

## Negative association between left prefrontal GABA concentration and BDNF serum concentration in young adults

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**Background:** The brain's major inhibitory neurotransmitter gamma-aminobutyric acid (GABA) and the brain-derived neurotrophic factor (BDNF) play important roles in several stress-related disorders. Magnetic resonance spectroscopy (MRS) allows for non-invasive quantification of GABA concentration in the brain. We investigated the relationship between GABA concentration in the left dorsolateral prefrontal cortex (DLPFC) and BDNF concentration in the serum in a community-based sample of young subjects.

**Methods:** For the GABA measurement a single voxel MR spectrum was assessed in the prefrontal lobe (25 × 40 × 30 mm) using the MEGA-PRESS method in 276 subjects. BDNF serum concentrations were assessed with an ELISA kit. For 147 subjects we had both MRS and BDNF serum data, and for 79 subjects we had genotype data on the BDNF rs6265 polymorphism. Depressive psychopathology was assessed using Beck's Depression Inventory (BDI), Montgomery-Asberg Depression Rating Scale (MADRS) and Structured Clinical Interviews for Diagnostic and Statistical Manual of Mental Disorders (SCID) for DSM-IV.

**Results:** GABA concentration in the left DLPFC was negatively associated with BDNF serum concentration ( $r = -.264$ ,  $p = .001$ ). This correlation remained significant if corrected for sex ( $r = -.264$ ,  $p = .001$ ). BDNF serum concentration was also positively associated with volumes and surface areas of the left prefrontal cortex ( $p = .048$ ,  $p = .005$ ). There were no significant associations or interaction with depressive psychopathology (BDI, MADRS, SCID) or rs6265.

**Conclusion:** The results of this study suggest that GABA, BDNF and prefrontal brain volumes are interrelated, but do not show a strong association to depressive psychopathology, possibly due to the mild forms of psychiatric conditions present in our community-based sample.

### 1. Introduction

There is growing evidence that the inhibitory neurotransmitter gamma-aminobutyric acid (GABA) and the brain-derived neurotrophic factor (BDNF) play crucial roles in depression, anxiety, epilepsy, and in the treatment of these conditions (Butler et al., 2018; Gabbay et al., 2017; Hasler, van der Veen, Grillon, Drevets and Shen, 2010; Helms et al., 2006; Long et al., 2013; Moreira et al., 2019; Sanacora et al., 2012).

GABA is the primary inhibitory neurotransmitter in the adult brain and shapes the dynamic neural network via GABAergic interneurons (Fukuchi et al., 2014; Schur et al., 2016). Magnetic resonance spectroscopy (MRS) allows for a non-invasive measurement of the GABA

concentration in the brain of living humans. Human and rat studies showed gender-related differences in GABA concentration, with men showing higher GABA concentrations compared to women (Al-Suwailem et al., 2018; Kirkovski et al., 2018; O'Gorman et al., 2011; Pandya et al., 2019).

MRS is an imaging technique to quantify metabolites such as GABA in the brain within one or several voxels of interest (Egerton et al., 2017; Lener et al., 2017; Long et al., 2013). With MRS, signals from total intra- and extracellular GABA as well as GABAergic interneurons across all tissue content are measured (Egerton et al., 2017; Schur et al., 2016). The MRS-visible GABA signal therefore reflects the dynamic balance between GABA synthesis, reuptake and degradation around the synapses (Hasler

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et al., 2010), and has been shown to be sensitive to medication-induced changes (e.g. from antiepileptic drugs like topiramate (Kuzniecky et al., 2002; Reis et al., 2002), which also induce changes in cortical excitability in humans (Dyke et al., 2017).

BDNF is a protein that belongs to the neurotrophin family of growth factors and it is involved in several aspects of neuronal functioning (Naegelin et al., 2018). BDNF can be found in the brain as well as in the blood, secreted by circulating platelets in humans, which release BDNF during blood coagulation. Its concentration can be measured non-invasively in serum (Lommatzsch et al., 2005; Yamamoto and Gurney, 1990). BDNF serum concentrations have been associated with the BDNF concentration within the brain (Karege et al., 2002; Klein et al., 2011). However, the mechanisms explaining this association remain to be fully elucidated (de Azevedo Cardoso et al., 2014; Karege et al., 2002; Naegelin et al., 2018; Vinberg et al., 2013).

The most plausible explanation for the presence of BDNF in the platelets are the platelet-generating megakaryocyte cells. These cells have been found to contain BDNF transcripts and may represent the link between brain and serum BDNF concentrations (Chacon-Fernandez et al., 2016; Naegelin et al., 2018). BDNF serum concentrations were shown to be decreased in untreated depressed participants compared to healthy subjects, and females tend to have lower BDNF serum concentrations compared to men (de Azevedo Cardoso et al., 2014; Elfving et al., 2012; Ellsworth et al., 2013; Kreinin et al., 2015; Lommatzsch et al., 2005; Molendijk et al., 2014; Ozan et al., 2010). Studies also found associations between BDNF and the volume of the prefrontal cortex (Forde et al., 2014; Frodl et al., 2008; Harrisberger et al., 2015; Matsuo et al., 2009; Pezawas et al., 2004), suggesting a link between prefrontal brain structure and function, and BDNF concentration in the serum.

Some rat and human studies have demonstrated interactions between GABA-ergic neurotransmission and BDNF release. On one hand, BDNF can suppress or presynaptically enhance GABA release through the activation of postsynaptic tropomyosin receptor kinase B (TrkB)-type receptors, which may result in an altered total GABA concentration (Bulleit and Hsieh, 2000; Colino-Oliveira et al., 2016; Kim et al., 2017; Pezet et al., 2002; Tanaka et al., 1997; Wardle and Poo, 2003). Conversely, changes in the balance between excitatory glutamatergic and inhibitory GABAergic neurotransmission has been shown to alter BDNF expression (Saffarpour et al., 2017). The activation of glutamate receptors enhances the synthesis of BDNF, whereas stimulation of the GABAergic system decreases BDNF synthesis and thus may alter BDNF concentration in the brain and in the blood (Fukuchi et al., 2014; Mar-migere et al., 2003; Zafra et al., 1991).

Diminished GABA-ergic transmission in the prefrontal cortex has been related to circuit dysfunction in mood- and stress-related disorders including major depressive disorder (MDD) (Germer et al., 1984; Ghosal et al., 2017; Hasler et al., 2010). Given the results from rat and human studies cited above, we hypothesized that a higher left dorsolateral prefrontal cortex (DLPFC) GABA concentration is associated with lower BDNF serum concentration, and that this association differs between genders. To elucidate this putatively inverse relationship between GABA and BDNF in healthy and depressed subjects, we investigated healthy subjects and current and remitted MDD patients in a community-based sample. In particular, we assessed left prefrontal GABA concentrations with single-voxel MRS and BDNF concentration in the serum. Clinical characterization of the research subjects included diagnostic interviews based on DSM-IV and standard rating instruments for depressive symptoms.

