

The transmembrane serine protease (TMPRSS3) mutated in deafness DFNB8/10 activates the epithelial sodium channel (ENaC) *in vitro*

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TMPRSS3 encodes a transmembrane serine protease that contains both LDLRA and SRCR domains and is mutated in non-syndromic autosomal recessive deafness (DFNB8/10). To study its function, we cloned the mouse ortholog which maps to *Mmu17*, which is structurally similar to the human gene and encodes a polypeptide with 88% identity to the human protein. RT-PCR and RNA *in situ* hybridization on rat and mouse cochlea revealed that *Tmprss3* is expressed in the spiral ganglion, the cells supporting the organ of Corti and the stria vascularis. RT-PCR on mouse tissues showed expression in the thymus, stomach, testis and E19 embryos. Transient expression of wild-type or tagged TMPRSS3 protein showed a primary localization in the endoplasmic reticulum. The epithelial amiloride-sensitive sodium channel (ENaC), which is expressed in many sodium-reabsorbing tissues including the inner ear and is regulated by membrane-bound channel activating serine proteases (CAPs), is a potential substrate of TMPRSS3. In the *Xenopus* oocyte expression system, proteolytic processing of TMPRSS3 was associated with increased ENaC mediated currents. In contrast, 6 TMPRSS3 mutants (D103G, R109W, C194F, W251C, P404L, C407R) causing deafness and a mutant in the catalytic triad of TMPRSS3 (S401A), failed to undergo proteolytic cleavage and activate ENaC. These data indicate that important signaling pathways in the inner ear are controlled by proteolytic cleavage and suggest: (i) the existence of an auto-catalytic processing by which TMPRSS3 would become active, and (ii) that ENaC could be a substrate of TMPRSS3 in the inner ear.

INTRODUCTION

Hearing loss is the most common sensory defect in humans, affecting about one in 1000 children (1). About 60% of all

cases of sensorineural deafness are of genetic origin (2) and non-syndromic autosomal recessive deafness accounts for ~70% of genetic hearing loss cases (2). Thirty-three loci that cause autosomal recessive non-syndromic hearing loss

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have been identified and seventeen of these genes have been isolated (<http://dnalab-www.uia.ac.be/dnalab/hhh/index.html>). Mutations in the *TMPRSS3* gene, which encodes a transmembrane serine protease, were identified in both familial and sporadic cases of non-syndromic sensorineural recessive deafness, DFNB8/10 (3–6). *TMPRSS3* (for transmembrane protease, serine 3) is a type II transmembrane serine protease (TTSP) and like the other members of this family, it contains a short N-terminal tail, a transmembrane (TM) domain and a large C-terminal segment characterized by: (i) a stem region containing LDLRA and SRCR domains; (ii) a potential proteolytic activation cleavage site between R216 and I217; and (iii) a serine protease domain containing the catalytic triad signature (H257, D304 and S401) (Fig. 1B). This structure suggests that *TMPRSS3* could be active as a membrane-bound enzyme (7).

Channel activating proteases (CAPs) are membrane-bound serine proteases that are able to activate ENaC *in vitro* (8,9). Recently, three CAPs have been identified in a mouse kidney cell line (mCAP1, mCAP2, mCAP3) (10). Interestingly, like *TMPRSS3*, mCAP2 is a type II transmembrane serine protease incorporating LDLRA and SRCR domains. mCAP2 is the mouse ortholog of hTMPRSS4. Recently, it has been suggested that ENaC plays a role in the establishment and maintenance of the remarkably low sodium concentration of the endolymph (11,12). This observation raises the possibility that ENaC could be a substrate of *TMPRSS3* in the inner ear.

To study the function of *TMPRSS3* in the inner ear, we isolated the mouse *Tmprss3* and characterized its expression profile and the intracellular localization of the human *TMPRSS3* protein. We show that *Tmprss3* co-localizes with ENaC in some inner ear tissues and that, unlike wild-type *TMPRSS3*, *TMPRSS3* mutants causing deafness fail to undergo proteolytic processing and to activate ENaC *in vitro*.

RESULTS

Isolation of the mouse *Tmprss3* cDNA and gene

Using the methods described later, we constructed a 2752 bp mouse *Tmprss3* cDNA encoding a putative protein of 453 amino acids that showed 88% identity to human *TMPRSS3* (Fig. 1B) (3). Similar to *TMPRSS3*, *Tmprss3* is divided into 12 coding exons over 18 kb of genomic sequence, but there was no experimental evidence for the existence of a 5'UTR exon in the mouse gene (Fig. 1A). No alternative mouse transcripts were identified.

Tmprss3 mRNA expression

RT-PCR was performed to study the expression profile of *Tmprss3*. Significant expression was found in thymus, stomach, testis, E19 embryos, stria vascularis, modiolus, organ of Corti and cultured spiral ganglion (Figs. 2A and B). No amplification from genomic DNA was observed.

