

Searching for targets for the systemic therapy of mesothelioma

R. A. Stahel^{1*}, W. Weder², E. Felley-Bosco¹, U. Petrausch¹, A. Curioni-Fontecedro¹, I. Schmitt-Opitz² & S. Peters³

Departments of ¹Oncology; ²Thoracic Surgery, University Hospital, Zürich; ³Department of Oncology, Centre Hospitalier Universitaire Vaudois, Lausanne, Switzerland

Received 25 August 2014; revised 15 January 2015 and 8 February 2015; accepted 12 February 2015

Malignant mesothelioma is an incurable disease associated with asbestos exposure arising in the pleural cavity and less frequently in the peritoneal cavity. Platinum-based combination chemotherapy with pemetrexed is the established standard of care. Multimodality approaches including surgery and radiotherapy are being investigated. Increasing knowledge about the molecular characteristics of mesothelioma had led to the identification of novel potential targets for systemic therapy. Current evidence suggests pathways activated in response to merlin deficiency, including Pi3K/mTOR and the focal adhesion kinase, as well as immunotherapeutic approaches to be most promising. This review elaborates on the rationale behind targeted approaches that have been and are undergoing exploration in mesothelioma and summarizes available clinical results and ongoing efforts to improve the systemic therapy of mesothelioma.

Key words: mesothelioma, targeted therapy, immunotherapy

Introduction

Mesothelioma is a fatal disease predominantly arising in the pleural cavity and less so in the peritoneal cavity. The association of mesothelioma with asbestos exposure is well established. The time from exposure to the diagnosis is on the average greater than 40 years, explaining why the disease incidence is still raising in many countries despite working bans on the use of asbestos in the early 1990s. Platinum-based chemotherapy, mostly combined with the folate antagonist pemetrexed, is the established standard of care [1]. In earlier stages of pleural mesothelioma, multimodality therapy including extrapleural pneumonectomy or more recently extended pleurectomy/decortication, with or without radiotherapy, are being investigated in selected patients [2]. There is currently no defined standard for second-line therapy. The rationale behind investigating novel targeted approaches, available results and ongoing efforts are summarized in this review.

Exploring molecular alterations

Data mining of version 71 of the catalog of somatic mutations in cancer (COSMIC, <http://www.sanger.ac.uk/cosmic>) reveals that the genes that are most frequently mutated in malignant pleural mesothelioma are ‘cyclin-dependent kinase inhibitor

2A’ (*CDKN2A*), ‘neurofibromatosis type 2’ (*NF2*) and *BRCA*-‘associated protein 1’.

Targeting the cell cycle

Mesothelioma lack expression of both *CDKN2A* encoded proteins p16 and ARF due to gene deletion or methylation (reviewed in [3]). Deletion in *CDKN2A* leads to loss of control of cyclin D-dependent kinases (CDK). Although CDK4/6-specific inhibitors are under investigation in clinical trials, animal models with *CDKN2A* deficiency showed that loss of *CDKN2A* function is not necessarily associated with CDK4/6 addiction [4].

Although only a minor fraction of mesothelioma presents with p53 mutations [5], this led to the hypothesis that this tumor might be dependent on G2 checkpoint and therefore vulnerable to a G2 checkpoint inhibition when combined with chemotherapy. In line with this hypothesis, the calmodulin-binding peptide (CBP501) was clinically tested in combination with cisplatin and pemetrexed in order to increase the sensitivity of mesothelioma cells to chemotherapy [6]. In patients receiving CBP501 with chemotherapy PFS of more than 4 months was achieved compared with 39% receiving chemotherapy alone (Table 1) [7].

Targeting NF2/Hippo deficiency

The NF2/Hippo signaling pathway has been shown to be disrupted in most mesothelioma [5, 8] characterized by mutation or inactivation of the *NF2* gene (reviewed in [3]). Experimental animal models indicate that this event, together with a deficiency in

*Correspondence to: Prof. Rolf Stahel, Clinic of Oncology, University Hospital, Ramistrasse 100, 8091 Zürich, Switzerland. Tel: +41442552219; Fax: +41446342872; E-mail: rolf.stahel@usz.ch

Table 1. Clinical trials investigating targeted therapy in mesothelioma

Target	Clinical trial ID	Experimental arm	Mechanism	Control arm	Phase	Primary end points	Expected completion date	
Chemotherapy sensitizer	NCT00700336	CBP501 plus Cis/Pem	G2 checkpoint inhibition	Cis/Pem	I/II	Safety, MTD, PFS	Completed	
Altered NF2 signaling	NCT01138033	GSK2256098	Focal adhesion kinase inhibitor	Placebo	I	MTD	December 2014	
	NCT01870609	VS-6063	Focal adhesion kinase inhibitor		II	OS, PFS	December 2016	
	NCT02004028	VS-6064	Focal adhesion kinase inhibitor		II	PK	November 2014	
Altered NF2 signaling	NCT00854152	GDC-0980	PI3K/mTOR inhibitor		I	PK, MTD	August 2014	
	NCT01655225	LY3023414	PI3K/mTOR inhibitor		I	Recommended phase II dose	September 2014	
Altered NF2 signaling	NCT00770120	Everolimus	mTOR inhibitor		II	PFS, RR	completed	
	NCT01024946	Everolimus	mTOR inhibitor		II	PFS, RR	completed	
Arginine dependency	NCT01279967	Pegylated arginine deiminase	Growth inhibition of ASS-negative tumors	Best supportive care	II	PFS	January 2014	
Receptor tyrosine kinases	NCT00402766	Imatinib mesylate plus Cis/Pem	bcr/abl, c-kit, and PDGFR TKI		I	MTD	August 2015	
	NCT00703638	Sorafenib plus Cis/Pem	VEGFR2, VEGFR3, Raf, PDGFR, and c-kit TKI		I	MTD	Completed	
	NCT01064648	Cediranib plus Cis/Pem	VEGFR 1-3		Placebo (phase II)	I/II	MTD	June 2014
	NCT00700336	Dasatinib	Src inhibition and PDGFR TKI		I	Modulation of p-Src Tyr419	March 2016	
	NCT01592383	Erlotinib	EGFR TKI		Cis/pem	II	RR	June 2015
	NCT00459862	Pazopanib	VEGFR1-3, PDGFR, and c-Kit			II	PFS	Completed
	NCT01211275	Axitinib	pan-VEGFR inhibitor			I/II	Safety/efficacy	Unknown
	NCT01861301	Tivantinib	MET inhibitor			II	ORR	February 2015
	NCT02049060	Tivantinib plus Carbo/Pem	MET inhibitor			I	DLTs	July 2014
	NCT01590160	Ganetespiib plus Cis/Pem	Hsp90 inhibitor			I/II	DLTs	December 2015
Proteasome	NCT01307100	Nintedanib plus Cis/Pem	VEGFR, PDGFR, and FGFR TKI	Cis/Pem	II	PFS	May 2016	
	NCT01769547	Dovitinib	FGFR3 inhibitor		II	PFS	June 2016	
	NCT00996385	Bortezomib and oxaliplatin	Proteasome inhibitor		II	RR	September 2013	
	Aminopeptidase N	NCT01358084	NGR-hTNF		TNF targeting tumor blood vessels	Best supportive care	II	PFS