## 2. Methods and materials

### 2.1. Recruitment

Participants were recruited by advertisements in local newspapers or by blackboard webpages of the University of Zurich for this ongoing cohort study. This provided us with a sample of young adults from the

general population. After a full explanation of the goals and risks of the study and after giving their consent, participants were admitted to the study. Inclusion criteria were age between 18 and 40 years and preferably being without any psychopharmacological medication for the last three months before testing to minimize the possible effects on the brain. Participants with current or remitted MDD were included into the study, but other severe psychiatric disorders, for example schizophrenia were excluded. The local ethics committee approved the study and the written consent (Kantonale Ethikkommission Zürich). The study described has been carried out in accordance with the Code of Ethics of the World Medical Association (Declaration of Helsinki) for experiments involving humans. In one of our previous studies (Hasler et al., 2019) we report data from a subset of the participants in the present study.

### 2.2. Procedure

After giving consent the participants were screened for magnetic resonance (MR) safety with a standard checklist and for psychological disorders with the screening part of the Structured Clinical Interview for Diagnostic and Statistical Manual of Mental Disorders (SCID) for DSM-IV. Later the participants completed an extensive battery of psychological and physiological questionnaires. Validated German versions of all instruments were used. Finally, an approximately five hour long face-to-face session at the Children's Hospital in Zurich was conducted.

The face-to-face session, during which participants had to complete paper-pencil versions of psychological questionnaires, was supervised by a trained psychologist. Fifty ml blood samples were collected by trained staff from the hospital after the psychological assessments. Subjects underwent an MR session in a 3.0T GE Discovery MR750 scanner.

### 2.3. Sample

276 subjects were assessed with MRS, in 147 we had both MRS and BDNF serum data (67.6% females) and in 79 we also had genotype data on the polymorphism rs6265 of the BDNF gene.

Exclusion criteria were age outside the range of 18–40 years, preferably no use of psychopharmacological drugs in the past three months, severe psychiatric disorders such as schizophrenia, drug or alcohol abuse or any illnesses or accidents affecting the brain or genetic variance, as for example brain injuries or diabetes.

Of the 276 subjects who underwent magnetic resonance imaging (MRI) scanning, two were excluded because they had mild abnormalities that could potentially influence the results (an arachnoid cyst in the right posterior lateral ventricle; a lipoma with dysplasia of the corpus callosum) leaving us with a sample of 274 subjects. Of these 274 subjects, 75 were diagnosed with MDD (16 currently depressed and 59 partially or fully remitted). Out of the reported 147 subjects with both GABA and BDNF serum data, 29 were diagnosed with MDD, including eight currently depressed and 21 partially or fully remitted (Table 1). Three of the MDD cases were on psychotropic medication (antidepressants or antipsychotics). Out of the 147 subjects, 99 were females and 54 of them were taking contraceptive medication.

### 2.4. Blood sampling

Venous blood samples were taken on the day of MR scanning in the Children's Hospital (mean time 2:12 p.m., standard deviation 1.1 h). The blood samples were analysed by specialized laboratories in Basel (Switzerland) and New York (United States).

BDNF serum samples were stored at  $-80^{\circ}\text{C}$  before assaying BDNF content. An enzyme-linked immunosorbent assay (ELISA) kit (Biosensis® Mature BDNF Rapid™ ELISA Kit: Human, Mouse, Rat; Thebarton, SA, Australia) was used to assess the serum BDNF concentrations. Serum samples were appropriately diluted (1:100) and detection of BDNF was carried out on a pre-coated mouse monoclonal anti-mature BDNF 96-well plate as described in the manufacturer's protocol. The absorbance was

**Table 1.** Clinical characteristics of the original sample (N = 274) and the subsample (N = 147) with available data concerning GABA concentration and BDNF serum concentration used in this study.

	GABA sample	GABA-BDNF subsample
N	274	147
Age Mean (SD) [years]	24.68 (4.57)	24.45 (4.35)
Females	179 (65.3%)	99 (67.3%)
MDD current or remitted (N)	75 (27.4%)	29 (19.7%)
Medicated Subjects	3 (1.1%)	2 (1.4%)
Mean BDNF serum concentration (pg/ml)	-	11500 (3400)
BDNF rs6265 genotypes (subsample N = 109/79)	c/c 61 c/t 43 t/t 5	c/c 43 c/t 33 t/t 3
BDI Mean (SD)	4.81 (6.87), 2 missing	3.81 (6.53), 2 missing
MADRS Mean (SD)	3.44 (5.56), 1 missing	2.96 (5.90), 1 missing
Volume left prefrontal cortex (gyri and sulci, cm <sup>3</sup> )	52.84 (6.10), 1 missing	53.18 (6.17)
Surface left prefrontal cortex (gyri and sulci, cm <sup>2</sup> )	165.16 (17.95), 1 missing	165.71 (17.33)

measured within 5 min after addition of the stop solution in a microplate reader set at 450nm and a correction wavelength set to 690nm, to determine BDNF concentrations according to the standard curve. All assays were carried out in duplicate and means were calculated.

### 2.5. MRI data acquisition and analyses

MRI data were obtained with a 3.0 GE Discovery MR750 scanner equipped with an eight-channel receive-only head coil. The measurements included a localizer and a 3D T1-weighted spoiled gradient recalled (SPGR) sequence for the assessment of global and local brain volumes (162 slices of 256 x 256 voxels, 1 x 1 x 1 mm resolution; TR = 11 ms, TE = 5 ms, TI = 600 ms, flip angle 8°), which was used for the localization of the MR spectra.

