also needed to limit the extent of damage inflicted on the heart once acute ischemia has occurred, in order to prevent ensuing congestive heart failure (CHF).

Atherosclerosis is a chronic inflammatory disease: clinical evidence

Recent epidemiological studies demonstrate a significant link between coronary artery disease (CAD) and chronic inflammatory diseases.

An increased incidence of myocardial infarction was found in patients diagnosed with rheumatoid arthritis (RA),⁴ systemic lupus erythematosus,⁵ psoriasis,⁶ and gout.⁷ The discovery of an association between a polymorphism in the promoter of the MHC class II transactivator and increased susceptibility to RA, multiple sclerosis and myocardial infarction⁸ provides evidence for a shared immune-mediated pathophysiology in CAD and chronic inflammatory diseases. Expression of MHC class II molecules is induced by the proinflammatory cytokine interferon- γ on several cell types such as macrophages, endothelial cells, and smooth muscle cells and is a prerequisite for antigen presentation and activation of T cells.

Several other genetic variants with known biological functions in inflammation and atherosclerosis were also reported to be associated with myocardial infarction. The products of these genes are involved in adaptive immunity, such as the tumor necrosis superfamily member OX40 ligand (TNFSF4)⁹ that promotes T cell activation. Mediators of innate immunity comprise interleukin (IL)-1 beta,¹⁰ the cytokine lymphotoxin- α (LTA)¹¹ and components of the leukotriene biosynthesis pathway including arachidonate 5-lipoxygenase (ALOX5), arachidonate 5-lipoxygenase-activating protein (ALOX5AP), and leukotriene A4 hydrolase (LTA4H).^{12–14}

* Corresponding author. Tel: +41 44 255 8700, Fax: +41 44 255 8701, Email: roland.klingenberg@usz.ch or rklingenberg@gmx.de

Published on behalf of the European Society of Cardiology. All rights reserved. © The Author 2009. For permissions please email: journals.permissions@oxfordjournals.org.

Research on chronic inflammatory diseases has pinpointed members of the tumor necrosis factor superfamily (TNFSF) (signature cytokine: TNF- α , signature cell-type: T lymphocyte)¹⁵ and the IL-1 family (signature cytokine: IL-1 β , signature cell type: monocyte) as key mediators.¹⁶ Chronic inflammatory diseases are now categorized into autoimmune diseases centred around the TNFSF and autoinflammatory diseases characterized by a clinical response to IL-1 β antagonism, respectively.¹⁶ Prominent examples of autoinflammatory diseases are gout and type 2 diabetes mellitus.¹⁶ Coronary artery disease appears to share features of both, autoimmune and autoinflammatory diseases. In CAD patients presenting with acute myocardial infarction, elevated blood levels of soluble and cell-bound members of the TNFSF (TNF- α , CD40L, LIGHT, RANKL, OPG, TRAIL)^{17–22} as well as from the IL-1 family (IL-1 β , IL-18 and IL-33)^{23–25} were detected and correlated with the subsequent risk of cardiovascular death or CHF. Soluble members of the TNFSF and IL-1 family are also present in stable CAD patients.^{26,27} Several other inflammatory mediators downstream of the TNF and IL-1 superfamilies were shown to contribute in the pathogenesis of atherosclerosis.²⁸ A full list of abbreviations can be found in the online supplementary data.

Clinical settings and anti-inflammatory treatment options in atherosclerosis

The dismal results of clinical trials aimed at inhibiting inflammation during reperfusion injury in a non-selective manner

(steroids)²⁹ or by targeting neutrophil recruitment³⁰ suggest placing the focus on a different time point. Coronary artery disease patients at high risk for future coronary events despite currently available treatments and evidence of vascular inflammation constitute the most promising study population for novel anti-inflammatory therapies with respect to sample size and tolerated side effects, placing post-ACS patients at centre-stage. Indeed, a trial termed Cardiovascular Inflammation Reduction Trial (CIRT) has recently been designed. It will compare the immunosuppressive drug methotrexate at very low dose against placebo in addition to the standard treatment in secondary prevention patients.³¹ In such trials, cardiovascular death should be the prime endpoint to move the field forward, complemented by biomarkers modifiable by anti-inflammatory therapies and vascular imaging for adequate patient identification and therapeutic monitoring. Figure 1 shows potential therapeutic strategies and time points during the evolution of atherosclerosis. Successful novel therapies will need to target inflammation directly without interfering with the cardiovascular risk profile (i.e. lipids) and have an acceptable safety profile. In light of the pathogenetic similarities between CAD and many autoimmune and autoinflammatory diseases, clinical trials conducted with novel immunomodulatory compounds should include registration of cardiovascular endpoints including traditional risk profiles for *post hoc* analysis. One such example may be the immunomodulatory agent fingolimod (FTY720) for treatment of patients with multiple sclerosis³² as experimental data on the role of FTY720 in atherosclerosis were dependent on the model and experimental conditions used.^{33–35}