To confirm these data, RNA *in situ* hybridization was performed on mouse embryos and inner ear sections. *Tmprss3* expression was not detected in the otocyst of mouse embryos from E8.5 to E10.5 d.p.c. (data not shown). Further analyses were performed on mouse postnatal day 5 when the inner ear development is almost complete. *Tmprss3* expression was

detected in the spiral ganglion and in the entire epithelium that supports cells of the organ of Corti but was not observed in sensory hair cells. Weak expression was also found in the stria vascularis (Fig. 3).

Intracellular localization of human *TMPRSS3* protein

The intracellular localization of the *TMPRSS3* protein was investigated by using wild-type and N- or C-terminal tagged *TMPRSS3* proteins. All *TMPRSS3* proteins tested, showed cytoplasmic localization in HeLa, Cos-7, U2OS and MDCK cells (Fig. 4A–D, G and data not shown). Co-localization experiments showed that the *TMPRSS3* protein is primarily localized in the endoplasmic reticulum membranes where it may be anchored by its transmembrane domain (Fig. 4F and I and data not shown).

Expression analysis of the α -, β - and γ -ENaC subunits in the rat inner ear

The ENaC channel is expressed in the inner ear and is known to be regulated by serine protease activity. These features make it a candidate substrate for *TMPRSS3*. To investigate this possibility, the co-expression of *Tmprss3* and the three ENaC subunits was analysed by RT-PCR on a panel of rat inner ear tissues. Similar to *Tmprss3*, α - and γ -subunits were detected in all the samples tested. The β -subunit was detected in all samples tested except the organ of Corti and the spiral ganglion, even after nested PCR (Fig. 2B).

Functional and biochemical analysis of *TMPRSS3* in Xenopus oocytes

The six *TMPRSS3* missense mutations associated deafness can be divided into three main groups: (i) LDLRA domain mutants (D103G, R109W); (ii) an SRCR domain mutant (C194F); and (iii) serine protease domain mutants (W251C, P404L, C407R) (4–6), (Fig. 5A). Amino acid sequence comparisons of a portion of the LDLRA, SRCR and serine protease domains of the human and mouse *TMPRSS3* with few selected such domains of other proteins are available at http://medgen.unige.ch/public_1.html

Co-expression of the human WT-*TMPRSS3* cRNA with the rat ENaC subunits (rENaC) led to an increase in ENaC mediated current (I_{Na}) ($P < 0.01$) compared to oocytes injected only with rENaC (Fig. 5B, lane 2 versus lane 1). Oocytes injected only with water or *TMPRSS3* did not generate measurable current (data not shown). The LDLRA domain mutants (D103G, R109W) retained a residual but significant ($P < 0.05$) activity (lanes 3 and 4 versus lane 2). The SRCR domain mutant (C194F) was totally inactive (lane 5 versus lane 2). The catalytic domain mutants were either totally inactive (W251C, P404L) (lanes 6 and 7 versus lane 2) or partially active (C407R), leading to a small but significant ($P < 0.05$) ENaC activation (lane 8 versus lane 2). As a negative control, the S401A catalytic triad mutant was totally inactive (lane 9 versus lane 2). Expression of the pro domain (M1-S215) of *TMPRSS3* (pro domain, pro) decreased baseline activity ($P < 0.01$, lane 10 versus lane 1) whereas the serine protease domain (M-C204-T454) of *TMPRSS3* (SP domain, SP) was inactive (lane 11 versus lane 2). As shown in

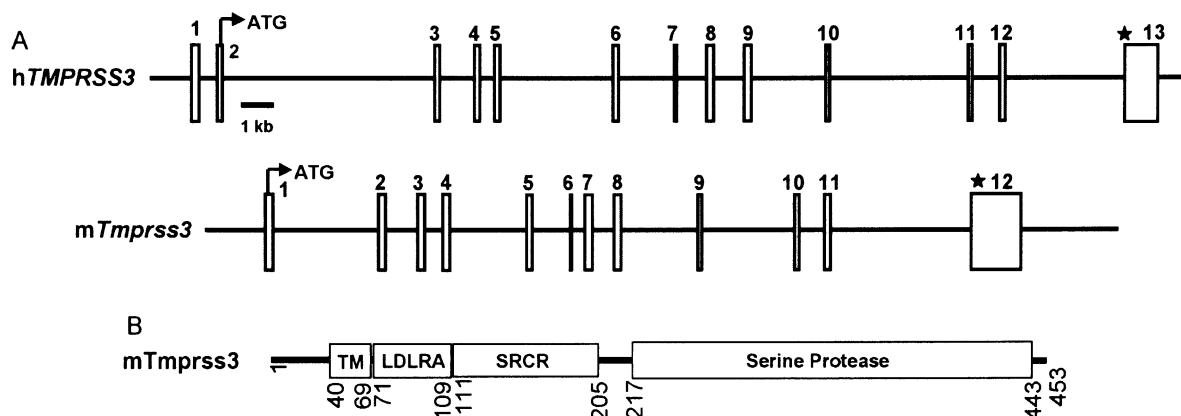


Figure 1. (A) Schematic representation of the human and mouse *TMPRSS3* genomic structure. Exons are represented as boxes numbered on top, with start and stop/codons indicated. Introns are shown as thin lines. (B) Schematic representation of the mouse *Tmprss3* protein showing the TM, LDLRA, SRCR and serine protease domains and their position in the 453 amino acid polypeptide.