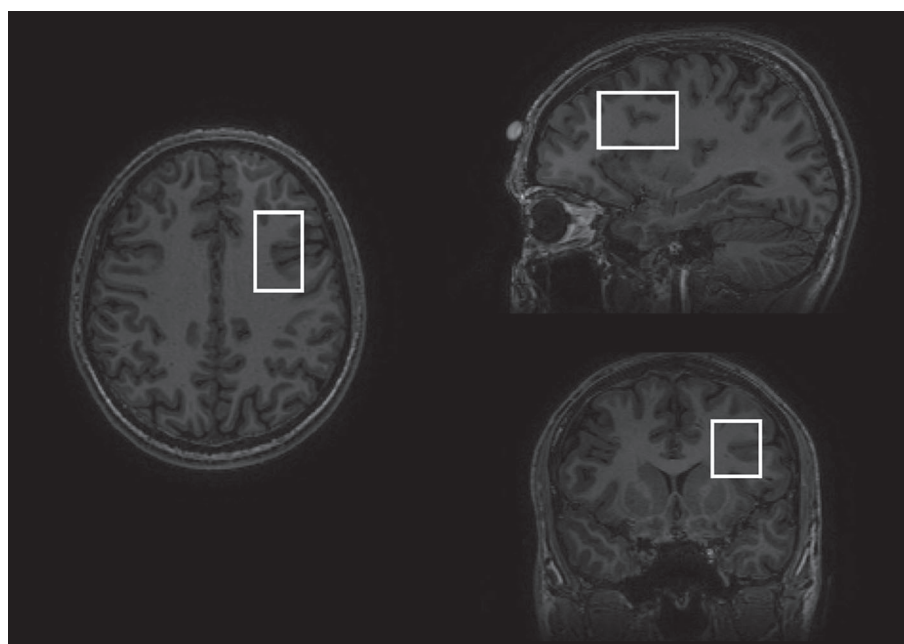
T1-weighted images were segmented and warped into MNI space using “new segment” procedure in SPM12 for Matlab. For volumetric measurements, freesurfer (version 5.3.0) was run over the 3-dimensional T1-weighted images (Dale et al., 1999; Fischl et al., 1999). Volumes of subcortical structures, whole brain volumes and areas and thicknesses of neocortical areas can be analysed by this software.

MR spectra were acquired from a 25 x 40 x 30 mm voxel in the left frontal lobe using the MEGA-PRESS method (TE = 69 ms, TR = 2000 ms, 320 averages and eight-step phase cycle; Figure 1). We chose the left

DLPFC (and not the right one) because most MRS studies examined the left side (Moriguchi et al., 2018), and because of the default direction of the chemical shift effect on the scanner, which by results in fat being shifted to the left (so for a voxel on the left side of the brain, scalp fat is shifted away from the voxel), and water being shifted to the right (so for a voxel on the left side of the brain, ventricular water is also shifted away from the voxel). Due to the major overlapping signals for GABA with those from N-acetyl aspartate (NAA) and Creatine, the MEGA-PRESS technique was used to edit the GABA signal (Schur et al., 2016). Due to the overlap of the edited GABA peak with co-edited macromolecules, findings represent GABA<sup>+</sup> rather than pure GABA values.

MR spectra were preprocessed with a vendor-provided pipeline, which included steps for coil combination, frequency alignment, phasing, residual water removal, and averaging, and both the edited data and water reference data were saved as RAW files for processing with LCModel. Spectral fitting for the edited data was performed with LCModel version 6.3 using a simulated basis set, including basis spectra for GABA, Glutamate, Glutamine, Glutathione, N-acetyl-aspartate (NAA), and N-acetyl-aspartyl-glutamate (NAAG). The control parameter sptype was set to ‘mega-press-2’ to control the baseline fitting, and spectra were visually inspected for the quality of the GABA fit.

Metabolite levels with a Cramer-Rao lower bound (CRLB) of more than 20% were excluded from the analysis. Water-scaled GABA+



**Figure 1.** MR images showing the position of the measured voxel in the left DLPC.

concentrations were calculated in institutional units after correcting the GABA+ and estimated water concentrations for cerebrospinal fluid contamination (atrophy) within the voxel (Chowdhury et al., 2015). Figure 2 shows the edited and unedited spectra for all participants, together with the group mean spectra and the mean LCModel fit across the group.

## 2.6. Psychological measurements

Subjects filled out the Becks Depression Inventory (BDI) (Beck et al., 1961) and the Montgomery-Asberg Depression Rating Scale (MADRS) (Montgomery and Asberg, 1979) shortly before MR scanning at the Children's Hospital in Zurich, using the validated German versions. Psychiatric diagnoses were made based on the SCID for DSM-IV (First et al., 2001).

## 2.7. Statistics

In explorative analyses, we tested associations between GABA+ concentration, biological measures (BDNF serum concentrations and left DLPFC volumes) and depressive psychopathology (BDI, MADRS, MDD DSM-IV diagnosis) using Pearson's bivariate correlation analyses in SPSS (IBM SPSS Statistics, Version 24).

Due to previously reported gender differences concerning both GABA and BDNF serum concentrations we repeated the analyses in the female and male subsample. MDD was added to the analyses as confounding parameter in a partial correlation. Post-hoc analyses were also conducted to examine the link between BDNF and other neuro-metabolites present in the edit OFF spectra, namely, NAA, Creatine, Choline, and myo-inositol.

## 3. Results

Figure 3 shows that GABA+ concentration in the DLPFC was significantly negatively associated with the BDNF serum concentration ( $r = -.264, p = .001, N = 147$ ; Figure 3). This negative association was significant in the female subsample ( $r = -.306, p = .002, N = 99$ ) but not in the male subsample ( $r = -.189, p = .197, N = 48$ ).

BDNF serum concentrations were positively associated with the volumes of the left prefrontal cortex (gyri and sulci;  $r = .164, p = .048, N = 147$ ), and the surface area of the left prefrontal cortex (gyri and sulci;  $r = .231, p = .005, N = 147$ ). GABA+ concentration showed no significant association with the volume or the surface of the left prefrontal cortex (volume:  $r = -.128, p = .122$ ; surface:  $r = -.089, p = .282, N = 147$ ).

GABA+ concentration in the left DLPFC was not significantly associated with the diagnosis of MDD ( $r = -.092, p = .268, N = 147$ ), BDI sum score ( $r = -.043, p = .604, N = 145$ ) and MADRS total score ( $r = -.081, p = .333, N = 146, 1$  missing). There was no significant association between BDNF concentration in the serum and MDD diagnosis ( $r = .105, p = .203, N = 147$ ), BDI sum score ( $r = -.031, p = .709, N = 145$ ) and MADRS total score ( $r = .099, p = .236, N = 146$ ). BDNF concentration in the serum was not significantly associated with hippocampal volumes (left hippocampus volume:  $r = -.058, p = .488$ ; right hippocampus volume:  $r = .041, p = .618, N = 147$ ). In  $N = 79$  subjects we had genotype data on the BDNF polymorphism rs6265. We did not find a significant association between this polymorphism and bilateral hippocampal brain volumes (left hippocampus volume:  $r = .159, p = .161$ ; right hippocampus volume:  $r = .170, p = .135, N = 79$ ), BDNF serum concentration ( $r = .059, p = .608, N = 79$ ) or prefrontal GABA+ concentration ( $r = -.038, p = .742, N = 79$ ). This result remained unchanged after excluding the three medicated subjects (data not shown). Using age, BMI or sports activity as a control variable did not change the results either.