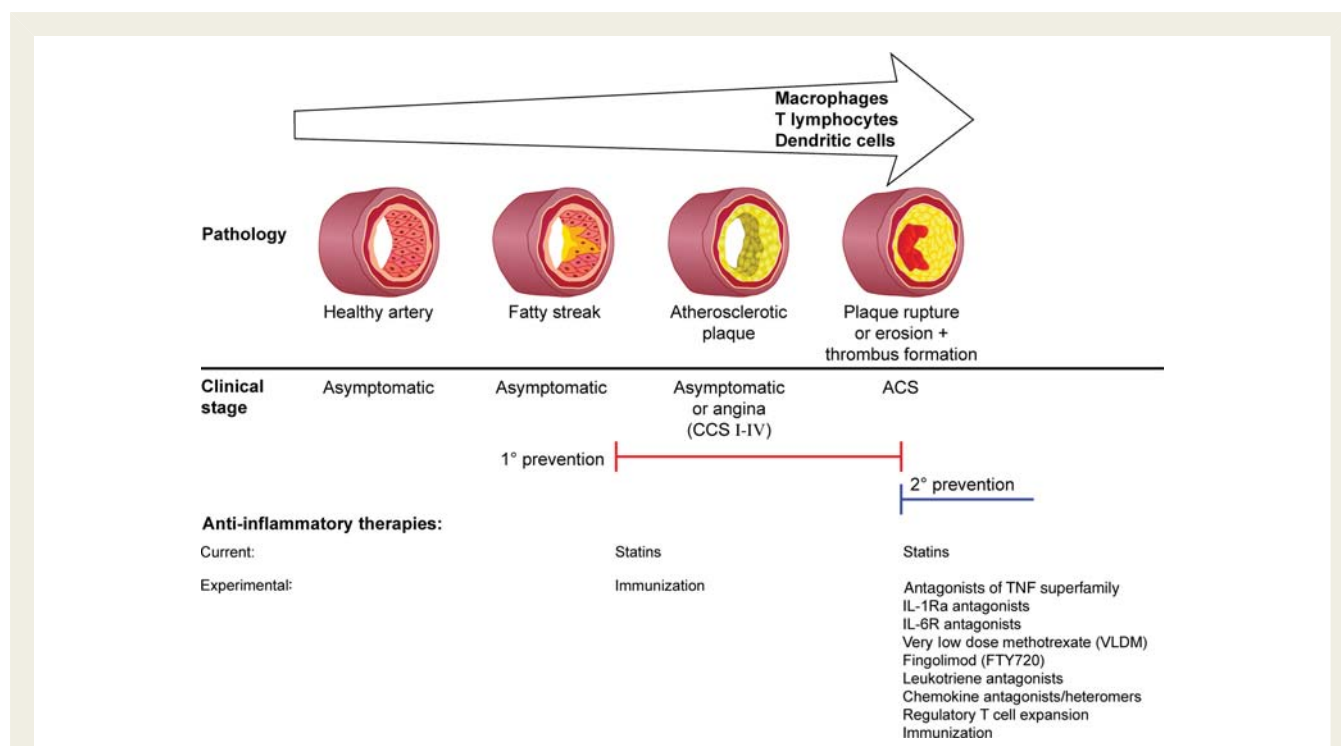


Figure 1 Emerging anti-inflammatory therapies in clinical atherosclerosis. Anti-inflammatory treatment options are shown for the distinct stages in the development of clinical atherosclerosis.

Statins

Statins constitute the best characterized anti-inflammatory class of drugs in primary and secondary prevention of CAD. Beyond their lipid-lowering activity, statins also exert anti-inflammatory effects.³⁶ Clinical evidence for a direct anti-inflammatory effect of statins comes from the *post hoc* C-reactive protein substudies of the PROVE-IT TIMI 22, A to Z, and REVERSAL trials^{37–39} documenting that statin-induced reductions of C-reactive protein and LDL cholesterol levels were only weakly correlated, whereas the decrease in C-reactive protein was significantly correlated with reduced atherosclerosis progression, independent of LDL cholesterol-lowering. The JUPITER trial prospectively confirmed these findings in primary prevention of individuals with elevated C-reactive protein but with low LDL cholesterol.⁴⁰ Further analysis scheduled *a priori* within the JUPITER trial showed that the magnitude of the decrease in C-reactive protein paralleled the magnitude of clinical benefit⁴¹ suggesting a beneficial role of targeting inflammation *per se* in the prevention of cardiovascular events. In addition, the Armyda trial showed that administration of high-dose statins prior to revascularization in ACS patients reduced major adverse cardiovascular events.⁴²

Tumor necrosis factor- α blockade

Tumor necrosis factor blockade has shown efficacy in autoimmune diseases⁴³ and reduced the incidence of cardiovascular events in RA patients, suggesting that attenuated TNF signaling reduces not only RA but also atherosclerosis.⁴⁴ Long-term safety of TNF blockade was documented in RA patients, with no adverse effect on the disease-inherent development of CHF,⁴⁵ adding information to previous reports on CHF patients without RA which showed no adverse effect on hospitalization rates and mortality.⁴⁶ The effects of TNF blockade on plasma lipids require further study.^{47,48} A common feature of TNF antagonists is that they reduce cellularity in inflamed tissues and inhibit expression of pro-inflammatory cytokines and chemokines in addition to TNF- α (IL-1 β , IL-6, IL-8, MCP-1, GM-CSF, VEGF). Furthermore, they dampen the TNF- α -driven production of matrix-degrading enzymes MMP-1 and MMP-3.⁴⁹ These enzymes are considered to be contributing factors to plaque instability.⁵⁰ Tumour necrosis factor antagonism normalized levels of circulating OPG and RANKL in RA patients⁵¹, suggesting an option for therapeutic monitoring. Recent data show that OPG administration can stabilize atherosclerotic plaques in mice.⁵² OPG probably exerts this effect due to its capacity to inhibit RANKL signalling; the latter molecule promotes protease activity and inhibits matrix formation. The pleiotropic cytokine IL-6 acts as a major inducer of the acute-phase response (i.e. C-reactive protein) and impacts on the function of diverse inflammatory and vascular cells. Inhibition of the IL-6 receptor recently shown to be an effective therapeutic in RA patients, however, was associated with elevated lipid levels.⁵³

Interleukin-1 receptor antagonism

Interleukin-1 receptor antagonism (IL-1Ra) has shown beneficial effects in several autoinflammatory diseases¹⁶ and recently also

stroke.⁵⁴ Genetic evidence in a prospective patient cohort documented a significant correlation between a variant in the *IL1-Ra* gene and carotid atherosclerosis.⁵⁵ Experimental data⁵⁶ provided the rationale to conduct clinical trials on the effects of IL-1Ra to prevent post-infarction remodeling (clinicaltrials.gov NCT00789724) and on inflammatory biomarkers in NSTEMI patients, respectively.⁵⁷