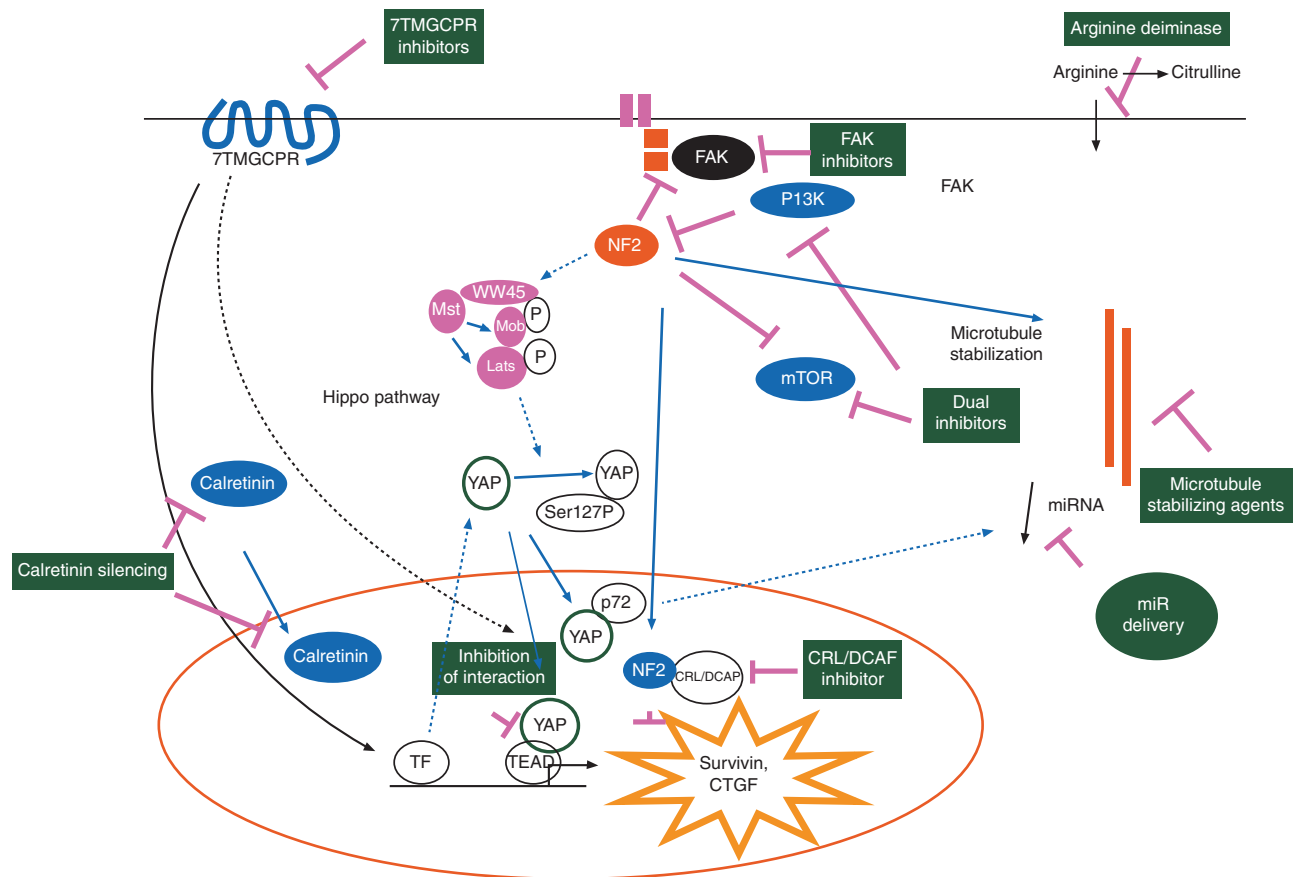


Figure 1. Genetic/epigenetic changes present in mesothelioma offer different possibilities for therapeutic intervention. Functional inactivation of NF2 and NF2/Hippo pathway offers the opportunity to intervene using focal adhesion kinase (FAK) inhibitors, PI3/mTOR dual inhibitors, microtubule stabilizing agents, E3 ubiquitin ligase CRL4/DCAF inhibitors, tumor suppressor miR delivery, inhibitors of the interaction between YAP and TEAD transcription activators, and 7 transmembrane G-coupled receptors (7TMGCCR) inhibitors regulating YAP activity. Auxotrophy for arginine render the tumor sensitive to the activity of arginine deiminase. Calretinin-dependent survival can be blocked by calretinin silencing.

cyclin-dependent kinase activator inhibitor (*CDKN2A*), is essential for mesothelioma development. Therefore, targeting molecules involved in the NF2/Hippo pathway is of major interest (Figure 1) for the treatment of mesothelioma. NF2 is an upstream regulator of the so-called Hippo signaling cascade, [9] which controls the transcriptional co-activator Yes-associated protein (YAP). The dysfunction of Hippo pathway, which leads to increased YAP activity [10], induces oncogenic transformation by the activation of transcription factors including transcription enhancer activation domain (TEAD) family members [11]. Upon binding TEADs, YAP/TAZ upregulates the expression of several growth promoting factors. YAP is constitutively active in more than 70% of primary mesotheliomas [8, 12]. Hedgehog signaling has a role in maintaining YAP protein stability in progenitor cells [13] and is activated in mesothelioma, consistent with the re-activation of a signaling known to be essential during embryonic mesothelial development [14]. Treatment of mesothelioma xenografts with the hedgehog antagonist HhAntag led to a decrease of the tumor volume accompanied by a decrease in Ki-67 labeling index.

Another possible approach is the direct inhibition of YAP activity. Three compounds related to porphyrin that could inhibit the transcriptional activity of YAP *in vitro* were identified by

screening of a Johns Hopkins Drug Library [15]. One of these, verteporfin, in clinical use as a photosensitizer in photocoagulation therapy for macular degeneration, was moderately effective at blocking mouse *Yap1*-overexpression- or loss of *Nf2*-driven hepatic tumorigenesis. These data suggest further investigation of these compounds as anticancer therapies.

NF2 suppresses tumorigenesis by migrating into the nucleus where it inhibits the E3 ubiquitin ligase CRL4 and through that controls a subset of Hippo pathway target genes [16]; therefore, CRL inhibitors such as MLN4924 should be investigated in mesothelioma.

Interestingly, expression of constitutively active YAP causes widespread miRNA suppression [17]. Thus, the Hippo pathway may be responsible for the widespread miRNA repression observed in cancer, including mesothelioma. To overcome the difficulties of directly delivering miRNA mimics, minicells composed of achromosomal bacterial cells and targeted by bispecific antibodies have been developed. These have been used to restore miR16 and induce growth arrest in mesothelioma xenografts [18]. Minicells can be given safely to patients with advanced cancer [19] and a clinical trial has started in mesothelioma (ACTRN12614001248651).

targeting NF2-associated cytoskeletal alterations

In a systematical screen for tumor suppressors whose functional inactivation would result in microtubular instability, NF2 was identified as a microtubule stabilizer [20], demonstrating that the microtubular network might be significantly affected in mesothelioma. Based on this and *in vitro* studies, epithilone B would be a candidate for clinical evaluation [21].

NF2 alterations result also in activation of the focal adhesion kinase (FAK) and merlin deficiency predicts for sensitivity to FAK inhibitors [22, 23]. The underlying mechanism is that survival and proliferation signals seem mediated through extracellular matrix-integrin signals promoting FAK activation in mesothelioma cells with inactivating NF2 mutations [23]. There is a phase I and two phase II trials ongoing testing the FAK inhibitors GSK2256098 and defactinib (VS-6063) in mesothelioma (Table 1). Determination of the NF2 status in these trials will allow exploring whether the clinical outcome is indeed associated with alteration of NF2.

targeting PI3K/mTOR

PI3K/mTOR signaling is activated in mesothelioma [24]. For the time being, the reason for this has not been elucidated, as neither PI3K nor receptor tyrosine kinase mutations/amplifications have been found in two recent high-throughput studies [5, 25]. NF2-null cells were shown sensitive to growth inhibitory effects of rapamycin [26] via mechanisms involving PI3K signaling-independent mammalian target of rapamycin complex (mTORC1) activation. However, preliminary results of a phase II trial [27] of the mTOR inhibitor, everolimus, did not show to be active in unselected MPM patients (Table 1). In addition, a peritoneal mesothelioma model was recently generated by deficiency in p53 and the tuberous sclerosis gene, a negative regulator of mTORC [28]. GDC-0980 is a small molecule inhibitor of class I PI3K and mTOR (mTORC1 and mTORC2) [29] has been tested in phase I studies (Table 1) and the preliminary result of the phase I extension cohort showed two objective responses among 26 patients with mesothelioma [30]. Another dual PI3K/mTOR inhibitor, LY3023414 is tested in a phase I trial (Table 1).

synthetic lethal approaches

A large proportion of mesothelioma [31] show reduced expression of arginosuccinate synthetase-1, the rate-limiting enzyme for arginine biosynthesis, rendering cells auxotrophic for arginine and consequently susceptible to the arginine degrading enzyme arginine deiminase (Adi-PEG20). Preliminary results of a randomized phase II study (Table 1) showed a significant PFS improvement delivering ADI-PEG20 versus best supportive care [32]. Almost all epithelioid and ~50% of sarcomatoid mesothelioma express calretinin [33] which is widely used as a robust diagnostic mesothelioma marker. Since its silencing inhibits mesothelioma cell survival *in vitro*, this may offer another opportunity for a therapeutic intervention [34].

tyrosine kinase inhibitors

Deregulated expression of growth factors or proteins involved in downstream signaling pathways has been shown to play an important role in malignant transformation of mesothelial cells.

Molecular studies in malignant pleural mesothelioma have confirmed that growth factors such as vascular endothelial growth factor (VEGF), platelet-derived growth factor receptor beta (PDGFR β) and the epidermal growth factor receptor family are frequently activated. Several clinical trials have tried to exploit these specific characteristics using tyrosine kinase inhibitors (TKIs).

multitargeted TKIs

Overexpression of platelet-derived growth factor (PDGF) has been observed and found to be associated with a poor prognosis [35]. While normal mesothelial cells express predominantly the PDGFR α subunit and less PDGFR β , mesothelioma was shown to overexpress PDGFR β [36]. *In vitro* studies show that VEGF stimulates the growth of mesothelioma cells and anti-VEGF rabbit polyclonal antibodies inhibit their growth [37–39].

The co-expression of c-kit in 26% [40] of mesothelioma has inspired the use of imatinib, an inhibitor of bcr/abl, c-kit, and PDGFR α and β . Four phase II clinical trials of imatinib as a single agent in mesothelioma have been published. Of a total of 94 patients treated, no objective response was seen and progression-free survival was less than 2 months [41–44].