The bivariate correlations were also calculated for genders separately, due to reported sex-differences in GABA and BDNF systems. There was a trend for an association between left DLPFC GABA+ concentration and MADRS total score in male subjects ( $r = -.267, p = .067, N = 48$ ) but not in female subjects ( $r = -.010, p = .922, N = 98$ ). The partial correlation showed a significant association between prefrontal GABA+ and BDNF serum concentration if corrected for sex ( $r = -.264, p = .001$ ).

The partial correlation revealed that MDD did not change the significant results between GABA+ and BDNF serum concentration ( $r = -.257, p = .002, N = 144$ ), between BDNF concentration in the serum and left prefrontal volume and surface-area (volume:  $r = .197, p = .017$ ; surface-area:  $r = .266, p = .001, N = 144$ ). The use of contraceptives was not associated with the BDNF polymorphism ( $r = .032, p = .780, N = 79$ ), BDNF serum concentration ( $r = .097, p = .241, N = 147$ ) and prefrontal GABA+ concentration ( $r = -.053, p = .525, N = 147$ ). The post-hoc tests for a link between BDNF and other neuro-metabolites present in the edit OFF spectra did not reveal any significant associations (all  $p > 0.12$ ). Using Creatinine instead of H2O as reference signal did not change the main findings of this study.

## 4. Discussion

In a cohort of young adults from the general population, we investigated the link between GABA+ levels in the left DLPFC, prefrontal brain volumes and surface-areas, serum BDNF concentrations, and depressive psychopathology.

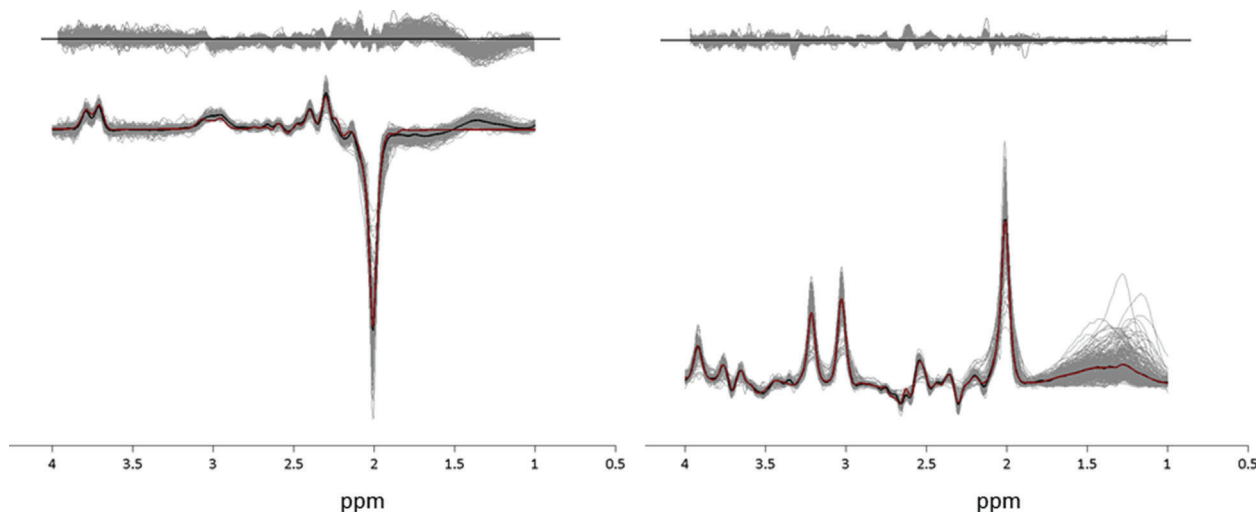
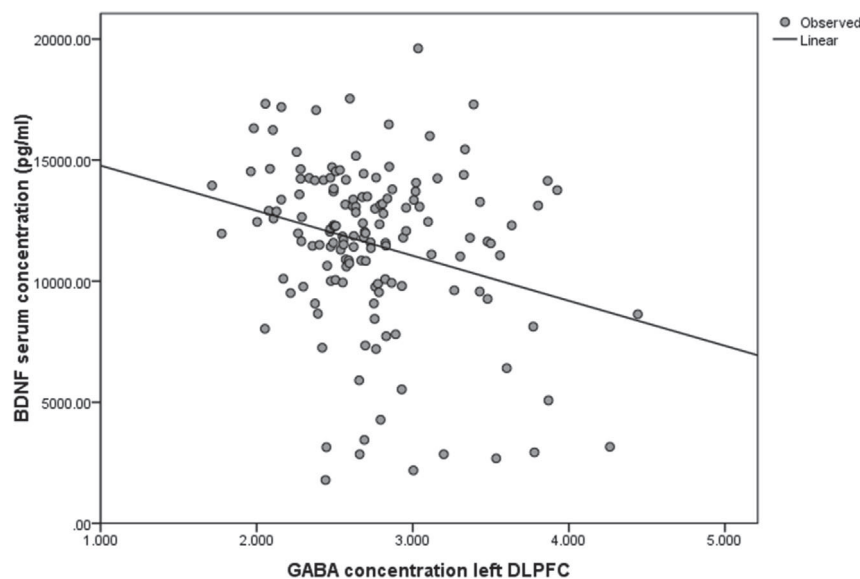


Figure 2. Edited and unedited spectra for all participants. The individual spectra for all participants are shown in grey, with the group-mean spectra overlaid in black and the mean LCModel fit overlaid in red. The residuals from each participant are also plotted in grey above the spectra.





**Figure 3.** Bivariate correlation between GABA concentration in the left prefrontal cortex, corrected for atrophy and water scaling, and BDNF serum concentration, normalized and averaged ( $r = -.264$ ,  $p = .001$ ,  $N = 147$ ).

The main finding of the present study is that higher GABA+ concentrations in the left DLPFC were associated with lower BDNF serum concentrations in young adults. No association was found between the BDNF polymorphism and GABA+ concentration. In the gender-specific analyses, the significant association between GABA+ and BDNF serum concentration held true in the female but not in the male subsample. BDNF serum concentration was associated with increased brain volumes and surface-areas in the left prefrontal cortex. No associations were found between GABA+ concentration, BDNF polymorphism and left prefrontal volumes and surface-areas. Depressive symptoms as measured by BDI and MADRS and MDD diagnosis were not associated with GABA+, BDNF polymorphism or BDNF serum concentrations, but we found a negative trend towards a correlation between GABA+ concentration and MADRS total score in males but not in females. Depression did not affect the association between GABA and BDNF.