Leukotrienes

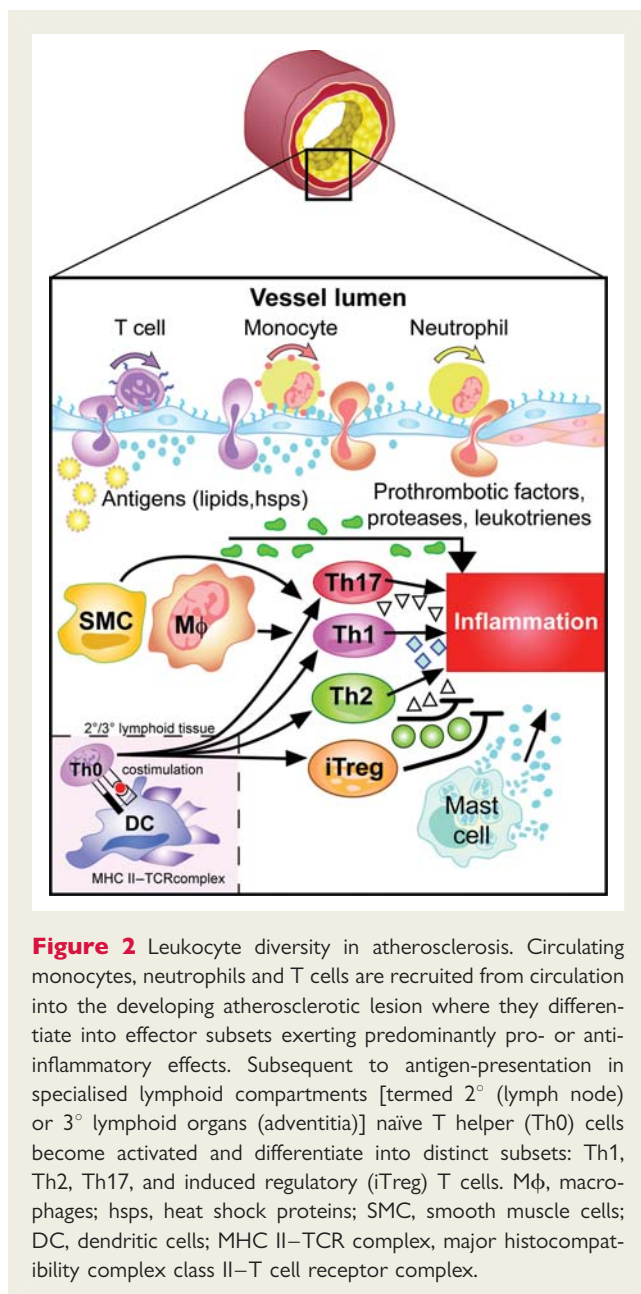
Leukotrienes belong to the family of eicosanoids and constitute potent pro-inflammatory and smooth muscle constrictive lipid mediators.⁵⁸ Activation of the 5-lipoxygenase pathway in patients with acute myocardial infarction was already documented in 1992.⁵⁹ Recent genetic evidence linked polymorphisms in several enzymes of the leukotriene biosynthesis pathways with myocardial infarction comprising ALOX5, ALOX5AP, and LTA4H.^{12–14} Expression of leukotrienes were detected in atherosclerotic plaques and correlated with symptoms of plaque instability.^{60,61} An ongoing trial aims to evaluate the role of cysteinyl-leukotriene blockade on peripheral endothelial function by administering montelukast to patients after an acute coronary event (clinicaltrials.gov NCT00351364). A recent clinical study showed a marked reduction in inflammatory biomarkers after administration of an inhibitor of the 5-lipoxygenase activating protein (FLAP) to patients carrying at-risk variants in the *FLAP* gene or the *LTA4H* gene.⁶² It remains to be determined whether this tailored therapy translates into a reduction in coronary events.

Leukocyte diversity

The concept of inherent atheroprotective immunity⁶³ has paved the way for therapeutic efforts to attenuate atherosclerosis. Increasing experimental evidence documents a role for several types of leukocytes in atherogenesis, including both pro- and anti-inflammatory subtypes (Figure 2). Circulating monocytes and tissue macrophages can be subdivided into 'inflammatory'/'classical' and 'resident'/'non-classical' subtypes based on the expression of chemokine and adhesion molecule receptors.⁶⁴ These differences may offer possibilities for selectively blocking entry of inflammatory monocytes into atherosclerotic lesions and modify plaque composition. The heterogeneity of T lymphocytes reflects their diverse functions in orchestrating adaptive immune reactions. Four types of CD4+ T helper (Th) cells have currently been identified: Th1, Th2, Th17 cells, and the regulatory T (Treg) cell lineage. Identifying the role of the distinct effector CD4+ T cell subsets in atherosclerosis has been of central interest in recent years.⁶⁵ Recent data identified lymphoid tissue in the adventitia as an additional site to secondary lymphoid tissue (lymph nodes) where naïve T helper (Th0) cells may become activated through the process of antigen presentation and co-stimulation.^{66,67}

Expansion of regulatory T cells

A deficiency in Treg cells in terms of number and/or function was shown in patients with a variety of autoimmune diseases that led to the concept that expansion of these cells may attenuate disease activity. Reduced numbers were reported in blood from ACS



patients⁶⁸ and Treg cells were detected in all stages of atherosclerotic lesions.⁶⁹ Expansion of the Treg cell pool can be achieved either by promoting Treg cell development and survival *in vivo* by administering drugs or by adoptive transfer of Treg cells following *ex vivo* expansion.

Drugs designed to target surface molecules on T cells selectively deplete activated effector T cells while promoting Treg cell expansion *in vivo*. A monoclonal antibody directed at the CD3- ϵ chain of T cells showed remarkable efficacy in type 1 diabetic patients⁷⁰ and reduced atherosclerosis in mice.⁷¹ A critical role was recently identified for the co-stimulatory molecules ICOS, PD-1, OX40L, and CD137 in Treg and cytotoxic T cell function in mice,^{72–75} suggesting further targets for modulation of immune homeostasis in atherosclerosis. Targeting the CD40-CD40L pathway appears cumbersome as treatment directed against CD40L produced

thrombosis *in vivo*,⁷⁶ highlighting the need for characterization of the cellular expression pattern and functional aspects of targeting co-stimulatory molecules.

Cytokine administration *in vivo* may be a valid short-term strategy to enhance Treg cells. Interleukin-10 and TGF- β have important roles in Treg cell generation and function. Administration of IL-10 was safe in phase II trials in subjects with psoriasis. TGF- β , however, appears as a less interesting candidate due to its pleiotropic effects.

Adoptive immunotherapy to rapidly increase the circulating Treg cell pool by re-infusion of autologous Treg cells after *in vitro* expansion constitutes an interesting approach. However, several hurdles have to be overcome before translation into the clinics. A pivotal step is the identification and isolation of Treg cells, which poses a major challenge in light of the heterogeneity of Treg cells and the absence of a unifying surface marker. Cell-sorting for CD4⁺ CD25^{high} CD127^{low} T cells may constitute a valid approach.⁷⁷ In order to provide a disease-specific therapy, re-infusion of antigen-specific Treg cells directed against antigens relevant in atherosclerosis can be achieved by *in vitro* expansion of isolated antigen-specific (adaptive) Treg cells, by means of expanding natural Treg cells isolated from patients against specific antigens *in vitro* or by induced expansion of naïve T cells against specific antigens under tolerable conditions *in vitro*. Several protocols were shown to effectively expand Treg cells *in vitro*.⁷⁸ Introduction of inducible suicide genes allows the control of potential unwanted *in vivo* effects such as leukaemia or generalized immunosuppression upon re-transfer of Treg cells into the host.

