

Reappraising the theme of breeding systems in *Echinococcus*: is outcrossing a rare phenomenon?

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SUMMARY

Selfing has been considered the most common mode of reproduction in *Echinococcus* flatworms. However, population genetic studies on the asexual larval stage involving nuclear co-dominant markers have not always revealed significant heterozygote deficiencies – the expected outcome of a regularly and highly inbred population. In this study, we analysed the genetic structure of *Echinococcus granulosus sensu lato* populations from Southern Brazil during their adult (sexual) stage using 1 mitochondrial and 1 nuclear marker (*cox 1* and *mdh*, respectively). We show that parasite genetic differentiation is largest among definitive hosts (domestic dogs) from different farms, suggesting that transmission is mostly maintained within a farm. Moreover, we show that heterozygote deficiencies are not significant, and we suggest that outbreeding is the most common mode of reproduction of the parasite in that region.

Key words: selfing, outcrossing, *Echinococcus granulosus*, flatworm, mating system.

INTRODUCTION

Echinococcus is a hermaphroditic cestode that lives in the intestine of carnivores (generally canids) during the adult phase of its life cycle. The larvae usually develop in the liver and lungs of herbivorous mammals such as rodents, artiodactyls and perissodactyls. The natural cycle is maintained through predator-prey interactions, and the domestic cycle is completed when dogs eat the uncooked viscera of infected intermediate hosts (sheep, cattle, goats etc). *Echinococcosis* is a globally prevalent zoonosis with considerable economical impacts. The genus *Echinococcus* is undergoing a taxonomic revision (Thompson and McManus, 2002) as its most common and widely distributed species, *E. granulosus*, contains a high degree of genetic variation correlating to its intermediate host species. Thus, some genotypes or strains of *E. granulosus* that had adapted to different intermediate hosts were recently split in separate species (Thompson and McManus, 2002; Nakao *et al.* 2007).

It has been generally accepted that selfing causes strain variation within the genus *Echinococcus* (Smyth and Smyth, 1964). An opposing but less-accepted explanation for strain divergence is natural selection

(revised by Thompson and Lymbery, 1988). Rausch (1986) proposed that gene flow and cross-fertilization are extensive in *E. granulosus*, and he attributed strain differentiation to the association with distinct domestic host species. Evidence for the traditional view is the highly significant deficiency of heterozygotes and linkage disequilibrium in *E. granulosus* populations from Australia and Brazil (Lymbery *et al.* 1997; Haag *et al.* 1999). Nevertheless, some degree of outcrossing must occur to account for the presence of heterozygotes (Badaraco *et al.* 2008).

Outcrossing is believed to be more advantageous than selfing because it produces more variable offspring that might better respond to various selection pressures and suffer less from the expression of deleterious mutations (Wright, 1977; Maynard Smith, 1978). In *Schistocephalus solidus*, a hermaphroditic tapeworm whose cycle is maintained through fish-eating birds (definitive hosts), copepods (first intermediate hosts) and sticklebacks (second intermediate hosts), outbred parasites showed higher infectivity and developmental rates (Christen *et al.* 2002). Model simulations used to estimate extinction probabilities have suggested that hermaphroditism associated with selfing in the free-living nematode *Caenorhabditis elegans* must have arisen relatively recently, or that low levels of outcrossing and other factors are key to the species' persistence into the present day (Loewe and Cutter, 2008).

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Most models on mating system evolution predict that a mixture of selfed and outcrossed progeny should not exist. Uyenoyama (1986) found that mixed-mating systems (with both outcrossing and selfing) would be evolutionary stable only when self-fertilization and bi-parental inbreeding occur simultaneously. Indeed, it was shown that adults of *S. solidus* self-fertilize a fraction of their eggs, but the amount of selfed progeny is highly variable among individuals (Lüscher and Milinski, 2003). A mixed-mating system seems to be maintained by stochastic density fluctuations in the host (uncertainty in finding a mate increases the benefit of selfing). In *Echinococcus*, microscopy studies of adult worms show contradictory results, indicating either that self-insemination is the normal process of sperm transfer in *E. granulosus* (Smyth and Smyth, 1969), or that mating between worms is the main form of reproduction (Wang, 1998).