Previous studies investigating the relationship between GABA and BDNF have suggested a complex and not yet fully understood interplay between the two factors, as discussed in the introduction (Bulleit and Hsieh, 2000; Colino-Oliveira et al., 2016; Marmigere et al., 2003; Pezet et al., 2002; Saffarpour et al., 2017; Zafra et al., 1991). The connection between GABA and BDNF may depend on pre- and postsynaptic effects of the two factors and may differ considerably between brain regions (Marmigere et al., 2003; Saffarpour et al., 2017; Tanaka et al., 1997; Zafra et al., 1991). Both GABA and BDNF serum concentrations have been shown to be reduced in subjects with MDD and subjects after acute and chronic stress (Fogaca and Duman, 2019; Hasler et al., 2010; Kishi et al., 2017). However, in remitted subjects with MDD, prefrontal GABA concentration was found to be normal (Hasler et al., 2005; Schur et al., 2016). Our findings of normal BDNF and GABA concentrations in MDD may therefore arise as a result of our community-based sampling method, leading to the inclusion of subjects with rather mild, partly remitted and unmedicated MDD.

Previous studies on serum BDNF concentrations have found gender differences, with men showing higher BDNF serum concentrations compared to women (de Azevedo Cardoso et al., 2014; Ozan et al., 2010). Our study suggests that gender differences in BDNF concentrations are unrelated to depression-related disorders, and could possibly be related to the effects of sex hormones (Elfving et al., 2012).

To date, our knowledge about sex-related differences concerning the GABAergic system is limited. Some studies have shown higher GABA concentrations in men compared to women (O'Gorman et al., 2011;

Pandya et al., 2019). Our significant correlation between GABA+ and BDNF serum concentration in the female but not in the male subsample may reflect both GABA and BDNF specific gender differences. The negative trend in our results between GABA+ concentrations and MADRS scores in the male but not in the female subsample may also reflect the gender differences concerning GABA but should be interpreted with caution. Although many previous studies investigated GABA and BDNF serum concentrations in subjects with psychiatric disorders, the apparent gender differences in both GABA+ and BDNF seen in our study of young adults from the general population indicates that gender should also be considered in group comparisons of healthy samples with psychiatric groups (Molendijk et al., 2014). In addition, since BDNF concentrations can be normalized by antidepressant treatment (Kreinin et al., 2015), medication status should be taken into account to avoid potential confounds from medication effects.

BDNF is strongly expressed in the limbic brain regions, hence many studies have tested for associations between hippocampal volumes and BDNF serum concentrations, yielding inconsistent results (Frodl et al., 2014; Harrisberger et al., 2015). In our study, we did not find a significant correlation between BDNF concentration in the serum or the BDNF polymorphism rs6265 and hippocampal brain volumes.

With regard to the BDNF polymorphism, we did not find a significant association with BDNF concentration in the serum. The BDNF gene is located on chromosome 11p13 and encodes proBDNF, which generates mature BDNF, which can be measured in plasma and serum (Elfving et al., 2012). Most studies have researched the single nucleotide polymorphism (SNP) rs6265 (Val66Met) of the BDNF gene, but the association between this polymorphism and BDNF serum concentrations show divergent results and the mechanism has yet to be demonstrated in detail (Karege et al., 2002; Kishi et al., 2017; Naegelin et al., 2018; Ozan et al., 2010). The Met allele of the rs6265 polymorphism is thought to impair the intracellular trafficking of proBDNF into the dendrites and vesicles, thus this SNP seems to play an important role in the regulation of extracellular BDNF concentrations (Kishi et al., 2017). A series of studies have examined the SNP rs6265 regarding the risk of depression and hippocampal volumes (Elfving et al., 2012). In healthy controls, homozygous Val/Val carriers of the rs6265 polymorphism showed larger hippocampal volumes compared to Met carriers (Hajek et al., 2012; Kishi et al., 2017). However, Harrisberger et al. found no such association between the rs6265 polymorphism and hippocampal volume in patients with MDD (Harrisberger et al., 2015). Forde et al. showed that the BDNF

Val66Met polymorphism may not be as specific as previously supposed, but may have a global effect on the cortical morphology in humans (Forde et al., 2014). Matsuo et al. presented smaller DLPFC volumes in Val/Met carriers as compared to the Val/Val subjects (Matsuo et al., 2009). Our results of increased BDNF serum concentration being associated with larger volumes and surface-areas in the prefrontal cortex in young adults may reflect the association between BDNF polymorphism and prefrontal brain volume, but still it remains unclear whether and how BDNF serum concentration is linked to BDNF in the brain (Naegelin et al., 2018).

Recently it has been suggested that, aside from platelet-generating megakaryocytes, epigenetics may play an important role in the association between BDNF serum and brain BDNF concentrations (Chacon-Fernandez et al., 2016; Chen and Chen, 2017; Mallei et al., 2015; Naegelin et al., 2018). Mallei et al. showed that homozygous mice Met carriers displayed repressive neuronal activation of BDNF and repressive epigenetic changes in certain BDNF promoters, which is accompanied by reduced transcription of BDNF promoters (Mallei et al., 2015).

Our non-significant results regarding serum BDNF concentration and depression-related symptoms may indicate that the downregulation of BDNF serum concentration may not be associated with healthy or mostly remitted MDD participants, but may be specific to severe MDD and reflect the complex molecular pathologies of psychiatric disorders (Guilloux et al., 2012).

The present study provides insight into the association of prefrontal GABA concentration, BDNF serum concentration, prefrontal brain volumes, and depressive symptoms in a cohort of young adults from the general population. The approach of a community-based sample of young subjects should be considered a strength of the present study, as the comparability to the general population is higher than in studies that included severely depressed participants under psychopharmacological drugs. To our knowledge, the association between prefrontal GABA and BDNF serum concentration has not been examined in humans, although animal studies have suggested an important connection between central GABA concentration and BDNF. Furthermore, our study was large enough to study the sexes separately. We sought to keep the time of the blood and MRS measurements constant, because BDNF shows a strong circadian variation (Begliuomini et al., 2008).

The following limitation of our study merits comment. MRS total GABA signal was measured within a relatively large voxel. Our method does not allow for discrimination between GABA concentrations of different cell types. The spectral overlap between GABA and other metabolites remain a challenging issue with MRS. We used the MEGA-PRESS technique to address this limitation, but it is important to note that some macromolecular contribution is likely to be present in the GABA signal detected by MEGA-PRESS, and this signal should therefore be regarded as GABA<sup>+</sup> rather than as pure GABA.