Atherosclerosis-specific immunization

Immunization has emerged as a promising therapeutic regimen against atherosclerosis enhancing protective antibody titers, altering the balance of pro- and anti-inflammatory T cell subtypes and expanding Treg cells. Several antigens have been identified and investigated for immunization against atherosclerosis in animal models using active immunization or antibody infusion.⁷⁹ Among those, epitopes recognized in the LDL particle including apolipoprotein B-100 appear most interesting from a clinical perspective in light of the role of LDL in the pathogenesis of atherosclerosis. In order to translate those findings to the clinics, however, antigens that can be easily manufactured under good manufacturing practice conditions and that have a reproducible quality without the risk of contamination are mandatory.

Clearly, more research into the cellular and inflammatory components at different stages of human CAD is needed to identify therapeutic targets, inflammatory biomarkers, and imaging modalities suitable for improved identification of patients and monitoring of anti-inflammatory therapies. Nonetheless, the stage is set to provide a rationale for anti-inflammatory therapies and several clinical studies are currently underway.

Supplementary material

Supplementary material is available at *European Heart Journal* online.

Funding

R.K. was supported by grants from the German Research Foundation (KL1398/2-2) and the Swiss National Fund (SPUM 33CM30-124112). G.K.H. received funding from the Swedish Heart-Lung Foundation, Swedish Research Council, European Commission (projects EVGN Molstroke, Eicosanox, CVDImmune, Immunath, and AtheroRemo), and the Leducq Transatlantic Network on Atherothrombosis.

Conflict of interest: none declared.