Understanding the mode of reproduction in the *Echinococcus* would enable us to predict how parasite populations respond to drugs or vaccines. Resistance mutations may spread rapidly in an outcrossing population. However, genetic studies within the group have focused mainly on uni-parental mitochondrial markers and on the larval (asexual) stage. To our knowledge, this is the first study that assesses the genetic structure of adult *E. granulosus* populations using both mitochondrial (uni-parental) and nuclear (bi-parental) markers.

MATERIALS AND METHODS

Sampling and preparation of parasite materials

Echinococcus adults measuring between 2 and 3 mm were obtained from 6 dogs on 3 farms in the rural area of Santana do Livramento, Southern Brazil. Dogs were purged with 3 mg/kg arecoline (Schantz, 1973). Worms were collected from the feces, washed in 1X PBS (pH 7.4), and stored at 4 °C in 70% ethanol. To account for the possibility that the eggs within a gravid proglottid originated from cross-fertilization between 2 genetically distinct worms, the last proglottid was separated from the rest of the body with a scalpel. Each body part was first dried for 30 min at 37 °C and then incubated in 50 µl of a 3 ng/µl proteinase K solution at 58 °C for 2 h. Proteinase K was inactivated at 95 °C for 20 min, and the eluates were stored at –20 °C until use.

PCR, SSCP and sequencing

Three targets were amplified by PCR and subsequently analysed: (1) a 366 bp fragment of the mitochondrial cytochrome oxidase gene (*cox1*, Bowles *et al.* 1992); (2) a 106 bp region including the first intron of the cytosolic malate dehydrogenase gene (*mdh*, referred to as *EgAg4* in Haag *et al.* 1999);

and (3) a 214 bp fragment containing the second intron of the same nuclear gene (Badaraco *et al.* 2008). PCR reactions were performed as previously described using 2 µl of the thawed eluates as templates.

To screen for nucleotide polymorphisms of the nuclear *mdh* gene, we used Single Strand Conformation Polymorphism (SSCP) to identify intron I and II alleles and to search for heterozygous genotypes. SSCP was performed using the GeneGel Clean SSCP kit on a GenePhor Electrophoresis Unit (GE Healthcare) following the manufacturer's instructions. The gels ran for 1 h at 14 °C and 500 V (intron I), or 2 h at 12 °C and 200 V (intron II). After electrophoresis, gels were silver stained using a conventional protocol.

We confirmed allele and genotype assignments for *mdh* introns I and II by sequencing the PCR fragments that showed distinctive SSCP patterns. All *cox1* PCR products were analysed by nucleotide sequencing. Briefly, amplicons were purified using ExoSAP-IT (USB) and sequenced automatically in both directions (Macrogen). Chromatograms were inspected for quality and used to search for nucleotide polymorphisms with the CodonCode Aligner software.

Statistical analyses

We hierarchically grouped parasite genotypes into infra-populations (worms collected from a single dog) and populations (worms from all dogs on a single farm). Genetic variance was analysed by AMOVA (Excoffier *et al.* 1992) using the Arlequin 3.5 software (Excoffier and Lischer, 2010), and was classified into the following categories: within dogs (infra-populations), among dogs within farms (populations) and among farms (meta-population). To estimate inbreeding, we performed a second molecular variance analysis for each individual worm.