Sorafenib is a potent inhibitor of VEGFR2, VEGFR3, Raf, PDGFR β , and c-kit. In a phase II trial with 50 patients assessable for response, 3 partial responses and 27 disease stabilizations were observed, results deemed insufficient for further evaluation of sorafenib [45]. Another phase II trial assessed sorafenib in a single-arm phase II study enrolling 53 patients using a Simon's two-stage design. Treatment was well tolerated and demonstrated a moderate activity with a median PFS of 5.1 months [46].

Sunitinib, a VEGFR1, VEGFR2, VEGFR3, PDGFR β , and c-kit TKI was tested in a phase II trial in 53 patients resulting in partial responses in 6 and stable disease in 34. An accompanying biomarker study was unsuccessful [47]. Another phase II trial using a Simon's two-stage design and a primary outcome of objective response rate did not meet the criteria for continuing to the second stage of accrual, with only one partial response observed among 35 patients [48]. A phase I trial with an expansion cohort in mesothelioma patients demonstrated that sunitinib was not well tolerated at 37.5 mg with standard pemetrexed and cisplatin doses, requiring dose reductions mainly due to cumulative myelosuppression and subsequent limited activity [49].

Pazopanib, a broad antiangiogenic broad TKI targeting VEGFR1-3, PDGFR α and β , and c-kit, has been evaluated in a phase II trial as a single agent in 34 mesothelioma patients resulting in a 6-month progression-free survival of 48% (Table 1). Vatalanib targets VEGFR1, VEGFR2, c-kit, PDGFR β , and c-Fms. It was tested in a phase II trial and did not achieve the protocol-specified 3-month PFS, ending its development mesothelioma [50]. Cediranib is a VEGFR 1–3 TKI. Two phase II trials were able to show only a modest single-agent activity with partial remission in 4 and 5 patients out of 54 and 51 patients, respectively, however with significant toxicities [51, 52].

Dasatinib is an inhibitor of the Src family of nonreceptor tyrosine kinases and PDGFR β . Single-agent dasatinib did not show any activity in mesothelioma and was associated with unacceptable pulmonary toxicities in a phase II trial enrolling 46 patients [53].

While the results from most of these trials were considered as negative, the fact remains that activity of cediranib, imatinib,

sunitinib, or sorafenib was observed in a low proportion of patients, suggesting a need for the identification of predictive biomarkers to support further development. However, given the multitargeted nature of these TKIs and the difficulties encountered in identifying biomarkers for antiangiogenic therapies in general this will unlikely be successful.

Due to the few responses to TKIs, combinatorial regimens with chemotherapy are ongoing. To this end, a study with cediranib in combination with chemotherapy is currently recruiting (Table 1) and another trial phase I/II trial has been randomizing patients to cisplatin and pemetrexed with or without axitinib, a pan-VEGFR inhibitor (Table 1). Several trials combining chemotherapy with multitargeted TKIs are still being discussed or currently in phase I (Table 1).

restricted TKI

Erlotinib and gefitinib are first-generation TKIs targeting specifically EGFR. EGFR expression has been demonstrated by immunohistochemistry in 70%–95% of mesothelioma specimens and its overexpression might be associated with a favorable prognosis [54–56]. Despite some encouraging *in vitro* data, phase II trials in patients with untreated mesothelioma using gefitinib or erlotinib have failed to demonstrate significant activity. Gefitinib demonstrated partial remissions in 2 of 43 untreated patients [56]. Erlotinib was ineffective in 63 untreated patients despite high expression of EGFR in patients' tumors. Here, the activation of the PI3K/Akt downstream pathways was proposed as a potential mechanism of primary resistance [57]. Also the combination of erlotinib and bevacizumab after platinum-based chemotherapy did not result in any responses among 24 mesothelioma patients [58].

Other targets including MET- and FGFR3-TKIs, tivantinib, and dovitinib, are under investigation (Table 1).

histone deacetylase inhibitors

The equilibrium between the acetylated and deacetylated forms of histone proteins is regulated by histone acetyltransferase and histone deacetylase (HDAC). HDAC inhibitors will alter the wrapping DNA around histones, modify the access of transcription factors and consequently impact the expression of various genes.

After two promising phase I trials including small numbers of mesothelioma treated by vorinostat as monotherapy or combined with chemotherapy, [59] a placebo-controlled phase III trial including 660 mesothelioma patients who had progressed after treatment with pemetrexed and platinum was launched. Results were reported at ECCO-ESMO 2011 and were negative for all end points [60]. Another small phase II trial with the HDAC inhibitor belinostat was also negative [61].

In vitro data suggested that valproic acid might have a proapoptotic effect in synergy with doxorubicine. A phase II trial evaluating valproic acid in combination with doxorubicine in 45 patients pretreated with chemotherapy demonstrated objective responses in seven with a median progression-free survival of 2.5 months [62].

proteasome inhibitors

Bortezomib was found to be inactive in a single-arm phase II trial in poor performance status first-line and second-line mesothelioma patients with only one confirmed response of 33 patients [63]. Bortezomib was also evaluated in combination with cisplatin in a prospective phase II trial with progression-free survival rate at 18 weeks as primary end point [64]. Eighty-two patients were treated with an 18-week progression-free survival of 53%. Based on statistical assumptions, the null hypothesis could not be rejected and the combination was considered not worthy of further investigation.

bevacizumab

A randomized phase II trial of untreated mesothelioma patients compared cisplatin-gemcitabine alone or with bevacizumab. The addition of bevacizumab did not improve response, progression-free survival, or overall survival compared with chemotherapy alone. A potential benefit in patients with low circulating levels of VEGF was suggested in subgroup analysis [65]. Another phase II trial combined treatment of cisplatin and pemetrexed with bevacizumab in 45 inoperable chemotherapy naïve mesothelioma patients. The response rate of 41%, median PFS of 6.9 months, and median OS 15.3 months were reported, with development of hypertension as a possible surrogate marker for bevacizumab activity [66].

A two-armed phase II/III trial compared the standard of care cisplatin and pemetrexed regimen with or without bevacizumab as first-line treatment and maintenance in inoperable mesothelioma patients. While tolerance was good, the preliminary analysis of the study revealed that disease control at 6 months favored the bevacizumab arm (73.5% and 43.2%, $P = 0.010$). Final results of this trial are eagerly awaited [67].

other antiangiogenic interventions

vascular disrupting agents

BNC105P is a small molecule inhibiting tubulin polymerization that functions as a vascular disrupting agent through selectively shutting down tumor blood vessels without affecting normal vasculature. Preclinical models have demonstrated significant tumor growth suppression and regression with BNC105P [68]. VDA BNC105P was tested as a second-line treatment in advanced mesothelioma and proven ineffective in a trial of 30 patients [69].

thalidomide

Thalidomide is the oldest and perhaps the most extensively studied drug classified as an antiangiogenic agent, which activity is attributed to the inhibition of VEGF, basic fibroblast growth factor, as well as Transforming growth factor- β (TGF- β) and tumor necrosis factor (TNF)- α [70, 71]. A phase I trial explored its activity in 40 mesothelioma patients, a third of them being treatment naïve. There were no responders, with an OS of 7.6 months and 11 were free of progression after 6 months [72]. Two parallel unpublished phase II studies evaluated thalidomide in combination with gemcitabine/cisplatin or thalidomide as a single agent. Thirty-one chemotherapy naïve patients received

thalidomide and gemcitabine/cisplatin with partial responses in 14% and an OS of 11 months [73]. Twenty-seven patients pre-treated or unsuitable for chemotherapy were treated with single-agent thalidomide. Responses occurred in 6% of the patients, and OS was 11 months.

The utility of thalidomide in mesothelioma as maintenance therapy for up to 1 year was evaluated in a large randomized phase III trial in 222 patients who had not progressed after 4–6 cycles of pemetrexed with or without platinum. The primary end point of time to progression was of 3.6 months in the experimental arm when compared with 3.5 months in the placebo arm, demonstrating the absence of benefit of thalidomide maintenance [74].

NGR-hTNF

NGR-hTNF consists of human tumor necrosis factor- α (hTNF- α) fused to the tumor-homing peptide asparagine-glycine-arginine (NGR) able to selectively bind an aminopeptidase N isoform over-expressed on tumor blood vessels. Based on an exploratory phase II trial in mesothelioma, a phase III trial comparing NGR-hTNF to best supportive care is ongoing [75].

immunotherapeutic approaches

In mesothelioma, a chronic inflammatory response represented by infiltrating lymphocytes and plasma cells was associated with a better prognosis [76]. As in other tumors, immunotherapeutic strategies aimed at balancing the immune system in favor of an anti-mesothelioma response are being explored (Figure 2).

transforming growth factor- β

TGF- β is a pleiotropic cytokine which can be produced by cancer, stroma, and immune cells [77–79]. TGF- β attenuates the antitumor immune response blocking priming and effector phase of tumor-specific T cells. Fresolimumab (GC1008), a humanized monoclonal antibody neutralizing active forms of TGF- β , has been examined in a phase I trial in 13 patients with mesothelioma [80]. No objective response was seen, but three patients had stable disease at 3 months and five patients developed an enhanced antibody response to mesothelioma lysates (Table 2).

interferon- β /interferon- α

Interferon- β (IFN- β) is type 1 cytokine with multiple functions resulting in antiviral, antiproliferative, antiangiogenic activity and immune cell stimulation. IFN- β has been delivered by an adenoviral vector into the pleural effusion of patient with mesothelioma in two phase I trials, using one or two injections [81, 82]. Both trials found a transient increase of IFN- β in the pleural cavity, mitigated by a neutralizing antibody response resulting in clearance of the adenovirus and decrease of IFN- β . There were no safety issues and antibody responses against the mesothelin could be induced in some patients. Of the 13 patients treated, 4 had stable disease as the best response.