References

- Lopez AD, Mathers CD, Ezzati M, Jamison DT, Murray CJ. Global and regional burden of disease and risk factors, 2001: systematic analysis of population health data. *Lancet* 2006;**367**:1747–1757.
- Baigent C, Keech A, Kearney PM, Blackwell L, Buck G, Pollicino C, Kirby A, Sourjina T, Peto R, Collins R, Simes R. Efficacy and safety of cholesterol-lowering treatment: prospective meta-analysis of data from 90,056 participants in 14 randomised trials of statins. *Lancet* 2005;**366**:1267–1278.
- Greenland P, Knoll MD, Stamler J, Neaton JD, Dyer AR, Garside DB, Wilson PW. Major risk factors as antecedents of fatal and nonfatal coronary heart disease events. *JAMA* 2003;**290**:891–897.
- Maradit-Kremers H, Nicola PJ, Crowson CS, Ballman KV, Gabriel SE. Cardiovascular death in rheumatoid arthritis: a population-based study. *Arthritis Rheum* 2005;**52**:722–732.
- Asanuma Y, Oeser A, Shintani AK, Turner E, Olsen N, Fazio S, Linton MF, Raggi P, Stein CM. Premature coronary-artery atherosclerosis in systemic lupus erythematosus. *N Engl J Med* 2003;**349**:2407–2415.
- Gelfand JM, Neimann AL, Shin DB, Wang X, Margolis DJ, Troxel AB. Risk of myocardial infarction in patients with psoriasis. *JAMA* 2006;**296**:1735–1741.
- Choi HK, Curhan G. Independent impact of gout on mortality and risk for coronary heart disease. *Circulation* 2007;**116**:894–900.
- Swanberg M, Lidman O, Padyukov L, Eriksson P, Akesson E, Jagodic M, Lobell A, Khademi M, Borjesson O, Lindgren CM, Lundman P, Brookes AJ, Kere J, Luthman H, Alfredsson L, Hillert J, Klareskog L, Hamsten A, Piel F, Olsson T. MHC2TA is associated with differential MHC molecule expression and susceptibility to rheumatoid arthritis, multiple sclerosis and myocardial infarction. *Nat Genet* 2005;**37**:486–494.
- Wang X, Ria M, Kelmenson PM, Eriksson P, Higgins DC, Samnegard A, Petros C, Rollins J, Bennet AM, Wiman B, de Faire U, Wennberg C, Olsson PG, Ishii N, Sugamura K, Hamsten A, Forsman-Semb K, Lagercrantz J, Paigen B. Positional identification of TNFSF4, encoding OX40 ligand, as a gene that influences atherosclerosis susceptibility. *Nat Genet* 2005;**37**:365–372.
- Iacoviello L, Di Castelnuovo A, Gattone M, Pezzini A, Assanelli D, Lorenzet R, Del Zotto E, Colombo M, Napoleone E, Amore C, D'Orazio A, Padovani A, de Gaetano G, Giannuzzi P, Donati MB. Polymorphisms of the interleukin-1beta gene affect the risk of myocardial infarction and ischemic stroke at young age and the response of mononuclear cells to stimulation in vitro. *Arterioscler Thromb Vasc Biol* 2005;**25**:222–227.
- Ozaki K, Ohnishi Y, Iida A, Sekine A, Yamada R, Tsunoda T, Sato H, Hori M, Nakamura Y, Tanaka T. Functional SNPs in the lymphotoxin-alpha gene that are associated with susceptibility to myocardial infarction. *Nat Genet* 2002;**32**:650–654.
- Dwyer JH, Allayee H, Dwyer KM, Fan J, Wu H, Mar R, Lusi AJ, Mehrabian M. Arachidonate 5-lipoxygenase promoter genotype, dietary arachidonic acid, and atherosclerosis. *N Engl J Med* 2004;**350**:29–37.
- Helgadottir A, Manolescu A, Thorleifsson G, Gretarsdottir S, Jonsdottir H, Thorsteinsdottir U, Samani NJ, Gudmundsson G, Grant SF, Thorgeirsson G, Sveinbjornsdottir S, Valdimarsson EM, Matthiasson SE, Johannsson H, Gudmundsdottir O, Gurney ME, Sainz J, Thorhallsdottir M, Andresdottir M, Frigge ML, Topol EJ, Kong A, Gudnason V, Hakonarson H, Gulcher JR, Stefansson K. The gene encoding 5-lipoxygenase activating protein confers risk of myocardial infarction and stroke. *Nat Genet* 2004;**36**:233–239.
- Helgadottir A, Manolescu A, Helgason A, Thorleifsson G, Thorsteinsdottir U, Gudbjartsson DF, Gretarsdottir S, Magnusson KP, Gudmundsson G, Hicks A, Jonsson T, Grant SF, Sainz J, O'Brien SJ, Sveinbjornsdottir S, Valdimarsson EM, Matthiasson SE, Levey AI, Abramson JL, Reilly MP, Vaccarino V, Wolfe ML, Gudnason V, Quyyumi AA, Topol EJ, Rader DJ, Thorgeirsson G, Gulcher JR, Hakonarson H, Kong A, Stefansson K. A variant of the gene encoding leukotriene A4 hydrolase confers ethnicity-specific risk of myocardial infarction. *Nat Genet* 2006;**38**:68–74.
- Watts TH. TNF/TNFR family members in costimulation of T cell responses. *Annu Rev Immunol* 2005;**23**:23–68.
- Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. *Annu Rev Immunol* 2009;**27**:519–550.
- Ridker PM, Rifai N, Pfeffer M, Sacks F, Lepage S, Braunwald E. Elevation of tumor necrosis factor-alpha and increased risk of recurrent coronary events after myocardial infarction. *Circulation* 2000;**101**:2149–2153.
- Heeschen C, Dimmeler S, Hamm CW, van den Brand MJ, Boersma E, Zeiher AM, Simoons-Sel A. Soluble CD40 ligand in acute coronary syndromes. *N Engl J Med* 2003;**348**:1104–1111.
- Otterdal K, Smith C, Oie E, Pedersen TM, Yndestad A, Stang E, Endresen K, Solum NO, Aukrust P, Damas JK. Platelet-derived LIGHT induces inflammatory responses in endothelial cells and monocytes. *Blood* 2006;**108**:928–935.
- Sandberg WJ, Yndestad A, Oie E, Smith C, Ueland T, Ovchinnikova O, Robertson AK, Muller F, Semb AG, Scholz H, Andreassen AK, Gullestad L, Damas JK, Froland SS, Hansson GK, Halvorsen B, Aukrust P. Enhanced T-cell expression of RANK ligand in acute coronary syndrome: possible role in plaque destabilization. *Arterioscler Thromb Vasc Biol* 2006;**26**:857–863.
- Omeland T, Ueland T, Jansson AM, Persson A, Karlsson T, Smith C, Herlitz J, Aukrust P, Hartford M, Caidahl K. Circulating osteoprotegerin levels and long-term prognosis in patients with acute coronary syndromes. *J Am Coll Cardiol* 2008;**51**:627–633.
- Secchiero P, Corallini F, Cecconi C, Parrinello G, Volpato S, Ferrari R, Zauli G. Potential prognostic significance of decreased serum levels of TRAIL after acute myocardial infarction. *PLoS ONE* 2009;**4**:e4442.
- Waehre T, Yndestad A, Smith C, Haug T, Tunheim SH, Gullestad L, Froland SS, Semb AG, Aukrust P, Damas JK. Increased expression of interleukin-1 in coronary artery disease with downregulatory effects of HMG-CoA reductase inhibitors. *Circulation* 2004;**109**:1966–1972.
- Mallat Z, Henry P, Fressonnet R, Alouani S, Scoazec A, Beauvais P, Chvatchko Y, Tedgui A. Increased plasma concentrations of interleukin-18 in acute coronary syndromes. *Heart* 2002;**88**:467–469.
- Shimpo M, Morrow DA, Weinberg EO, Sabatine MS, Murphy SA, Antman EM, Lee RT. Serum levels of the interleukin-1 receptor family member ST2 predict mortality and clinical outcome in acute myocardial infarction. *Circulation* 2004;**109**:2186–2190.
- Blankenberg S, McQueen MJ, Smieja M, Pogue J, Balion C, Lonn E, Rupprecht HJ, Bickel C, Tietel L, Cambien F, Gerstein H, Munzel T, Yusuf S. Comparative impact of multiple biomarkers and N-Terminal pro-brain natriuretic peptide in the context of conventional risk factors for the prediction of recurrent cardiovascular events in the Heart Outcomes Prevention Evaluation (HOPE) Study. *Circulation* 2006;**114**:201–208.
- Semb AG, Ueland T, Aukrust P, Wareham NJ, Luben R, Gullestad L, Kastelein JJ, Khaw KT, Boekholdt SM. Osteoprotegerin and soluble receptor activator of nuclear factor-kappaB ligand and risk for coronary events: a nested case-control approach in the prospective EPIC-Norfolk population study 1993–2003. *Arterioscler Thromb Vasc Biol* 2009;**29**:975–980.
- Tedgui A, Mallat Z. Cytokines in atherosclerosis: pathogenic and regulatory pathways. *Physiol Rev* 2006;**86**:515–581.
- Roberts R, DeMello V, Sobel BE. Deleterious effects of methylprednisolone in patients with myocardial infarction. *Circulation* 1976;**53**(Suppl. 3):I204–I206.
- Yellon DM, Hausenloy DJ. Myocardial reperfusion injury. *N Engl J Med* 2007;**357**:1121–1135.
- Ridker PM. Testing the inflammatory hypothesis of atherothrombosis: scientific rationale for the cardiovascular inflammation reduction trial (CIRT). *J Thromb Haemost* 2009;**7**(Suppl. 1):332–339.
- Kappos L, Antel J, Comi G, Montalban X, O'Connor P, Polman CH, Haas T, Korn AA, Karlsson G, Radue EW. Oral fingolimod (FTY720) for relapsing multiple sclerosis. *N Engl J Med* 2006;**355**:1124–1140.
- Nofer JR, Bot M, Brodde M, Taylor PJ, Salm P, Brinkmann V, van Berkel T, Assmann G, Biessen EA. FTY720, a synthetic sphingosine 1-phosphate analogue, inhibits development of atherosclerosis in low-density lipoprotein receptor-deficient mice. *Circulation* 2007;**115**:501–508.
- Keul P, Tolle M, Lucke S, von Wnuck Lipinski K, Heusch G, Schuchardt M, van der Giet M, Levkau B. The sphingosine-1-phosphate analogue FTY720 reduces atherosclerosis in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* 2007;**27**:607–613.
- Klingenberg R, Nofer JR, Rudling M, Bea F, Blessing E, Preusch M, Grone HJ, Katus HA, Hansson GK, Dengler TJ. Sphingosine-1-phosphate analogue FTY720 causes lymphocyte redistribution and hypercholesterolemia in ApoE-deficient mice. *Arterioscler Thromb Vasc Biol* 2007;**27**:2392–2399.
- Steffens S, Mach F. Drug insight: immunomodulatory effects of statins—potential benefits for renal patients? *Nat Clin Pract Nephrol* 2006;**2**:378–387.