RESULTS

From a total of 49 worms, 36 showed successful amplification for the 3 markers (see Supplementary table in online version). Results for each body segment were always identical, with no exception, allowing us to pool the data from a single individual. Malate dehydrogenase introns I and II also showed consistent results (Table 1). Four mitochondrial haplotypes were present in our sample: G1 and G1b (sheep strain), G3 (buffalo strain) and G5 (cattle strain). According to the taxonomic classification proposed by Nakao *et al.* (2007), G1, G1b and G3 correspond to *E. granulosus sensu stricto*, and G5 to *E. ortleppi*. For convenience, we use here the suffix *sensu lato* to refer to all mitochondrial haplotypes found in our study. As Table 1 shows, every dog was

Table 1. Genotypes of the 36 *Echinococcus granulosus sensu lato* adults analysed in our study

Farm	Coordinates	Dog	n	Genotype		
				<i>cox1</i>	<i>mdh</i> intron I	<i>mdh</i> intron II
P01	S:30°54'16" W:55°52'08"	D03	2	G1b	A1/A1	Md1/Md1
P12	S:30°51'51" W:55°38'56"	D34	1	G3	A1/A2	Md1/Md2
		D35	6	G5	A3/A3	Md3/Md3
P20	S:30°51'34" W:55°48'43"	D58	2	G1	A1/A1	Md1/Md1
			2	G1	A1/A2	Md1/Md2
			1	G1	A2/A2	Md2/Md2
			6	G1	A1/A1	Md1/Md1
			6	G1	A1/A2	Md1/Md2
		D61	5	G1	A2/A2	Md2/Md2
			1	G1	A1/A1	Md1/Md1
			2	G1	A1/A2	Md1/Md2
			2	G1	A2/A2	Md2/Md2
Total			36			

Table 2. Analysis of molecular variance (AMOVA) without the individual level

Source of variation	Sum of squares	Variance components	% Variation
Among farms	27.11	0.85	61.87
Among dogs on farms	4.71	0.10	7.78
Within dogs	27.54	0.42	30.35
Total	59.36	1.37	

parasitized by a single mitochondrial haplotype with 1 exception: dog P20D59 contained 22 worms with haplotype G1 and 1 with haplotype G1b (Supplementary table). However, because the eluate derived from this individual did not amplify the nuclear markers, it was excluded from further analyses.

There was a slight but not significant, deficiency in *mdh* heterozygotes in farm P20 ($n=27$). Other populations have rather low sample sizes for testing Hardy-Weinberg proportions. One infra-population (dog P12D35, $n=6$) was monomorphic (Table 1). The inbreeding coefficient (F) could be estimated by the deviation from $H-W$ proportions as $F=1-H/He$ (Nei, 1987), where H is the actual population heterozygosity and He is the expected heterozygosity under equilibrium. In farm P20, F equals 0.26. So if F is interpreted as the proportion of selfed individuals (McCauley *et al.* 1985), that means the parasite on that farm outcrosses 74% of the time.

When genetic variance is partitioned using AMOVA (Table 2), more than half of the total variation (61.87%) is found among farms (i.e. within the entire meta-population). However, an expressive fraction of the total genetic variation (30.35%) occurs among worms from the same definitive host (within infra-populations). Conducting the molecular variance analyses on the individual level shows

Table 3. Analysis of molecular variance (AMOVA) considering the individual level

Source of variation	Sum of squares	Variance components	% Variation
Among farms	27.11	0.85	61.89
Among dogs on farms	4.71	0.10	6.88
Within dogs	16.54	0.12	8.96
Within individual worms	11.00	0.30	22.26
Total	59.36	1.37	

that the largest fraction of the intra-population variability occurs within individual worms (22.26%, see Table 3). The estimated fixation indexes are as follows: $F_{IS}=0.29$ ($P=0.02$); $F_{SC}=0.18$ ($P=0.03$); $F_{CT}=0.62$ ($P=0.00$); $F_{IT}=0.77$ ($P=0.00$).