IFN- α can promote the differentiation and activity of host immune cells and moreover correlates with generation of a durable antitumor response [83]; one clinical study exploring the efficacy of this cytokine in mesothelioma showed a response

in ~30% of patients when associated with chemotherapy, however with major toxicity [84]. One phase I trial have shown potential therapeutic benefit of IFN- α 2a gene transfer mediated by an adenoviral vector [85] and a new study with this approach is still ongoing (see Table 2).

intrapleural viruses

Viruses are strong stimulants to the immune system by the activation of the innate as well as the adaptive responses. The measles virus is oncolytic, resulting in tumor cell death and antigen release, allowing T-cell priming through dendritic cells. Also, immunoadjuvant properties of the measles virus were shown by loading dendritic cells with measles-infected mesothelioma cells, which resulted in dendritic cell maturation [86]. Phase I clinical trials are under way testing the intrapleural application of measles, herpes, and vaccinia virus in patients with malignant pleural mesothelioma (Table 2).

immune checkpoint inhibition

Cancer cells often inhibit T-cell activation to escape immune surveillance. After activation T cells express the cytotoxic T-lymphocyte antigen 4 (CTLA-4). When CTLA-4 binds members of the B7 family the T-cell response becomes abrogated [87]. Blocking monoclonal antibodies have been developed to prevent the negative feedback loop via CTLA-4. Tremelimumab, is a humanized monoclonal IgG2 antibody binding to CTLA-4. Tremelimumab has been tested as second line in mesothelioma in a phase II trial [88]. Two of 29 patients had durable partial responses. Although the primary end point of the study was not met, the disease control rate of 31% and progression-free survival of 6 months prompted further evaluation in an ongoing randomized phase II study (Table 2).

Expression of PD-L1 allows cancers to evade the host immune system by interaction with PD1. Treatment with antibodies targeting these molecules has resulted in extraordinary responses for advanced melanoma and lung cancer. Recently, the expression of PD-L1 has been demonstrated in mesothelioma [89]. Therefore, therapies targeting this pathway are of major interest and under development for mesothelioma patients.

mesothelin

Mesothelin is a cell surface glycoprotein expressed in mesothelial and peritoneal cells. Even if the biological function of mesothelin it not fully understood, it is known that mesothelin binds to CA-125 and is involved in cell adhesion (reviewed in [90]). Amatuximab is a chimeric IgG1 antibody targeting mesothelin. Studies demonstrated that it blocks the binding of mesothelin to CA 125 and thus could be used also as a strategy to prevent tumor metastasis [91]. Amatuximab was well tolerated in a phase I trial [92] and currently is tested in phase II in patients with mesothelioma (Table 2). Antibodies can be used to deliver cytotoxic agents to antigen expressing malignant cells. The potent bacterial toxin *Pseudomonas* exotoxin A (PE38) was linked to a disulfide-stabilized variable fragment based on the affinity modified variable light and heavy chain of amatuximab (SS1(dsFv)PE38) and showed preclinical activity [93]. SS1(dsFv)PE38 is under clinical investigation and was shown to be safe in two phase I trials, in which 16 patients with mesothelioma were

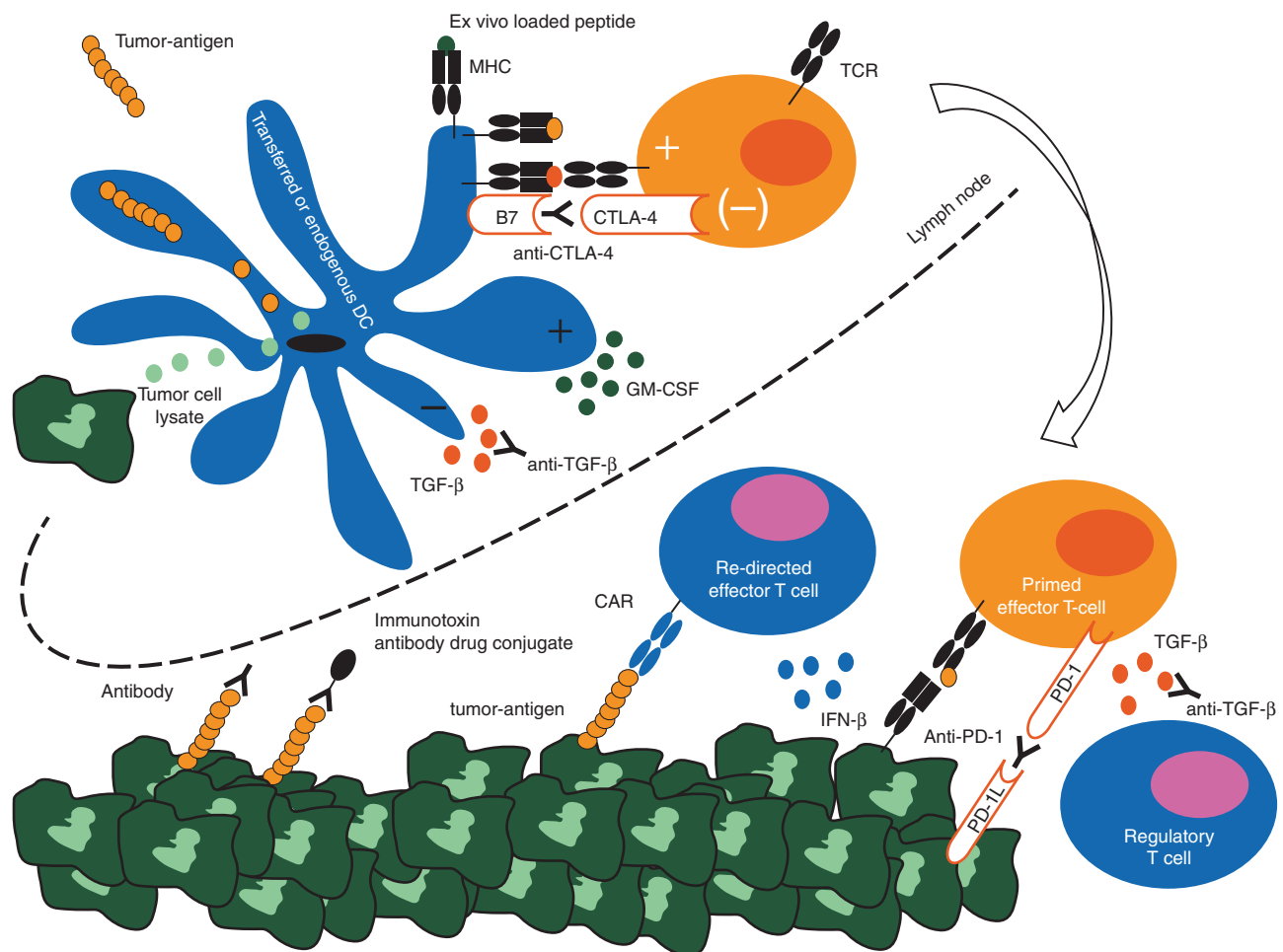


Figure 2. Interplay of the immune system with malignant mesothelioma and possible immunotherapeutic interventions. Malignant mesothelioma cells express tumor antigens such as WT1, FAP, mesothelin. Tumor antigens or other components from the malignant cells can be picked up by dendritic cells and presented to T cells inducing a T-cell response. However, molecules like TGF- β can block T-cell priming in the lymph node or T-cell function in malignant tissue. T-cell function can be also abrogated by activation of CTLA-4 and PD-1 on primed T cells. In addition, effector T cells can be converted in immunosuppressive regulatory T cells. Dendritic cell and re-directed T cells can be adoptively transferred. Dendritic cells loaded with peptides derived from tumor antigen induced T-cell responses. To enhance priming, GM-CSF is given in combination with dendritic cells. To circumvent T-cell priming, re-directed effector T cells can be transferred. IFN- γ is used to augment MHC expression on tumor cells increasing immunogenicity.

treated. Only minor antitumor activity could be observed. Additionally, the development of neutralizing antibodies was observed in 24% of patients prevented its use for more than one cycle [94]. In a subsequent pilot study using immunosuppressive pretreatment with pentostatin and cyclophosphamide to prevent neutralizing antibodies and allow delivery of more courses of treatment, 3 of 10 assessable patients had major responses [95].

MF-T is a fully humanized anti-mesothelin antibody conjugated to microtubule-targeting toxophore DM4 (BAY 94-9343). It showed selective cytotoxicity against mesothelioma cells, while sparing normal mesothelial cells, and potent *in vivo* activity against cell line and tumor xenografts [96]. This compound is thus a likely relevant candidate for further clinical testing.