In the present study, two specific aspects of BDNF, namely its concentration in the serum and the rs6265 polymorphisms were investigated. The relationship between brain and serum BDNF has still to be elucidated and thus the results should be considered with caution. Although contraceptive medication does not seem to impact our results, we cannot rule out the possibility that differences between females and males are moderated by sex-hormones. As our sample consisted of young subjects from the general population with low depression scores, our results may differ from samples with current MDD subjects.

In conclusion, our data suggest a complex but important association between GABA<sup>+</sup> concentration and BDNF serum concentration, prefrontal brain volumes and possible gender related differences in young adults. To understand the mechanisms concerning the interaction between BDNF and GABA concentration better, it would be interesting to combine the factors investigated in this study with stress-related epigenetics in a bigger sample. We are planning to conduct follow-up assessments of our cohort to evaluate the predictive value of our measures. For the gender differences concerning GABA and BDNF concentration future studies should focus on gender-matching between groups and should

consider the menstrual cycle, contraceptive use and sex-hormones of female participants. In addition, in future studies it would be interesting to examine the effects of neurotrophic factors and epigenetic markers and their influence on depression and stress-related disorders.

## Declarations

### Author contribution statement

Sabrina Theresia Müller, Christopher Ritter: Performed the experiments; Analyzed and interpreted the data; Wrote the paper.

Andreas Buchmann, Carmen Ghisleni: Performed the experiments; Wrote the paper.

Melanie Haynes, Gregor Hasler: Conceived and designed the experiments; Wrote the paper.

Ruth Tuura: Conceived and designed the experiments; Contributed reagents, materials, analysis tools or data; Wrote the paper.

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### Competing interest statement

The authors declare no conflict of interest.

### Additional information

No additional information is available for this paper.

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## References

- Al-Suwailem, E., Abdi, S., El-Ansary, A., 2018. Sex differences in the glutamate signaling pathway in juvenile rats. *J. Neurosci. Res.* 96 (3), 459–466.
- Beck, A.T., Ward, C.H., Mendelson, M., Mock, J., Erbaugh, J., 1961. An inventory for measuring depression. *Arch. Gen. Psychiatr.* 4, 561–571.
- Begliuomini, S., Lenzi, E., Ninni, F., Casarosa, E., Merlini, S., Pluchino, N., Genazzani, A.R., 2008. Plasma brain-derived neurotrophic factor daily variations in men: correlation with cortisol circadian rhythm. *J. Endocrinol.* 197 (2), 429–435.
- Bulleit, R.F., Hsieh, T., 2000. MEK inhibitors block BDNF-dependent and -independent expression of GABA(A) receptor subunit mRNAs in cultured mouse cerebellar granule neurons. *Brain Res. Dev. Brain Res.* 119 (1), 1–10.
- Butler, K.M., Moody, O.A., Schuler, E., Coryell, J., Alexander, J.J., Jenkins, A., Escayg, A., 2018. De novo variants in GABRA2 and GABRA5 alter receptor function and contribute to early-onset epilepsy. *Brain* 141 (8), 2392–2405.
- Chacon-Fernandez, P., Sauberli, K., Colzani, M., Moreau, T., Ghevaert, C., Barde, Y.A., 2016. Brain-derived neurotrophic factor in megakaryocytes. *J. Biol. Chem.* 291 (19), 9872–9881.
- Chen, K.W., Chen, L., 2017. Epigenetic regulation of BDNF gene during development and diseases. *Int. J. Mol. Sci.* 18 (3).
- Chowdhury, F.A., O'Gorman, R.L., Nashef, L., Elwes, R.D., Edden, R.A., Murdoch, J.B., Richardson, M.P., 2015. Investigation of glutamine and GABA levels in patients with idiopathic generalized epilepsy using MEGAPRESS. *J. Magn. Reson. Imag.* 41 (3), 694–699.
- Colino-Oliveira, M., Rombo, D.M., Dias, R.B., Ribeiro, J.A., Sebastiao, A.M., 2016. BDNF-induced presynaptic facilitation of GABAergic transmission in the hippocampus of young adults is dependent of TrkB and adenosine A2A receptors. *Purinergic Signal.* 12 (2), 283–294.
- Dale, A.M., Fischl, B., Sereno, M.I., 1999. Cortical surface-based analysis. I. Segmentation and surface reconstruction. *Neuroimage* 9 (2), 179–194.
- de Azevedo Cardoso, T., Mondin, T.C., Wiener, C.D., Marques, M.B., Fucolo Bde, A., Pinheiro, R.T., Oses, J.P., 2014. Neurotrophic factors, clinical features and gender differences in depression. *Neurochem. Res.* 39 (8), 1571–1578.