DISCUSSION

Transmission and implications for echinococcosis control

Four *Echinococcus granulosus sensu lato* haplotypes (G1, G1b, G3 and G5) are circulating among the 6 farms included in our study. However, these genetic variants are not randomly distributed among dogs and, with a single exception, no dog ever harboured more than 1 parasite haplotype. Transmission seems to occur mainly within farms. The 3 dogs from farm P20 played host to only G1 and G1b adults, while dogs from farm P12 hosted haplotypes G3 and G5. AMOVA shows that the largest fraction of genetic variance occurs among farms (61.87%). The genetic differentiation (fixation index) of infra-populations (dogs) from different farms ($F_{CT}=0.62$) is much larger than within farms ($F_{SC}=0.18$), pointing to a scenario where the parasite is transmitted primarily

by the habit of feeding dogs uncooked viscera. Dogs in our study area do not move far from the main farmhouse (de la Rue, personal observation). Furthermore, although taeniid eggs might be passively dispersed over distances up to 10 km (possibly by the wind or insects), their viability is rather low (Lawson and Gemmel, 1983).

These findings suggest that educational or other control programmes performed on a farm-by-farm basis would have a good chance of successfully reducing parasite transmission. They also explain the success of a similar control programme against echinococcosis in Tasmania in 1965, which led to the eradication of the parasite (see Jenkins (2005) for a review). With no wild definitive hosts transmitting the parasite across farms, denying domestic dogs access to offal and dosing them with anti-helminthics, together with public education and vigilance at abattoirs and farm quarantine policies, would interrupt the transmission cycle. A similar programme conducted in Uruguay in 1991, in which all domestic dogs were treated with praziquantel monthly, resulted in a remarkable decrease in the prevalence of ovine echinococcosis (Oku *et al.* 2004). Farias *et al.* (2004) also conducted a dog-treatment programme in the region in Brazil where the present samples were obtained. This programme also successfully decreased the prevalence in dogs; however, because it was difficult to introduce behavioural changes in the rural population and stop the feeding of uncooked sheep viscera to dogs, the infection rate increased quickly soon after the programme finished.

Selfing versus outcrossing

Previous studies from our group on a large set of markers using the larval stage of *E. granulosus sensu lato* (Haag *et al.* 1999) suggest that the Smyth and Smyth (1964) and Rausch (1986) models of strain evolution in *Echinococcus* are not mutually exclusive. We proposed that both selfing and outcrossing might occur in *E. granulosus* populations. No study since then has taken up the question, leaving reproduction modes in *Echinococcus* populations still a matter of some speculation, despite their clear epidemiological implications (e.g. evolution and spread of resistant phenotypes). Mitochondrial genes became the most popular markers for studying *Echinococcus* molecular epidemiology, but due to their uni-parental and non-recombining properties, they are inadequate to assess questions about breeding systems. Nuclear markers have frequently been considered to be less variable, with the exception of a microsatellite (EmsB, Bart *et al.* 2006), which is unfortunately repeated inside the genome, and therefore useless for assessing breeding systems as well.

Moreover, population studies have focused almost exclusively on the asexual metacestode stage.

However, comparing *Echinococcus* population polymorphisms in both intermediate and definitive hosts is essential to understanding the dynamics of genetic variation during the parasite's life cycle. Although it does not provide direct evidence for the mode of reproduction, it can help test hypotheses. In this present work on the adult stage of *Echinococcus*, as well as in our former study on the larval stage (Haag *et al.* 1999), we found no significant heterozygote deficiencies. Lymbery *et al.* (1997) and Badaraco *et al.* (2008) on the other hand, found significant heterozygote deficiencies. In light of the results obtained in the present study, which show that transmission among domestic animals seems to be restricted to a single farm, it is possible that the heterozygote deficiencies previously described are a consequence of subdivision (Wahlund effect). Pooling genotypes from different farms with distinct allele frequencies could lead to statistically significant heterozygote deficiencies, resembling an inbreeding effect. Badaraco *et al.* (2008), for example, tested the Hardy-Weinberg equilibrium for the *mdh* locus in a pooled sample of bovine isolates from southern Brazil corresponding to haplotype G1 ($n=115$). The frequency of alleles Md1, Md2 and Md3, was 0.42, 0.56 and 0.02, respectively, leading to an expected frequency of genotype Md1/Md2 under H-W equilibrium equal to 0.47. However, the bovine hosts came from different farms, which may have had different parasite allele frequencies. If, for instance, samples from 2 farms showing allele frequencies of Md1 equal to 0.8 and 0.04 (0.42 on average) and of Md2 equal to 0.18 and 0.94 (0.56 on average) would have been pooled, the expected heterozygosity under H-W equilibrium would be 0.29 for the first farm and 0.08 for the second! Heterozygotes have consistently been found both in *E. granulosus sensu lato* (Lymbery *et al.* 1997; Haag *et al.* 1999; Badaraco *et al.* 2008) and in *E. multilocularis* (Knapp *et al.* 2007), suggesting that cross-fertilization is not uncommon.