In contrast to these described passive immunological interventions mesothelin can be targeted by active-specific vaccination. Live-attenuated *Listeria* vaccine expressing mesothelin has been tested in a phase I study with mesothelin-expressing

tumors. CRS-207 was well tolerated and mesothelin-specific CD8T-cell responses were detected [97]. Recent data, presented at ASCO 2014, showed that CRS-207 can be safely combined with standard of care chemotherapy and showed encouraging antitumor activity with 9 of 15 subjects having confirmed durable partial response and 4 stable disease (Table 2).

Wilms tumor suppressor gene 1

The Wilms tumor suppressor gene 1 (WT1), is a transcription factor highly expressed in mesothelioma and WT1 immunohistochemistry is among the routine procedures used for the diagnosis of mesothelioma. WT1 peptides are immunogenic and induce T-cell responses against mesothelioma cell lines [98]. In a first clinical trial class I and II, WT1 peptides were used for vaccination with s.c. GM-CSF, which is used to mature dendritic cells to augment T-cell priming. Of 9 patients with mesothelioma treated, one remained without progression after 3 years

Table 2. Clinical trials investigating immunotherapy in mesothelioma

Target	Clinical trial ID	Experimental arm	Mechanism	Control arm	Phase	Primary end points	Expected completion date
Immunosuppressive cytokine	NCT01112293	Anti-TGF- β monoclonal antibody (GC1008)	Blocking TGF- β		II	PFS	October 2012
Immunomodulating cytokine	NCT01212367	Gene transfer IFN- α 2a	Immunomodulating cytokine		I	Safety	December 2027
Immunoadjuvant	NCT01503177	Intraleural measles virus	Dendritic cell maturation; oncolytic virus		I	Safety	September 2014
	NCT01721018	Intraleural herpes virus	Oncolytic virus		I/II	Safety, PFS	April 2014
	NCT01766739	Intraleural vaccinia virus (GL-ONC1)	Oncolytic virus		I	Safety	January 2015
Immune checkpoint Tumor-antigen passive	NCT01843374	Tremelimumab	Blocking CTLA-4	Placebo	II	OS	May 2016
	NCT00738582	MORAb-009 (Amatuximab)	Anti-mesothelin monoclonal antibody with pemetrexed and cisplatin		II	PFS	November 2014
Tumor-antigen active-specific	NCT01675765	CRS-207 live-attenuated Listeria vaccine expressing mesothelin	Active-specific immune response against mesothelin followed pemetrexed and cisplatin		I	Safety	December 2015
	NCT01265433	WT-1 analog peptide vaccine plus GM-CSF	Adjuvant, active-specific immune response against WT-1	Montanide adjuvant + GM-CSF	II	PFS	December 2014
	NCT01890980	WT-1 analog peptide vaccine plus GM-CSF	Adjuvant, active-specific immune response against WT-1	Montanide adjuvant + GM-CSF	II	PFS	December 2017
	NCT01258868	Autologous tumor cell vaccine	Active-specific immune response against autologous tumor cells in combination with celecoxib and ISCOMATRIX		I	Safety	November 2018
	NCT01143545	Allogeneic tumor cell vaccine	Active-specific immune response against allogeneic tumor cells in combination with cyclophosphamide and celecoxib		I	Safety	May 2017
	NCT01569919	TroVax: pox virus specific for antigen 5T4	Pox virus carrying the 5T4 antigen plus Cis/Pem		II	Immune responses to 5T4, safety	June 2014
	NCT00280982	Tumor lysate-loaded autologous dendritic cells	Active-specific immune response against autologous tumor cells		I	Safety	Completed
Tumor-antigen active-specific adoptive transfer	NCT01241682	Tumor lysate-loaded autologous dendritic cells low-dose cyclophosphamide	Active-specific immune response against autologous tumor cells		I	Safety	Completed
Tumor-antigen adoptive transfer	NCT01583686	Adoptive transfer of mesothelin-specific re-directed T cells	T-cell response		I/II	Safety/PFS	March 2019
	NCT01355965	Adoptive transfer of mesothelin-specific re-directed T cells	T-cell response		I	Safety	May 2014
	NCT01722149	Adoptive transfer of FAP-specific re-directed T cells	T-cell response		I	Safety	May 2015

and five were documented to have a CD8 T-cell response [99]. Currently, randomized phase II trials with this vaccine are ongoing in patients after completion of multimodality therapy (Table 2).

vaccination with tumor cell lysate

Mesothelioma cell lysates are used for vaccination and can induce an antitumoral response. Twenty-two patients were treated with autologous tumor cell lysates and GM-CSF. In 32% of the patients, an immune response could be induced, but there was no objective response [100]. One clinical phase I trial is testing an autologous tumor cell vaccine with an adjuvant (ISCOMATRIX) and celecoxib to augment antigen presentation. The tumor cell vaccine is exposed *ex vivo* to demethylating agents to increase expression of tumor antigens (Table 2). Another phase one trial evaluates an allogeneic tumor cell vaccine (K526-GM) in combination with cyclophosphamide and celecoxib (Table 2). Cyclophosphamide is intended to eradicate regulatory T cells, which can inhibit dendritic cells to prime effector T cells [101]. Celecoxib is a COX-2 inhibitor resulting in decreased prostaglandin E2 (PGE2) and was used to block PGE2-mediated conversion of regulatory T cells and to allow dendritic cell maturation [102].

cellular therapy

Adoptive transfer of dendritic cells pulsed *ex vivo* with tumor antigens led to the first FDA approved cellular therapy (Sipuleucel-T) in prostate cancer [103]. In mesothelioma, a comparable approach was tested in 10 patients vaccinated with autologous dendritic cells. Each vaccine was composed of mature dendritic cells pulsed with autologous tumor lysate [104]. In four patients, dendritic cell vaccination induced cytotoxic T cells. Results from a trial evaluating dendritic cell-based vaccination in combination with low-dose cyclophosphamide are awaited (Table 2).

T cells can be re-directed against specific antigens. After gene transfer, autologous T cells express a chimeric antigen receptor (CAR), which enables the T cell to destroy target cells. Mesothelin-specific re-directed T cells were developed and showed *in vitro* and *in vivo* activities [105, 106]. To achieve transient expression, the plasmid with the gene sequence of the CAR was transferred in the T cells by electroporation [107]. Mesothelin-specific re-directed T cells are tested in early clinical trials (Table 2). An alternative target in malignant mesothelioma is the fibroblast activation protein (FAP) [108, 109]. FAP-specific re-directed T cells demonstrated antigen-specific activity *in vitro* and *in vivo* and are close to early clinical investigation (Table 2) [108, 110]. In this clinical trial, the adoptive transfer is planned to be carried out into the pleural effusion to overcome blocked T-cell trafficking [111].

discussion

In contrast to lung cancer, oncogenic driver mutations are absent in malignant mesothelioma. The development of targeted therapy therefore hinges on the exploration of pathways indirectly activated by the loss of tumor suppressor genes or targets associated with the disease phenotype. Efforts of targeting angiogenesis and cancer-associated receptor tyrosine kinases have shown disappointing results despite the enrollment of hundreds of mesothelioma patients in clinical trials. HDAC and proteasome

inhibitors were found to be inactive. The promising avenues for targeted therapies in mesothelioma include the functional consequences of alterations in NF2/Hippo pathway, and immunotherapeutic approaches. The inactivation of NF-2 and resulting merlin deficiency leads to a significantly increased activity of several pathways, including the hedgehog pathway, the activity of the focal adhesion kinase (FAK) and the PI3K/mTOR pathway. Inhibition of these pathways resulted in reproducible growth reduction of mesothelioma in preclinical models. Phase I trials in mesothelioma demonstrated clinical activity of FAK and of PI3K/mTOR inhibitors and a randomized phase II trial of the FAK inhibitor defactinib as maintenance therapy after chemotherapy has been initiated. In regard to immunotherapeutic approaches the jury is still out. However, based on early results in nonsmall-cell lung cancer and other solid tumors, it appears likely that immune checkpoint inhibitors will find a place in the therapy of mesothelioma.

funding

There was no direct funding for this manuscript. EF-B was supported by the Stiftung für Angewandte Krebsforschung, the Krebsliga Zürich and the Swiss National Science Foundation (CRSII3_147697); IS-O was supported by the Swiss National Science Foundation (PP00P3_133657).

disclosure

The authors have declared no conflicts of interest.