37. Ridker PM, Cannon CP, Morrow D, Rifai N, Rose LM, McCabe CH, Pfeffer MA, Braunwald E. C-reactive protein levels and outcomes after statin therapy. *N Engl J Med* 2005;**352**:20–28.
38. Morrow DA, de Lemos JA, Sabatine MS, Wiviott SD, Blazing MA, Shui A, Rifai N, Califf RM, Braunwald E. Clinical relevance of C-reactive protein during follow-up of patients with acute coronary syndromes in the Aggrastat-to-Zocor Trial. *Circulation* 2006;**114**:281–288.
39. Nissen SE, Tuzcu EM, Schoenhagen P, Crowe T, Sasiela WJ, Tsai J, Orazem J, Magorien RD, O'Shaughnessy C, Ganz P. Statin therapy, LDL cholesterol, C-reactive protein, and coronary artery disease. *N Engl J Med* 2005;**352**:29–38.
40. Ridker PM, Danielson E, Fonseca FA, Genest J, Gotto AM Jr, Kastelein JJ, Koenig W, Libby P, Lorenzatti AJ, MacFadyen JG, Nordestgaard BG, Shepherd J, Willerson JT, Glynn RJ. Rosuvastatin to prevent vascular events in men and women with elevated C-reactive protein. *N Engl J Med* 2008;**359**:2195–2207.
41. Ridker PM, Danielson E, Fonseca FA, Genest J, Gotto AM Jr, Kastelein JJ, Koenig W, Libby P, Lorenzatti AJ, MacFadyen JG, Nordestgaard BG, Shepherd J, Willerson JT, Glynn RJ. Reduction in C-reactive protein and LDL cholesterol and cardiovascular event rates after initiation of rosuvastatin: a prospective study of the JUPITER trial. *Lancet* 2009;**373**:1175–1182.
42. Patti G, Pasceri V, Colonna G, Miglionico M, Fischetti D, Sardella G, Montinaro A, Di Sciascio G. Atorvastatin pretreatment improves outcomes in patients with acute coronary syndromes undergoing early percutaneous coronary intervention: results of the ARMYDA-ACS randomized trial. *J Am Coll Cardiol* 2007;**49**:1272–1278.
43. Tracey D, Klareskog L, Sasso EH, Salfeld JG, Tak PP. Tumor necrosis factor antagonist mechanisms of action: a comprehensive review. *Pharmacol Ther* 2008;**117**:244–279.
44. Jacobsson LT, Turesson C, Gulfe A, Kapetanovic MC, Petersson IF, Saxne T, Geborek P. Treatment with tumor necrosis factor blockers is associated with a lower incidence of first cardiovascular events in patients with rheumatoid arthritis. *J Rheumatol* 2005;**32**:1213–1218.
45. Listing J, Strangfeld A, Kekow J, Schneider M, Kapelle A, Wassenberg S, Zink A. Does tumor necrosis factor alpha inhibition promote or prevent heart failure in patients with rheumatoid arthritis? *Arthritis Rheum* 2008;**58**:667–677.
46. Mann DL, McMurray JJ, Packer M, Swedberg K, Borer JS, Colucci WS, Djian J, Drexler H, Feldman A, Kober L, Krum H, Liu P, Nieminen M, Tavazzi L, van Veldhuisen DJ, Waldenstrom A, Warren M, Westheim A, Zannad F, Fleming T. Targeted anticytokine therapy in patients with chronic heart failure: results of the Randomized Etorcept Worldwide Evaluation (RENEWAL). *Circulation* 2004;**109**:1594–1602.
47. Serio B, Paolino S, Sulli A, Facciolo D, Cutolo M. Effects of anti-TNF-alpha treatment on lipid profile in patients with active rheumatoid arthritis. *Ann N Y Acad Sci* 2006;**1069**:414–419.
48. van Eijk IC, de Vries MK, Levels JH, Peters MJ, Huizer EE, Dijkman BA, van der Horst-Bruinsma IE, Hazenberg BP, van de Stadt RJ, Wolbink GJ, Nurmohamed MT. Improvement of lipid profile is accompanied by atheroprotective alterations in high-density lipoprotein composition upon tumor necrosis factor blockade: a prospective cohort study in ankylosing spondylitis. *Arthritis Rheum* 2009;**60**:1324–1330.
49. Catrina AI, Lampa J, Ernestam S, af Klint E, Bratt J, Klareskog L, Ulfgren AK. Anti-tumor necrosis factor (TNF)-alpha therapy (etanercept) down-regulates serum matrix metalloproteinase (MMP)-3 and MMP-1 in rheumatoid arthritis. *Rheumatology (Oxford)* 2002;**41**:484–489.
50. Aikawa M, Rabkin E, Sugiyama S, Voglic SJ, Fukumoto Y, Furukawa Y, Shiomi M, Schoen FJ, Libby P. An HMG-CoA reductase inhibitor, cerivastatin, suppresses growth of macrophages expressing matrix metalloproteinases and tissue factor in vivo and in vitro. *Circulation* 2001;**103**:276–283.
51. Ziolkowska M, Kurowska M, Radzikowska A, Luszczkiewicz G, Wiland P, Dziewczopolski W, Filipowicz-Sosnowska A, Pazdur J, Szechinski J, Kowalczewski J, Rell-Bakalarska M, Maslinski W. High levels of osteoprotegerin and soluble receptor activator of nuclear factor kappa B ligand in serum of rheumatoid arthritis patients and their normalization after anti-tumor necrosis factor alpha treatment. *Arthritis Rheum* 2002;**46**:1744–1753.
52. Ovchinnikova O, Gylfe A, Bailey L, Nordstrom A, Rudling M, Jung C, Bergstrom S, Waldenstrom A, Hansson GK, Nordstrom P. Osteoprotegerin promotes fibrous cap formation in atherosclerotic lesions of ApoE-deficient mice. *Arterioscler Thromb Vasc Biol* 2009;**29**:1478–1480.
53. Smolen JS, Beaulieu A, Rubbert-Roth A, Ramos-Remus C, Rovinsky J, Alecock E, Woodworth T, Alten R. Effect of interleukin-6 receptor inhibition with tocilizumab in patients with rheumatoid arthritis (OPTION study): a double-blind, placebo-controlled, randomised trial. *Lancet* 2008;**371**:987–997.
54. Emsley HC, Smith CJ, Georgiou RF, Vail A, Hopkins SJ, Rothwell NJ, Tyrrell PJ. A randomised phase II study of interleukin-1 receptor antagonist in acute stroke patients. *J Neurol Neurosurg Psychiatry* 2005;**76**:1366–1372.
55. Markus HS, Labrum R, Bevan S, Reindl M, Egger G, Wiedermann CJ, Xu Q, Kiehl S, Willeit J. Genetic and acquired inflammatory conditions are synergistically associated with early carotid atherosclerosis. *Stroke* 2006;**37**:2253–2259.
56. Abbate A, Salloum FN, Vecile E, Das A, Hoke NN, Straino S, Biondi-Zoccai GG, Houser JE, Qureshi IZ, Ownby ED, Gustini E, Biasucci LM, Severino A, Capogrossi MC, Vetovec GW, Crea F, Baldi A, Kukreja RC, Dobrina A. Anakinra a recombinant human interleukin-1 receptor antagonist inhibits apoptosis in experimental acute myocardial infarction. *Circulation* 2008;**117**:2670–2683.
57. Crossman DC, Morton AC, Gunn JP, Greenwood JP, Hall AS, Fox KA, Lucking AJ, Flather MD, Lees B, Foley CE. Investigation of the effect of interleukin-1 receptor antagonist (IL-1ra) on markers of inflammation in non-ST elevation acute coronary syndromes (The MRC-ILA-HEART Study). *Trials* 2008;**9**: 8.
58. Radmark O, Samuelsson B. 5-lipoxygenase: regulation and possible involvement in atherosclerosis. *Prostaglandins Other Lipid Mediat* 2007;**83**:162–174.
59. Carry M, Korley V, Willerson JT, Weigelt L, Ford-Hutchinson AW, Tagari P. Increased urinary leukotriene excretion in patients with cardiac ischemia. In vivo evidence for 5-lipoxygenase activation. *Circulation* 1992;**85**:230–236.
60. Qiu H, Gabrielsen A, Agardh HE, Wan M, Wetterholm A, Wong CH, Hedin U, Swedenborg J, Hansson GK, Samuelsson B, Paulsson-Berne G, Haeggstrom JZ. Expression of 5-lipoxygenase leukotriene A4 hydrolase in human atherosclerotic lesions correlates with symptoms of plaque instability. *Proc Natl Acad Sci USA* 2006;**103**:8161–8166.
61. Back M, Bu DX, Brannstrom R, Sheikine Y, Yan ZQ, Hansson GK. Leukotriene B4 signaling through NF-kappaB-dependent BLT1 receptors on vascular smooth muscle cells in atherosclerosis and intimal hyperplasia. *Proc Natl Acad Sci USA* 2005;**102**:17501–17506.
62. Hakonarson H, Thorvaldsson S, Helgadóttir A, Gudbjartsson D, Zink F, Andresdóttir M, Manolescu A, Arnar DO, Andersen K, Sigurdsson A, Thorgeirsson G, Jonsson A, Agnarsson U, Bjornsdóttir H, Gottskalksson G, Einarsson A, Gudmundsdóttir H, Adalsteinsdóttir AE, Gudmundsson K, Kristjánsson K, Hardarson T, Kristinnsson A, Topol EJ, Gulcher J, Kong A, Gurney M, Stefansson K. Effects of a 5-lipoxygenase-activating protein inhibitor on biomarkers associated with risk of myocardial infarction: a randomized trial. *JAMA* 2005;**293**:2245–2256.
63. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* 2005;**352**:1685–1695.
64. Weber C, Zernecke A, Libby P. The multifaceted contributions of leukocyte subsets to atherosclerosis: lessons from mouse models. *Nat Rev Immunol* 2008;**8**:802–815.
65. Ait-Oufella H, Taleb S, Mallat Z, Tedgui A. Cytokine network and T cell immunity in atherosclerosis. *Semin Immunopathol* 2009;**31**:23–33.
66. Galkina E, Kadl A, Sanders J, Varughese D, Sarembock IJ, Ley K. Lymphocyte recruitment into the aortic wall before and during development of atherosclerosis is partially L-selectin dependent. *J Exp Med* 2006;**203**:1273–1282.
67. Grabner R, Lotzer K, Dopping S, Hildner M, Radke D, Beer M, Spanbroek R, Lippert B, Reardon CA, Getz GS, Fu YX, Hehlhans T, Mebius RE, van der Wall M, Kruspe D, Englert C, Lovas A, Hu D, Randolph GJ, Weih F, Habenicht AJ. Lymphotoxin beta receptor signaling promotes tertiary lymphoid organogenesis in the aorta adventitia of aged ApoE^{-/-} mice. *J Exp Med* 2009;**206**:233–248.
68. Mor A, Luboshits G, Planer D, Keren G, George J. Altered status of CD4(+)CD25(+) regulatory T cells in patients with acute coronary syndromes. *Eur Heart J* 2006;**27**:2530–2537.
69. de Boer OJ, van der Meer JJ, Teeling P, van der Loos CM, van der Wal AC. Low numbers of FOXP3 positive regulatory T cells are present in all developmental stages of human atherosclerotic lesions. *PLoS ONE* 2007;**2**:e779.
70. Herold KC, Gitelman SE, Masharani U, Hagopian W, Bisikirska B, Donaldson D, Rother K, Diamond B, Harlan DM, Bluestone JA. A single course of anti-CD3 monoclonal antibody hOKT3gamma1(Ala-Ala) results in improvement in C-peptide responses and clinical parameters for at least 2 years after onset of type 1 diabetes. *Diabetes* 2005;**54**:1763–1769.
71. Steffens S, Burger F, Pelli G, Dean Y, Elson G, Kosco-Vilbois M, Chatenoud L, Mach F. Short-term treatment with anti-CD3 antibody reduces the development and progression of atherosclerosis in mice. *Circulation* 2006;**114**:1977–1984.
72. Gotsman I, Gräbie N, Gupta R, Dacosta R, MacConmara M, Lederer J, Sukhova G, Witztum JL, Sharpe AH, Lichtman AH. Impaired regulatory T-cell response and enhanced atherosclerosis in the absence of inducible costimulatory molecule. *Circulation* 2006;**114**:2047–2055.
73. Gotsman I, Gräbie N, Dacosta R, Sukhova G, Sharpe A, Lichtman AH. Proatherogenic immune responses are regulated by the PD-1/CD28 pathway in mice. *J Clin Invest* 2007;**117**:2974–2982.
74. van Wanrooij EJ, van Puijvelde GH, de Vos P, Yagita H, van Berkel TJ, Kuiper J. Interruption of the Tnfrsf4/Tnfrsf4 (OX40/OX40L) pathway attenuates