Indeed, the difference between the observed and expected heterozygosity in farm P20 ($F=0.26$), and the fixation index of infra-populations averaged over all loci ($F_{IS}=0.29$) suggests that outcrossing is actually the most common mode of reproduction. In the extreme case of a completely selfed population, F should equal 1, although selfing rates may vary from one region to another, depending on particular ecological conditions. Cheptou and Dieckmann (2002) showed that, contrary to previous models of breeding system evolution, the outcomes of breeding system evolution are not confined to either complete selfing or full outcrossing – even under demographic equilibrium – and that intermediate selfing rates arise under a wide range of conditions depending on the nature of competitive interactions between inbred and outbred individuals. Thus, it seems more plausible that the Smyth and Smyth (1964) and Rausch (1986) models of *Echinococcus* strain

evolution represent two ends of a continuum of selfing rates.

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REFERENCES

- Badaraco, J. L., Ayala, F. J., Bart, J.-M., Gottstein, B. and Haag, K. L.** (2008). Using mitochondrial and nuclear markers to evaluate the degree of genetic cohesion among *Echinococcus* populations. *Experimental Parasitology* **119**, 453–459.
- Bart, J. M., Knapp, J., Gottstein, B., El-Garch, F., Giraudoux, P., Glowatzki, M. L., Berthoud, H., Maillard, S. and Piarroux, R.** (2006). EmsB, a tandem repeated multi-loci microsatellite, new tool to investigate the genetic diversity of *Echinococcus multilocularis*. *Infection Genetics and Evolution* **6**, 390–400.
- Bowles, J., Blair, D. and McManus, D. P.** (1992). Genetic variants within the genus *Echinococcus* identified by mitochondrial DNA sequencing. *Molecular and Biochemical Parasitology* **54**, 165–174.
- Cheptou, P.-O. and Dieckmann, U.** (2002). The evolution of self-fertilization in density-regulated populations. *Proceedings of the Royal Society of London, B* **269**, 1177–1186.
- Christen, M., Kurtz, J. and Milinski, M.** (2002). Outcrossing increases infection success and competitive ability: experimental evidence from a hermaphrodite parasite. *Evolution* **56**, 2243–2251.
- Excoffier, L. and Lischer, H. E. L.** (2010). Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources* **10**, 564–567.
- Excoffier, L., Smouse, P. E. and Quattro, J. M.** (1992). Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* **131**, 479–491.
- Farias, L. N., Malgor, R., Cassaravilla, C., Bragança, C. and De La Rue, M.** (2004). Echinococcosis in Southern Brazil: efforts toward implementation of a control program in Santana do Livramento, Rio Grande do Sul. *Revista do Instituto de Medicina Tropical de São Paulo* **46**, 153–156.
- Haag, K. L., Araújo, A. M., Gottstein, B., Siles-Lucas, M., Thompson, R. C. A. and Zaha, A.** (1999). Breeding systems in *Echinococcus granulosus* (Cestoda; Taeniidae): selfing or outcrossing? *Parasitology* **118**, 63–71.
- Jenkins, D. J.** (2005). Hydatid control in Australia: where it began, what we have achieved and where to from here. *International Journal for Parasitology* **35**, 733–740.