references

- Vogelzang NJ, Rusthoven JJ, Symanowski J et al. Phase III study of pemetrexed in combination with cisplatin versus cisplatin alone in patients with malignant pleural mesothelioma. *J Clin Oncol* 2003; 21: 2636–2644.
- Van Schil PE, Opitz I, Weder W et al. Multimodal management of malignant pleural mesothelioma: where are we today? *Eur Respir J* 2014; 44: 754–764.
- Felley-Bosco E, Stahel R. Hippo/YAP pathway for targeted therapy. *Transl Lung Cancer Res* 2014; 3: 75–83.
- Johnson SM, Torrice CD, Bell JF et al. Mitigation of hematologic radiation toxicity in mice through pharmacological quiescence induced by CDK4/6 inhibition. *J Clin Invest* 2010; 120: 2528–2536.
- Bott M, Brevet M, Taylor BS et al. The nuclear deubiquitinase BAP1 is commonly inactivated by somatic mutations and 3p21.1 losses in malignant pleural mesothelioma. *Nat Genet* 2011; 43: 668–672.
- Shapiro GI, Tibes R, Gordon MS et al. Phase I studies of CBP501, a G2 checkpoint abrogator, as monotherapy and in combination with cisplatin in patients with advanced solid tumors. *Clin Cancer Res* 2011; 17: 3431–3442.
- Krug LM, Wozniak AJ, Kindler HL et al. Randomized phase II trial of pemetrexed/cisplatin with or without CBP501 in patients with advanced malignant pleural mesothelioma. *Lung Cancer* 2014; 85: 429–434.
- Murakami H, Mizuno T, Taniguchi T et al. LATS2 is a tumor suppressor gene of malignant mesothelioma. *Cancer Res* 2011; 71: 873–883.
- Dong J, Feldmann G, Huang J et al. Elucidation of a universal size-control mechanism in *Drosophila* and mammals. *Cell* 2007; 130: 1120–1133.
- Striedinger K, VandenBerg SR, Baia GS et al. The neurofibromatosis 2 tumor suppressor gene product, merlin, regulates human meningioma cell growth by signaling through YAP. *Neoplasia* 2008; 10: 1204–1212.

11. Zhao B, Ye X, Yu J et al. TEAD mediates YAP-dependent gene induction and growth control. *Genes Dev* 2008; 22: 1962–1971.
12. Mizuno T, Murakami H, Fujii M et al. YAP induces malignant mesothelioma cell proliferation by upregulating transcription of cell cycle-promoting genes. *Oncogene* 2012; 31: 5117–5122.
13. Fernandez LA, Northcott PA, Dalton J et al. YAP1 is amplified and up-regulated in hedgehog-associated medulloblastomas and mediates Sonic hedgehog-driven neural precursor proliferation. *Genes Dev* 2009; 23: 2729–2741.
14. Dixit R, Ai X, Fine A. Derivation of lung mesenchymal lineages from the fetal mesothelium requires hedgehog signaling for mesothelial cell entry. *Development* 2013; 140: 4398–4406.
15. Liu-Chittenden Y, Huang B, Shim JS et al. Genetic and pharmacological disruption of the TEAD-YAP complex suppresses the oncogenic activity of YAP. *Genes Dev* 2012; 26: 1300–1305.
16. Li W, You L, Cooper J et al. Merlin/NF2 suppresses tumorigenesis by inhibiting the E3 ubiquitin ligase CRL4(DCAF1) in the nucleus. *Cell* 2010; 140: 477–490.
17. Mori M, Triboulet R, Mohseni M et al. Hippo signaling regulates microprocessor and links cell-density-dependent miRNA biogenesis to cancer. *Cell* 2014; 156: 893–906.
18. Reid G, Pel ME, Kirschner MB et al. Restoring expression of miR-16: a novel approach to therapy for malignant pleural mesothelioma. *Ann Oncol* 2013; 24: 3128–3135.
19. Solomon B, Desai J, Scott A et al. First-in-man, multicenter, phase I trial evaluating the safety of first-in-class therapeutic, EGFR-targeted, paclitaxel-packaged micellules. In 24th EORTC-NCI-AACR Symposium on Molecular Targets and Cancer Therapeutics. Dublin: 2012.
20. Smole Z, Thoma CR, Applegate KT et al. Tumor suppressor NF2/Merlin is a microtubule stabilizer. *Cancer Res* 2014; 74: 353–362.
21. Suraokar MB, Nunez MI, Diao L et al. Expression profiling stratifies mesothelioma tumors and signifies deregulation of spindle checkpoint pathway and microtubule network with therapeutic implications. *Ann Oncol* 2014; 25: 1184–1192.
22. Poulkakos PI, Xiao GH, Gallagher R et al. Re-expression of the tumor suppressor NF2/merlin inhibits invasiveness in mesothelioma cells and negatively regulates FAK. *Oncogene* 2006; 25: 5960–5968.
23. Shapiro IM, Kolev VN, Vidal CM et al. Merlin deficiency predicts FAK inhibitor sensitivity: a synthetic lethal relationship. *Sci Transl Med* 2014; 6: 237ra268.
24. Altomare DA, You H, Xiao GH et al. Human and mouse mesotheliomas exhibit elevated AKT/PKB activity, which can be targeted pharmacologically to inhibit tumor cell growth. *Oncogene* 2005; 24: 6080–6089.
25. Thomas RK, Baker AC, Debiasi RM et al. High-throughput oncogene mutation profiling in human cancer. *Nat Genet* 2007; 39: 347–351.
26. Lopez-Lago MA, Okada T, Murillo MM et al. Loss of the tumor suppressor gene NF2, encoding merlin, constitutively activates integrin-dependent mTORC1 signaling. *Mol Cell Biol* 2009; 29: 4235–4249.
27. Garland LL, Ou SH, Moon J et al. WOG 0722: a phase II study of mTOR inhibitor everolimus (RAD001) in malignant pleural mesothelioma (MPM). *J Clin Oncol* (In ASCO MEETING) 2012; 7083.
28. Guo Y, Chirieac LR, Bueno R et al. Tsc1-Tp53 loss induces mesothelioma in mice, and evidence for this mechanism in human mesothelioma. *Oncogene* 2014; 33: 3151–3160.
29. Salphati L, Pang J, Plise EG et al. Preclinical assessment of the absorption and disposition of the phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor GDC-0980 and prediction of its pharmacokinetics and efficacy in human. *Drug Metab Dispos* 2012; 40: 1785–1796.
30. Dolly S, Bendell JC, Kindler H et al. Evaluation of tolerability and anti-tumor activity of GDC-0980, an oral PI3K/mTOR inhibitor, administered to patients with advanced solid tumors or non-Hodgkin's lymphoma. In ECCO-ESMO-ESTRO 2013. Amsterdam: EJC 2013.
31. Szlosarek PW, Klabatsa A, Pallaska A et al. In vivo loss of expression of argininosuccinate synthetase in malignant pleural mesothelioma is a biomarker for susceptibility to arginine depletion. *Clin Cancer Res* 2006; 12: 7126–7131.
32. Phillips MM, Sheaff MT, Szlosarek PW. Targeting arginine-dependent cancers with arginine-degrading enzymes: opportunities and challenges. *Cancer Res Treat* 2013; 45: 251–262.
33. Ordonez NG. Value of caretinin immunostaining in diagnostic pathology: a review and update. *Appl Immunohistochem Mol Morphol* 2014; 22: 401–415.
34. Blum W, Schwaller B. Calretinin is essential for mesothelioma cell growth/survival in vitro: a potential new target for malignant mesothelioma therapy? *Int J Cancer* 2013; 133: 2077–2088.
35. Filiberti R, Marroni P, Neri M et al. Serum PDGF-AB in pleural mesothelioma. *Tumour Biol* 2005; 26: 221–226.
36. Ascoli V, Scalzo CC, Facciolo F, Nardi F. Platelet-derived growth factor receptor immunoreactivity in mesothelioma and non-neoplastic mesothelial cells in serous effusions. *Acta Cytol* 1995; 39: 613–622.
37. Konig JE, Tolnay E, Wiethage T, Muller KM. Expression of vascular endothelial growth factor in diffuse malignant pleural mesothelioma. *Virchows Arch* 1999; 435: 8–12.
38. Konig J, Tolnay E, Wiethage T, Muller K. Co-expression of vascular endothelial growth factor and its receptor flt-1 in malignant pleural mesothelioma. *Respiration* 2000; 67: 36–40.
39. Strizzi L, Catalano A, Vianale G et al. Vascular endothelial growth factor is an autocrine growth factor in human malignant mesothelioma. *J Pathol* 2001; 193: 468–475.
40. Arber DA, Weiss LM, West RB. CD117 expression in mesothelioma. *Mod Pathol* 2004; 17: 1021.
41. Porta C, Mutti L, Tassi G. Negative results of an Italian Group for Mesothelioma (G.I.Me.) pilot study of single-agent imatinib mesylate in malignant pleural mesothelioma. *Cancer Chemother Pharmacol* 2007; 59: 149–150.
42. Mathy A, Baas P, Dalesio O, van Zandwijk N. Limited efficacy of imatinib mesylate in malignant mesothelioma: a phase II trial. *Lung Cancer* 2005; 50: 83–86.
43. Villano J, Husain AN, Stadler MB et al. A phase II trial of imatinib mesylate in patients (pts) with malignant mesothelioma (MM). *J Clin Oncol* 2004; 22: 14.
44. Millward M, Parnis F, Byrne M et al. Phase II trial of imatinib mesylate in patients with advanced pleural mesothelioma. *Am J Clin Oncol* 2003; 22: 912.
45. Dubey S, Janne PA, Krug L et al. A phase II study of sorafenib in malignant mesothelioma: results of Cancer and Leukemia Group B 30307. *J Thorac Oncol* 2010; 5: 1655–1661.
46. Papa S, Popat S, Shah R et al. Phase 2 study of sorafenib in malignant mesothelioma previously treated with platinum-containing chemotherapy. *J Thorac Oncol* 2013; 8: 783–787.
47. Nowak AK, Millward MJ, Creaney J et al. A phase II study of intermittent sunitinib malate as second-line therapy in progressive malignant pleural mesothelioma. *J Thorac Oncol* 2012; 7: 1449–1456.
48. Laurie SA, Gupta A, Chu Q et al. Brief report: a phase II study of sunitinib in malignant pleural mesothelioma. The NCIC Clinical Trials Group. *J Thorac Oncol* 2011; 6: 1950–1954.
49. Camidge DR, Blais N, Jonker DJ et al. Sunitinib combined with pemetrexed and cisplatin: results of a phase I dose-escalation and pharmacokinetic study in patients with advanced solid malignancies, with an expanded cohort in non-small cell lung cancer and mesothelioma. *Cancer Chemother Pharmacol* 2013; 71: 307–319.
50. Jahan T, Gu L, Kratzke R et al. Vatalanib in malignant mesothelioma: a phase II trial by the Cancer and Leukemia Group B (CALGB 30107). *Lung Cancer* 2012; 76: 393–396.
51. Garland LL, Chansky K, Wozniak AJ et al. Phase II study of cediranib in patients with malignant pleural mesothelioma: SWOG S0509. *J Thorac Oncol* 2011; 6: 1938–1945.
52. Campbell NP, Kunnavakkam R, Leigh N et al. Cediranib in patients with malignant mesothelioma: a phase II trial of the University of Chicago Phase II Consortium. *Lung Cancer* 2012; 78: 76–80.
53. Dudek AZ, Pang H, Kratzke RA et al. Phase II study of dasatinib in patients with previously treated malignant mesothelioma (cancer and leukemia group B 30601): a brief report. *J Thorac Oncol* 2012; 7: 755–759.
54. Destro A, Ceresoli GL, Falleni M et al. EGFR overexpression in malignant pleural mesothelioma. An immunohistochemical and molecular study with clinicopathological correlations. *Lung Cancer* 2006; 51: 207–215.
55. Edwards JG, Swinson DE, Jones JL et al. EGFR expression: associations with outcome and clinicopathological variables in malignant pleural mesothelioma. *Lung Cancer* 2006; 54: 399–407.
56. Govindan R, Kratzke RA, Herndon JE, III et al. Gefitinib in patients with malignant mesothelioma: a phase II study by the Cancer and Leukemia Group B. *Clin Cancer Res* 2005; 11: 2300–2304.