- Dyke, K., Pepes, S.E., Chen, C., Kim, S., Sigurdsson, H.P., Draper, A., Jackson, S.R., 2017. Comparing GABA-dependent physiological measures of inhibition with proton magnetic resonance spectroscopy measurement of GABA using ultra-high-field MRI. *Neuroimage* 152, 360–370.
- Egerton, A., Modinos, G., Ferrera, D., McGuire, P., 2017. Neuroimaging studies of GABA in schizophrenia: a systematic review with meta-analysis. *Transl. Psychiatry* 7 (6), e1147.
- Elfving, B., Buttenschon, H.N., Foldager, L., Poulsen, P.H., Andersen, J.H., Grynderup, M.B., Mors, O., 2012. Depression, the Val66Met polymorphism, age, and gender influence the serum BDNF level. *J. Psychiatr. Res.* 46 (9), 1118–1125.
- Ellsworth, K.A., Moon, I., Eckloff, B.W., Fridley, B.L., Jenkins, G.D., Batzler, A., Wang, L., 2013. FKBP5 genetic variation: association with selective serotonin reuptake inhibitor treatment outcomes in major depressive disorder. *Pharmacogenetics Genom.* 23 (3), 156–166.
- First, M.B., Spitzer, R.L., Gibbon, M., Williams, J.B.W., 2001. Structured Clinical Interview for DSM-IV-TR Axis I Disorders, Research Version, Patient Edition (SCID-I/P). Biometrics Research, New York State Psychiatric Institute, New York.
- Fischl, B., Sereno, M.I., Dale, A.M., 1999. Cortical surface-based analysis. II: inflation, flattening, and a surface-based coordinate system. *Neuroimage* 9 (2), 195–207.
- Fogaca, M.V., Duman, R.S., 2019. Cortical GABAergic dysfunction in stress and depression: new insights for therapeutic interventions. *Front. Cell. Neurosci.* 13, 87.
- Forde, N.J., Ronan, L., Suckling, J., Scanlon, C., Neary, S., Holleran, L., Cannon, D.M., 2014. Structural neuroimaging correlates of allelic variation of the BDNF val66met polymorphism. *Neuroimage* 90, 280–289.
- Frodl, T., Carballedo, A., Frey, E.M., O'Keane, V., Skokauskas, N., Morris, D., Connor, T., 2014. Expression of glucocorticoid inducible genes is associated with reductions in cornu ammonis and dentate gyrus volumes in patients with major depressive disorder. *Dev. Psychopathol.* 26 (4 Pt 2), 1209–1217.
- Frodl, T., Moller, H.J., Meisenzahl, E., 2008. Neuroimaging genetics: new perspectives in research on major depression? *Acta Psychiatr. Scand.* 118 (5), 363–372.
- Fukuchi, M., Kirikoshi, Y., Mori, A., Eda, R., Ihara, D., Takasaki, I., Tsuda, M., 2014. Excitatory GABA induces BDNF transcription via CRTCL and phosphorylated CREB-related pathways in immature cortical cells. *J. Neurochem.* 131 (2), 134–146.
- Gabbay, V., Bradley, K.A., Mao, X., Ostrover, R., Kang, G., Shungu, D.C., 2017. Anterior cingulate cortex gamma-aminobutyric acid deficits in youth with depression. *Transl. Psychiatry* 7 (8), e1216.
- Gerner, R.H., Fairbanks, L., Anderson, G.M., Young, J.G., Scheinin, M., Linnoila, M., Cohen, D.J., 1984. CSF neurochemistry in depressed, manic, and schizophrenic patients compared with that of normal controls. *Am. J. Psychiatr.* 141 (12), 1533–1540.
- Ghosal, S., Hare, B., Duman, R.S., 2017. Prefrontal cortex GABAergic deficits and circuit dysfunction in the pathophysiology and treatment of chronic stress and depression. *Curr. Opin. Behav. Sci.* 14, 1–8.
- Guilloux, J.P., Douillard-Guilloux, G., Kota, R., Wang, X., Gardier, A.M., Martinowich, K., Sibille, E., 2012. Molecular evidence for BDNF- and GABA-related dysfunctions in the amygdala of female subjects with major depression. *Mol. Psychiatr.* 17 (11), 1130–1142.
- Hajek, T., Kopecek, M., Hoschl, C., 2012. Reduced hippocampal volumes in healthy carriers of brain-derived neurotrophic factor Val66Met polymorphism: meta-analysis. *World J. Biol. Psychiatr.* 13 (3), 178–187.
- Harrisberger, F., Smieskova, R., Schmidt, A., Lenz, C., Walter, A., Wittfeld, K., Borgwardt, S., 2015. BDNF Val66Met polymorphism and hippocampal volume in neuropsychiatric disorders: a systematic review and meta-analysis. *Neurosci. Biobehav. Rev.* 55, 107–118.
- Hasler, G., Buchmann, A., Haynes, M., Müller, S., Ghisleni, C., Brechbühl, S., Tuura, R., 2019. Association between prefrontal glutamine levels and neuroticism determined using proton magnetic resonance spectroscopy. *Transl. Psychiatry* in press.
- Hasler, G., Neumeister, A., van der Veen, J.W., Tuminis, T., Bain, E.E., Shen, J., Charney, D.S., 2005. Normal prefrontal gamma-aminobutyric acid levels in remitted depressed subjects determined by proton magnetic resonance spectroscopy. *Biol. Psychiatr.* 58 (12), 969–973.
- Hasler, G., van der Veen, J.W., Grillon, C., Drevets, W.C., Shen, J., 2010. Effect of acute psychological stress on prefrontal GABA concentration determined by proton magnetic resonance spectroscopy. *Am. J. Psychiatr.* 167 (10), 1226–1231.
- Helms, G., Ciumas, C., Kyaga, S., Savic, L., 2006. Increased thalamus levels of glutamate and glutamine (Glx) in patients with idiopathic generalised epilepsy. *J. Neurol. Neurosurg. Psychiatr.* 77 (4), 489–494.
- Karege, F., Perret, G., Bondolfi, G., Schwald, M., Bertschy, G., Aubry, J.M., 2002. Decreased serum brain-derived neurotrophic factor levels in major depressed patients. *Psychiatr. Res.* 109 (2), 143–148.
- Kim, J., Lee, S., Kang, S., Kim, S.H., Kim, J.C., Yang, M., Moon, C., 2017. Brain-derived neurotrophic factor and GABAergic transmission in neurodegeneration and neuroregeneration. *Neural Regen. Res.* 12 (10), 1733–1741.
- Kirkovski, M., Suo, C., Enticott, P.G., Yucel, M., Fitzgerald, P.B., 2018. Short communication: sex-linked differences in gamma-aminobutyric acid (GABA) are related to social functioning in autism spectrum disorder. *Psychiatry Res. Neuroimaging.* 274, 19–22.
- Kishi, T., Yoshimura, R., Ikuta, T., Iwata, N., 2017. Brain-derived neurotrophic factor and major depressive disorder: evidence from meta-analyses. *Front. Psychiatr.* 8, 308.
- Klein, A.B., Williamson, R., Santini, M.A., Clemmensen, C., Etrrup, A., Rios, M., Aznar, S., 2011. Blood BDNF concentrations reflect brain-tissue BDNF levels across species. *Int. J. Neuropsychopharmacol.* 14 (3), 347–353.