- atherogenesis in low-density lipoprotein receptor-deficient mice. *Arterioscler Thromb Vasc Biol* 2007;**27**:204–210.
75. Olofsson PS, Soderstrom LA, Wagsater D, Sheikine Y, Ocaya P, Lang F, Rabu C, Chen L, Rudling M, Aukrust P, Hedin U, Paulsson-Berne G, Sirsjo A, Hansson GK. CD137 is expressed in human atherosclerosis and promotes development of plaque inflammation in hypercholesterolemic mice. *Circulation* 2008;**117**: 1292–1301.
76. Sidiropoulos PI, Boumpas DT. Lessons learned from anti-CD40L treatment in systemic lupus erythematosus patients. *Lupus* 2004;**13**:391–397.
77. Seddiki N, Santner-Nanan B, Martinson J, Zaunders J, Sasson S, Landay A, Solomon M, Selby W, Alexander SI, Nanan R, Kelleher A, Fazekas de St Groth B. Expression of interleukin (IL)-2 and IL-7 receptors discriminates between human regulatory and activated T cells. *J Exp Med* 2006;**203**:1693–1700.
78. Gregori S, Bacchetta R, Passerini L, Levings MK, Roncarolo MG. Isolation, expansion, and characterization of human natural and adaptive regulatory T cells. *Methods Mol Biol* 2007;**380**:83–105.
79. Hansson GK, Nilsson J. Vaccination against atherosclerosis? Induction of athero-protective immunity. *Semin Immunopathol* 2009;**31**:95–101.