57. Garland LL, Rankin C, Gandara DR et al. Phase II study of erlotinib in patients with malignant pleural mesothelioma: a Southwest Oncology Group Study. *J Clin Oncol* 2007; 25: 2406–2413.
58. Jackman DM, Kindler HL, Yeap BY et al. Erlotinib plus bevacizumab in previously treated patients with malignant pleural mesothelioma. *Cancer* 2008; 113: 808–814.
59. Kelly WK, O'Connor OA, Krug LM et al. Phase I study of an oral histone deacetylase inhibitor, suberoylanilide hydroxamic acid, in patients with advanced cancer. *J Clin Oncol* 2005; 23: 3923–3931.
60. Krug L. Vorinostat in patients with advanced malignant pleural mesothelioma who have failed prior pemetrexed and either cisplatin or carboplatin therapy: a phase III randomized, double-blind, placebo-controlled trial. In ECCO-ESMO 2011. Stockholm: 2011.
61. Ramalingam SS, Belani CP, Ruel C et al. Phase II study of belinostat (PXD101), a histone deacetylase inhibitor, for second line therapy of advanced malignant pleural mesothelioma. *J Thorac Oncol* 2009; 4: 97–101.
62. Scherpereel A, Berghmans T, Lafitte JJ et al. Valproate-doxorubicin: promising therapy for progressing mesothelioma. A phase II study. *Eur Respir J* 2011; 37: 129–135.
63. Fennell DA, McDowell C, Busacca S et al. Phase II clinical trial of first or second-line treatment with bortezomib in patients with malignant pleural mesothelioma. *J Thorac Oncol* 2012; 7: 1466–1470.
64. O'Brien ME, Gaafar RM, Popat S et al. Phase II study of first-line bortezomib and cisplatin in malignant pleural mesothelioma and prospective validation of progression free survival rate as a primary end-point for mesothelioma clinical trials (European Organisation for Research and Treatment of Cancer 08052). *Eur J Cancer* 2013; 49: 2815–2822.
65. Kindler HL, Karrison TG, Gandara DR et al. Multicenter, double-blind, placebo-controlled, randomized phase II trial of gemcitabine/cisplatin plus bevacizumab or placebo in patients with malignant mesothelioma. *J Clin Oncol* 2012; 30: 2509–2515.
66. Dowell J, Gerber DE, Dunphy F. Association of hypertension (HTN) and clinical outcome in a phase II trial of cisplatin (C), pemetrexed (P), and bevacizumab (B) in patients with untreated malignant mesothelioma. *J Clin Oncol* 2010; 28: 15.
67. Zalcman G, Margery J, Scherpereel A et al. IFCT-GFFC-0701 MAPS trial, a multicenter randomized phase II/III trial of pemetrexed-cisplatin with or without bevacizumab in patients with malignant pleural mesothelioma. *J Clin Oncol* 2010; 28: 7020.
68. Siemann DW. The unique characteristics of tumor vasculature and preclinical evidence for its selective disruption by tumor-vascular disrupting agents. *Cancer Treat Rev* 2011; 37: 63–74.
69. Nowak AK, Brown C, Millward MJ et al. A phase II clinical trial of the vascular disrupting agent BNC105P as second line chemotherapy for advanced malignant pleural mesothelioma. *Lung Cancer* 2013; 81: 422–427.
70. Tamilarasan KP, Kolluru GK, Rajaram M et al. Thalidomide attenuates nitric oxide mediated angiogenesis by blocking migration of endothelial cells. *BMC Cell Biol* 2006; 7: 17.
71. De Sanctis JB, Mijares M, Suarez A et al. Pharmacological properties of thalidomide and its analogues. *Recent Pat Inflamm Allergy Drug Discov* 2010; 4: 144–148.
72. Baas P, Boogerd W, Dalesio O et al. Thalidomide in patients with malignant pleural mesothelioma. *Lung Cancer* 2005; 48: 291–296.
73. Pavlakis N, Abraham R, Harvie R. Thalidomide alone or in combination with cisplatin/gemcitabine in malignant pleural mesothelioma (MM); interim results from two parallel non randomized phase II studies. *Lung Cancer* 2003; 41: 11.
74. Buikhuisen WA, Burgers JA, Vincent AD et al. Thalidomide versus active supportive care for maintenance in patients with malignant mesothelioma after first-line chemotherapy (NVALT 5): an open-label, multicentre, randomised phase 3 study. *Lancet Oncol* 2013; 14: 543–551.
75. Gregorc V, Zucali PA, Santoro A et al. Phase II study of asparagine-glycine-arginine-human tumor necrosis factor alpha, a selective vascular targeting agent, in previously treated patients with malignant pleural mesothelioma. *J Clin Oncol* 2010; 28: 2604–2611.
76. Suzuki K, Kadota K, Sima CS et al. Chronic inflammation in tumor stroma is an independent predictor of prolonged survival in epithelioid malignant pleural mesothelioma patients. *Cancer Immunol Immunother* 2011; 60: 1721–1728.
77. Suzuki E, Kapoor V, Cheung HK et al. Soluble type II transforming growth factor-beta receptor inhibits established murine malignant mesothelioma tumor growth by augmenting host antitumor immunity. *Clin Cancer Res* 2004; 10: 5907–5918.
78. Fitzpatrick DR, Peroni DJ, Bielefeldt-Ohmann H. The role of growth factors and cytokines in the tumorigenesis and immunobiology of malignant mesothelioma. *Am J Respir Cell Mol Biol* 1995; 12: 455–460.
79. Marzo AL, Fitzpatrick DR, Robinson BW, Scott B. Antisense oligonucleotides specific for transforming growth factor beta2 inhibit the growth of malignant mesothelioma both in vitro and in vivo. *Cancer Res* 1997; 57: 3200–3207.
80. Stevenson JP, Kindler HL, Papasavvas E et al. Immunological effects of the TGFbeta-blocking antibody GC1008 in malignant pleural mesothelioma patients. *Oncoimmunology* 2013; 2: e26218.
81. Serman DH, Recio A, Carroll RG et al. A phase I clinical trial of single-dose intrapleural IFN-beta gene transfer for malignant pleural mesothelioma and metastatic pleural effusions: high rate of antitumor immune responses. *Clin Cancer Res* 2007; 13: 4456–4466.
82. Serman DH, Recio A, Haas AR et al. A phase I trial of repeated intrapleural adenoviral-mediated interferon-beta gene transfer for mesothelioma and metastatic pleural effusions. *Mol Ther* 2010; 18: 852–860.
83. Dunn GP, Koebel CM, Schreiber RD. Interferons, immunity and cancer immunoeediting. *Nat Rev Immunol* 2006; 6: 836–848.
84. Parra HS, Tixi L, Latteri F et al. Combined regimen of cisplatin, doxorubicin, and alpha-2b interferon in the treatment of advanced malignant pleural mesothelioma: a phase II multicenter trial of the Italian Group on Rare Tumors (GTR) and the Italian Lung Cancer Task Force (FONICAP). *Cancer* 2001; 92: 650–656.
85. Serman DH, Haas A, Moon E et al. A trial of intrapleural adenoviral-mediated Interferon-alpha2b gene transfer for malignant pleural mesothelioma. *Am J Respir Crit Care Med* 2011; 184: 1395–1399.
86. Gauvrit A, Brandler S, Sapede-Peroz C et al. Measles virus induces oncolysis of mesothelioma cells and allows dendritic cells to cross-prime tumor-specific CD8 response. *Cancer Res* 2008; 68: 4882–4892.
87. Salama AK, Hodi FS. Cytotoxic T-lymphocyte-associated antigen-4. *Clin Cancer Res* 2011; 17: 4622–4628.
88. Calabro L, Morra A, Fonsatti E et al. Tremelimumab for patients with chemotherapy-resistant advanced malignant mesothelioma: an open-label, single-arm, phase 2 trial. *Lancet Oncol* 2013; 14: 1104–1111.
89. Mansfield AS, Roden AC, Peikert T et al. B7-H1 expression in malignant pleural mesothelioma is associated with sarcomatoid histology and poor prognosis. *J Thorac Oncol* 2014; 9: 1036–1040.
90. Tang Z, Qian M, Ho M. The role of mesothelin in tumor progression and targeted therapy. *Anticancer Agents Med Chem* 2013; 13: 276–280.
91. Ma J, Tang WK, Esser L et al. Recognition of mesothelin by the therapeutic antibody MORAb-009: structural and mechanistic insights. *J Biol Chem* 2012; 287: 33123–33131.
92. Hassan R, Cohen SJ, Phillips M et al. Phase I clinical trial of the chimeric anti-mesothelin monoclonal antibody MORAb-009 in patients with mesothelin-expressing cancers. *Clin Cancer Res* 2010; 16: 6132–6138.
93. Li Q, Verschraegen CF, Mendoza J, Hassan R. Cytotoxic activity of the recombinant anti-mesothelin immunotoxin, SS1(dsFv)PE38, towards tumor cell lines established from ascites of patients with peritoneal mesotheliomas. *Anticancer Res* 2004; 24: 1327–1335.
94. Kreitman RJ, Hassan R, Fitzgerald DJ, Pastan I. Phase I trial of continuous infusion anti-mesothelin recombinant immunotoxin SS1P. *Clin Cancer Res* 2009; 15: 5274–5279.
95. Hassan R, Miller AC, Sharon E et al. Major cancer regressions in mesothelioma after treatment with an anti-mesothelin immunotoxin and immune suppression. *Sci Transl Med* 2013; 5: 208ra147.
96. Golfier S, Kopitz C, Kahmert A et al. Anetumab ravtansine—a novel mesothelin-targeting antibody-drug conjugate cures tumors with heterogeneous target expression favored by bystander effect. *Mol Cancer Ther* 2014; 13: 1537–1548.
97. Le DT, Brockstedt DG, Nir-Paz R et al. A live-attenuated *Listeria* vaccine (ANZ-100) and a live-attenuated *Listeria* vaccine expressing mesothelin (CRS-207) for advanced cancers: phase I studies of safety and immune induction. *Clin Cancer Res* 2012; 18: 858–868.