- Kreinin, A., Lissou, S., Neshet, E., Schneider, J., Bergman, J., Farhat, K., Pinhasov, A., 2015. Blood BDNF level is gender specific in severe depression. *PLoS One* 10 (5), e0127643.
- Kuzniecky, R., Ho, S., Pan, J., Martin, R., Gilliam, F., Faught, E., Hetherington, H., 2002. Modulation of cerebral GABA by topiramate, lamotrigine, and gabapentin in healthy adults. *Neurology* 58 (3), 368–372.
- Lener, M.S., Niciu, M.J., Ballard, E.D., Park, M., Park, L.T., Nugent, A.C., Zarate Jr., C.A., 2017. Glutamate and gamma-aminobutyric acid systems in the pathophysiology of major depression and antidepressant response to ketamine. *Biol. Psychiatr.* 81 (10), 886–897.
- Lommatzsch, M., Zingler, D., Schuhbaeck, K., Schloetcke, K., Zingler, C., Schuff-Werner, P., Virchow, J.C., 2005. The impact of age, weight and gender on BDNF levels in human platelets and plasma. *Neurobiol. Aging* 26 (1), 115–123.
- Long, Z., Medlock, C., Dziedzic, M., Shin, Y.W., Goddard, A.W., Dydak, U., 2013. Decreased GABA levels in anterior cingulate cortex/medial prefrontal cortex in panic disorder. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 44, 131–135.
- Mallei, A., Baj, G., Ieraci, A., Corna, S., Musazzi, L., Lee, F.S., Popoli, M., 2015. Expression and dendritic trafficking of BDNF-6 splice variant are impaired in knock-in mice carrying human BDNF Val66Met polymorphism. *Int. J. Neuropsychopharmacol.* 18 (12).
- Marmigier, F., Rage, F., Tapia-Arancibia, L., 2003. GABA-glutamate interaction in the control of BDNF expression in hypothalamic neurons. *Neurochem. Int.* 42 (4), 353–358.
- Matsuo, K., Walss-Bass, C., Nery, F.G., Nicoletti, M.A., Hatch, J.P., Frey, B.N., Soares, J.C., 2009. Neuronal correlates of brain-derived neurotrophic factor Val66Met polymorphism and morphometric abnormalities in bipolar disorder. *Neuropsychopharmacology* 34 (8), 1904–1913.
- Molendijk, M.L., Spinhoven, P., Polak, M., Bus, B.A., Penninx, B.W., Elzinga, B.M., 2014. Serum BDNF concentrations as peripheral manifestations of depression: evidence from a systematic review and meta-analyses on 179 associations (N=9484). *Mol. Psychiatr.* 19 (7), 791–800.
- Montgomery, S.A., Asberg, M., 1979. A new depression scale designed to be sensitive to change. *Br. J. Psychiatry* 134, 382–389.
- Moreira, F.P., de Azevedo Cardoso, T., Mondin, T.C., Wiener, C.D., de Mattos Souza, L.D., Oses, J.P., da Silva, R.A., 2019. Serum level of nerve growth factor is a potential biomarker of conversion to bipolar disorder in women with major depressive disorder. *Psychiatr. Clin. Neurosci.*
- Moriguchi, S., Takamiya, A., Noda, Y., Horita, N., Wada, M., Tsugawa, S., Nakajima, S., 2018. Glutamatergic neurometabolite levels in major depressive disorder: a systematic review and meta-analysis of proton magnetic resonance spectroscopy studies. *Mol. Psychiatr.*
- Naegelin, Y., Dingsdale, H., Sauberli, K., Schadelin, S., Kappos, L., Barde, Y.A., 2018. Measuring and validating the levels of brain-derived neurotrophic factor in human serum. *eNeuro* 5 (2).
- O'Gorman, R.L., Michels, L., Edden, R.A., Murdoch, J.B., Martin, E., 2011. In vivo detection of GABA and glutamate with MEGA-PRESS: reproducibility and gender effects. *J. Magn. Reson. Imag.* 33 (5), 1262–1267.
- Ozan, E., Okur, H., Eker, C., Eker, O.D., Gonul, A.S., Akarsu, N., 2010. The effect of depression, BDNF gene val66met polymorphism and gender on serum BDNF levels. *Brain Res. Bull.* 81 (1), 61–65.
- Pandya, M., Palpagama, T.H., Turner, C., Waldvogel, H.J., Faull, R.L., Kwakowsky, A., 2019. Sex- and age-related changes in GABA signaling components in the human cortex. *Biol. Sex Differ.* 10 (1), 5.
- Pezawas, L., Verchinski, B.A., Mattay, V.S., Callicott, J.H., Kolachana, B.S., Straub, R.E., Weinberger, D.R., 2004. The brain-derived neurotrophic factor val66met polymorphism and variation in human cortical morphology. *J. Neurosci.* 24 (45), 10099–10102.
- Pezet, S., Cunningham, J., Patel, J., Grist, J., Gavazzi, I., Lever, I.J., Malcangio, M., 2002. BDNF modulates sensory neuron synaptic activity by a facilitation of GABA transmission in the dorsal horn. *Mol. Cell. Neurosci.* 21 (1), 51–62.
- Reis, J., Tergau, F., Hamer, H.M., Müller, H.H., Knake, S., Fritsch, B., Rosenow, F., 2002. Topiramate selectively decreases intracortical excitability in human motor cortex. *Epilepsia* 43 (10), 1149–1156.
- Saffarpour, S., Shaabani, M., Naghd, N., Farahmandfar, M., Janzadeh, A., Nasirinezhad, F., 2017. In vivo evaluation of the hippocampal glutamate, GABA and the BDNF levels associated with spatial memory performance in a rodent model of neuropathic pain. *Physiol. Behav.* 175, 97–103.
- Sanacora, G., Treccani, G., Popoli, M., 2012. Towards a glutamate hypothesis of depression: an emerging frontier of neuropsychopharmacology for mood disorders. *Neuropharmacology* 62 (1), 63–77.
- Schur, R.R., Draisma, L.W., Wijnen, J.P., Boks, M.P., Koevoets, M.G., Joels, M., Vinkers, C.H., 2016. Brain GABA levels across psychiatric disorders: a systematic literature review and meta-analysis of (1) H-MRS studies. *Hum. Brain Mapp.* 37 (9), 3337–3352.
- Tanaka, T., Saito, H., Matsuki, N., 1997. Inhibition of GABA synaptic responses by brain-derived neurotrophic factor (BDNF) in rat hippocampus. *J. Neurosci.* 17 (9), 2959–2966.
- Vinberg, M., Bukh, J.D., Bennike, B., Kessing, L.V., 2013. Are variations in whole blood BDNF level associated with the BDNF Val66Met polymorphism in patients with first episode depression? *Psychiatr. Res.* 210 (1), 102–108.
- Wardle, R.A., Poo, M.M., 2003. Brain-derived neurotrophic factor modulation of GABAergic synapses by postsynaptic regulation of chloride transport. *J. Neurosci.* 23 (25), 8722–8732.
- Yamamoto, H., Gurney, M.E., 1990. Human platelets contain brain-derived neurotrophic factor. *J. Neurosci.* 10 (11), 3469–3478.
- Zafra, F., Castren, E., Thoenen, H., Lindholm, D., 1991. Interplay between glutamate and gamma-aminobutyric acid transmitter systems in the physiological regulation of brain-derived neurotrophic factor and nerve growth factor synthesis in hippocampal neurons. *Proc. Natl. Acad. Sci. U. S. A.* 88 (22), 10037–10041.