- Knapp, J., Bart, J. M., Glowatzki, M. L., Ito, A., Gerard, S., Maillard, S., Piarroux, R. and Gottstein, B.** (2007). Assessment of use of microsatellite polymorphism analysis for improving spatial distribution tracking of *Echinococcus multilocularis*. *Journal of Clinical Microbiology* **45**, 2943–2950.
- Lawson, R. and Gemmel, M. A.** (1983). Hydatidosis and cysticercosis: The dynamics of transmission. *Advances in Parasitology* **22**, 331–369.
- Loewe, L. and Cutter, A. D.** (2008). On the potential for extinction by Muller's Ratchet in *Caenorhabditis elegans*. *BMC Evolutionary Biology* **8**, 125.
- Lüscher, A. and Milinski, M.** (2003). Simultaneous hermaphrodites reproducing in pairs self-fertilize some of their eggs: an experimental test of predictions of mixed-mating and Hermaphrodite's Dilemma theory. *Journal of Evolutionary Biology* **16**, 1030–1037.
- Lymbery, A. J., Constantine, C. C. and Thompson, R. C. A.** (1997). Self-fertilization without genomic or population structuring in a parasitic tapeworm. *Evolution* **51**, 289–294.
- Maynard Smith, J.** (1978). *The Evolution of Sex*. Cambridge University Press, Cambridge, UK.
- McCauley, D. E., Whittier, D. P. and Reilly, L. M.** (1985). Inbreeding and the rate of self-fertilization in a grape fern, *Botrychium dissectum*. *American Journal of Botany* **72**, 1978–1981.
- Nakao, M., McManus, D. P., Schantz, P. M., Craig, P. S. and Ito, A.** (2007). A molecular phylogeny of the genus *Echinococcus* inferred from complete mitochondrial genomes. *Parasitology* **134**, 713–722.
- Nei, M.** (1987). *Molecular Evolutionary Genetics*. Columbia University Press, New York, USA.
- Oku, Y., Malgor, R., Benavidez, U., Carmona, C. and Kamiya, H.** (2004). Control program against hydatidosis and the decreased prevalence in Uruguay. *International Congress Series* **1267**, 98–104.
- Rausch, R. L.** (1986). Life-cycle patterns and distribution of *Echinococcus* species. In *The Biology of Echinococcus and Hydatid Disease* (ed. Thompson, R. C. A.), pp. 44–80. George Allen & Unwin, London, UK.
- Schantz, P. M.** (1973). Guía para el empleo del bromhidrato de arecolina en el diagnóstico de la infección por *Echinococcus granulosus* en el perro. *Boletín Chileno de Parasitología* **28**, 81–90.
- Smyth, J. D. and Smyth, M. M.** (1964). Natural and experimental hosts of *Echinococcus granulosus* and *E. multilocularis*, with comments on the genetics of speciation in the genus *Echinococcus*. *Parasitology* **54**, 493–514.
- Smyth, J. D. and Smyth, M. M.** (1969). Self insemination of *Echinococcus granulosus* in vivo. *Journal of Helminthology* **43**, 383–388.
- Thompson, R. C. A. and Lymbery, A. J.** (1988). The nature, extent and significance of variation within the genus *Echinococcus*. *Advances in Parasitology* **27**, 209–249.
- Thompson, R. C. A. and McManus, D. P.** (2002). Towards a taxonomic revision of the genus *Echinococcus*. *Trends in Parasitology* **18**, 452–457.
- Uyenoyama, M. K.** (1986). Inbreeding and the cost of meiosis: the evolution of selfing in populations practicing biparental inbreeding. *Evolution* **40**, 388–404.
- Wang, H.** (1998). A study on morphology of reproductive organs of *Echinococcus granulosus* by light microscopy, transmission and scanning electron. *Endemic Diseases Bulletin* **3**, 31–33.
- Wright, S.** (1977). *Experimental Results and Evolutionary Deductions*. University of Chicago Press, Chicago, IL, USA.