98. May RJ, Dao T, Pinilla-Ibarz J et al. Peptide epitopes from the Wilms' tumor 1 oncoprotein stimulate CD4+ and CD8+ T cells that recognize and kill human malignant mesothelioma tumor cells. *Clin Cancer Res* 2007; 13: 4547–4555.
99. Krug LM, Dao T, Brown AB et al. WT1 peptide vaccinations induce CD4 and CD8T cell immune responses in patients with mesothelioma and non-small cell lung cancer. *Cancer Immunol Immunother* 2010; 59: 1467–1479.
100. Powell A, Creaney J, Broomfield S et al. Recombinant GM-CSF plus autologous tumor cells as a vaccine for patients with mesothelioma. *Lung Cancer* 2006; 52: 189–197.
101. Le DT, Jaffee EM. Regulatory T-cell modulation using cyclophosphamide in vaccine approaches: a current perspective. *Cancer Res* 2012; 72: 3439–3444.
102. Whittaker DS, Bahjat KS, Moldawer LL, Clare-Salzler MJ. Autoregulation of human monocyte-derived dendritic cell maturation and IL-12 production by cyclooxygenase-2-mediated prostanoid production. *J Immunol* 2000; 165: 4298–4304.
103. Kantoff PW, Higano CS, Shore ND et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med* 2010; 363: 411–422.
104. Hegmans JP, Veltman JD, Lambers ME et al. Consolidative dendritic cell-based immunotherapy elicits cytotoxicity against malignant mesothelioma. *Am J Respir Crit Care Med* 2010; 181: 1383–1390.
105. Lanitis E, Poussin M, Hagemann IS et al. Redirected antitumor activity of primary human lymphocytes transduced with a fully human anti-mesothelin chimeric receptor. *Mol Ther* 2012; 20: 633–643.
106. Moon EK, Carpenito C, Sun J et al. Expression of a functional CCR2 receptor enhances tumor localization and tumor eradication by retargeted human T cells expressing a mesothelin-specific chimeric antibody receptor. *Clin Cancer Res* 2011; 17: 4719–4730.
107. Zhao Y, Moon E, Carpenito C et al. Multiple injections of electroporated autologous T cells expressing a chimeric antigen receptor mediate regression of human disseminated tumor. *Cancer Res* 2010; 70: 9053–9061.
108. Schuberth PC, Hagedorn C, Jensen SM et al. Treatment of malignant pleural mesothelioma by fibroblast activation protein-specific re-directed T cells. *J Transl Med* 2013; 11: 187.
109. Garin-Chesa P, Old LJ, Rettig WJ. Cell surface glycoprotein of reactive stromal fibroblasts as a potential antibody target in human epithelial cancers. *Proc Natl Acad Sci USA* 1990; 87: 7235–7239.
110. Petrusch U, Schuberth PC, Hagedorn C et al. Re-directed T cells for the treatment of fibroblast activation protein (FAP)-positive malignant pleural mesothelioma (FAPME-1). *BMC Cancer* 2012; 12: 615.
111. Scherpereel A, Grigoriu BD, Noppen M et al. Defect in recruiting effector memory CD8+ T-cells in malignant pleural effusions compared to normal pleural fluid. *BMC Cancer* 2013; 13: 324.

Annals of Oncology 26: 1660–1667, 2015
doi:10.1093/annonc/mdv245
Published online 22 May 2015

Should docetaxel be standard of care for patients with metastatic hormone-sensitive prostate cancer? Pro and contra

K. Fizazi¹, C. Jenkins² & I. F. Tannock^{3*}

¹Department of Cancer Medicine, Gustave Roussy, University of Paris Sud, Paris, France; ²Med Ed, RMC, Exeter, UK; ³Princess Margaret Cancer Centre and University of Toronto, Toronto, Canada

Received 21 March 2015; revised 10 May 2015; accepted 13 May 2015

Following the results of the TAX-327 study, questions have been raised as to whether administering chemotherapy to men with prostate cancer before symptomatic disease progression when receiving standard hormonal treatment can improve the duration and quality of patient survival. The GETUG-AFU-15 and CHAARTED studies both assessed the efficacy and tolerability of androgen deprivation therapy (ADT) with or without docetaxel in men with metastatic hormone-naïve prostate cancer. Both studies included a mix of patients with *de novo* metastatic disease (~75%) and patients who developed metastases following treatment of localized disease. A short course of ADT was allowed in both trials prior to accrual. Key differences between the two studies include the number of patients with high-volume metastases (GETUG-AFU-15: 52%; CHAARTED: 65%) and number of docetaxel cycles (GETUG-AFU-15: up to nine cycles; CHAARTED six cycles). Both studies reported an improvement in progression-free survival with docetaxel plus ADT versus ADT alone. The GETUG-AFU-15 did not find a significant difference in the primary end point of overall survival (OS) [hazard ratio (HR) 0.9 [95% confidence interval (CI) 0.7–1.2]; $P = 0.44$] for ADT plus docetaxel versus ADT alone. The CHAARTED study met the primary end point of OS [HR 0.61 (95% CI 0.47–0.80); $P = 0.0003$], and in a subset analysis reported the greatest improvement in OS for patients with high-volume disease [HR 0.60 (95% CI 0.45–0.81); $P = 0.0006$]. The following article

*Correspondence to: Prof. Ian F. Tannock, Department of Medical Oncology, Princess Margaret Cancer Centre and University of Toronto, 610 University Avenue, Toronto, ON, Canada M5G 2M9. Tel: +1-416-946-2245; Fax: +1-416-946-4563; E-mail: ian.tannock@uhn.ca