CARDIOVASCULAR FLASHLIGHT

doi:10.1093/eurheartj/ehp407

Online publish-ahead-of-print 12 October 2009

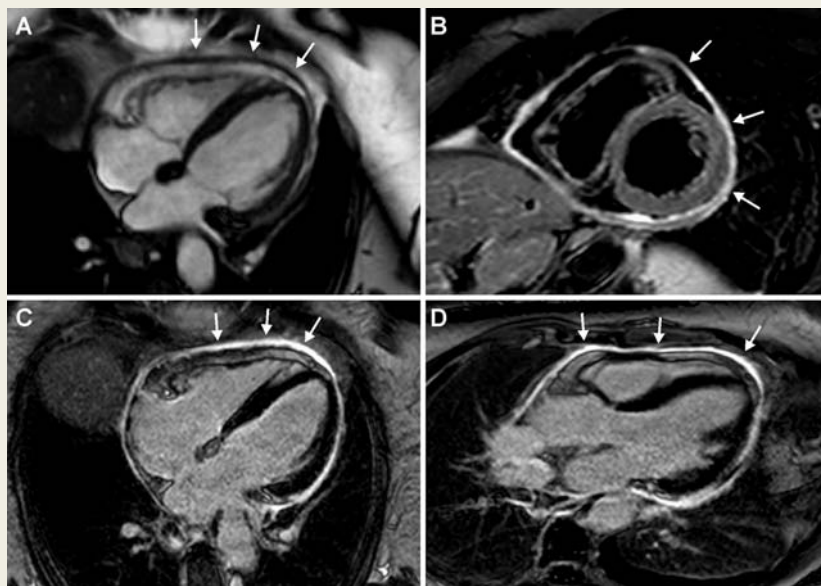
Acute viral pericarditis without typical electrocardiographic changes assessed by cardiac magnetic resonance imaging

Ralf Koos*, Jörg Schröder, and Harald Peter Kühl

Department of Cardiology, Medical Faculty, RWTH Aachen University, Pauwelsstrasse 30, 52074 Aachen, Germany

* Corresponding author. Tel: 49 241 8035443, Fax: 49 241 8082545, Email: rkoos@ukaachen.de

A 59-year-old female was admitted with tachycardia and atypical chest pain. Physical examination was normal. The initial electrocardiogram (ECG) showed atrial fibrillation with a heart rate of 140/min. Laboratory results on admission were unremarkable except for a markedly elevated C-reactive protein of 180 mg/dL (normal value <5 mg/dL). Troponin T levels on admission and follow-up were negative. A follow-up ECG after 4 h documented a spontaneous conversion in sinus rhythm and normal ST-segments. Echocardiography showed a normal left ventricular function (ejection fraction 60%) with no regional wall-motion abnormalities. A small circular pericardial effusion was noted. Additional laboratory tests revealed an acute parvovirus B19 infection by detecting parvovirus B19-specific IgM antibodies in the serum.



Cardiac magnetic resonance (CMR) imaging was scheduled to rule out acute myocarditis. A thickened pericardium (5.5 mm) and a small pericardial effusion were noted at cine-imaging (Panel A). At T2-weighted imaging (Panel B), a hyperintense signal from the thickened pericardium was noted suggesting pericardial oedema. No signs of myocardial oedema were present. Ten minutes after contrast administration of 0.2 mmol/kg gadolinium–DTPA inversion recovery, CMR revealed bright hyperenhancement of the complete pericardium (Panels C and D). No foci of delayed enhancement in the myocardium were noted.

Based on the findings at CMR, acute pericarditis was diagnosed. The patient was treated with non-steroidal anti-inflammatory medication for 4 weeks and remained asymptomatic thereafter.

This case shows that CMR is a valuable non-invasive tool in the differential diagnosis of acute chest pain even in the absence of typical ECG changes. Based on the findings of oedema and contrast enhancement of the pericardium suggesting an acute diffuse inflammatory process of the pericardium, the definite diagnosis of acute pericarditis could be established. Moreover, involvement of the myocardium in the inflammatory process could be excluded.

Panel A. Cardiac magnetic resonance image were obtained with the balanced steady-state free precession technique showing thickened pericardium (5.5 mm, arrows) and a small pericardial effusion.

Panel B. Short-time inversion recovery T2-weighted image show also hyperintense signal from the pericardium (arrows).

Panels C and D. Contrast-enhanced phase-sensitive inversion recovery images [four-chamber view (C) and three-chamber view (D)] in the late phase after gadolinium injection reveal hyperenhancement from the pericardium (arrows).