Figure 5C, trypsin was able to activate ENaC significantly in the absence of TMPRSS3 (lane 1). When ENaC was co-expressed with WT-TMPRSS3, however, subsequent exposure to trypsin had only a small additional effect (lane 2) indicating a non-additive, common signalling pathway between trypsin and TMPRSS3. By contrast, when ENaC was co-expressed with TMPRSS3 mutants (lanes 3–9), pro (lane 10) or SP domains (lane 11), sodium currents were significantly enhanced by subsequent perfusion with trypsin.

When expressed in *Xenopus* oocytes, WT-TMPRSS3 was detected as three protein species on a western blot analysis: a predominant doublet of 50 and 54 kDa and a faint 30 kDa fragment suggesting that a significant fraction of TMPRSS3 was proteolytically processed (Fig. 5D). The faint 30 kDa fragment migrated at the same position as the catalytic serine protease domain (SP domain, lane 11) consistent with an auto-proteolytic cleavage at the activation site between R216 and I217, as occurs on other TTSPs (7). The 25 kDa non-catalytic subunit of TMPRSS3 (pro domain, lane 10) could not be detected for WT-TMPRSS3, suggesting additional proteolytic cleavage(s). The unprocessed isoform of TMPRSS3 was detected for all TMPRSS3 deafness mutants (lanes 3–8) and the S401A mutant (lane 9) indicating that they were all well expressed but not cleaved in the *Xenopus* oocyte expression system. These results indicate that the LDLRA and SRCR domains and the serine protease domain are all implicated in the ENaC channel activation by TMPRSS3 and in the processing of TMPRSS3.

DISCUSSION

After our identification of TMPRSS3 as the gene responsible for DFNB8/10 (3), we identified additional mutations in *TMPRSS3* (4–6) and pursued a variety of approaches to investigate the role of TMPRSS3 in the hearing process. In this paper, we report the isolation of the mouse ortholog and subsequent expression studies. We also report that TMPRSS3 which may localize in the endoplasmic reticulum membranes, undergoes cleavage and activates ENaC whereas the TMPRSS3 mutant proteins causing deafness are all unprocessed and

inactive. We also demonstrate that ENaC is co-expressed with *Tmprss3* in the inner ear.

The specific ion composition of the endolymph, characterized by high potassium and low sodium concentrations, is essential for the proper function of the sensory hair cells. Genes important for potassium recycling are mutated in different forms of syndromic and non-syndromic deafness (13,14). In contrast, little is known about the transport systems and sites involved in sodium homeostasis. The ENaC channel is a highly selective sodium channel that is expressed in many sodium-reabsorbing tissues and is a good candidate for sodium reabsorption from the endolymph. Indeed, the three ENaC subunits mRNAs have been detected in rat inner ear. Their expression levels were found to increase from postnatal day 2 reaching the highest levels of expression at postnatal day 12 when the concentration of sodium in endolymph reaches adult levels (11,15). These data suggest that ENaC may be involved in the regulation of sodium concentration in the endolymph (11,12).

Using a *Xenopus* oocyte expression system, we showed that TMPRSS3 is proteolytically cleaved and significantly activates ENaC. In addition, we showed that TMPRSS3 missense mutations causing deafness, that lie in all identified domains except the TM, and a catalytic triad S401A mutant, all fail to undergo proteolytic cleavage and to activate ENaC. The mechanism of ENaC activation by TMPRSS3 remains elusive and could be direct or indirect. TMPRSS3 anchored in the endoplasmic reticulum membranes may play a role in the processing of ENaC. ENaC is assembled and acquires core glycosylation in the endoplasmic reticulum; this subunit assembly occurs early in channel biosynthesis and is an important determinant of ENaC maturation (16).

All the TMPRSS3 pathogenic missense mutations described so far, D103G, R109W, C194F, W251C, P404L and C407R (Figs. 5A), affect highly conserved residues except R109W in the LDLRA domain and C407R in the serine protease domain (4–6). Based on our functional screening of TMPRSS3 mutants for their activation of ENaC in a *Xenopus* oocyte expression system, the mutations can be classified into three groups: (i) LDLRA domain mutants D103G and R109W which are partially active; (ii) an SRCR domain mutant, C194F, which

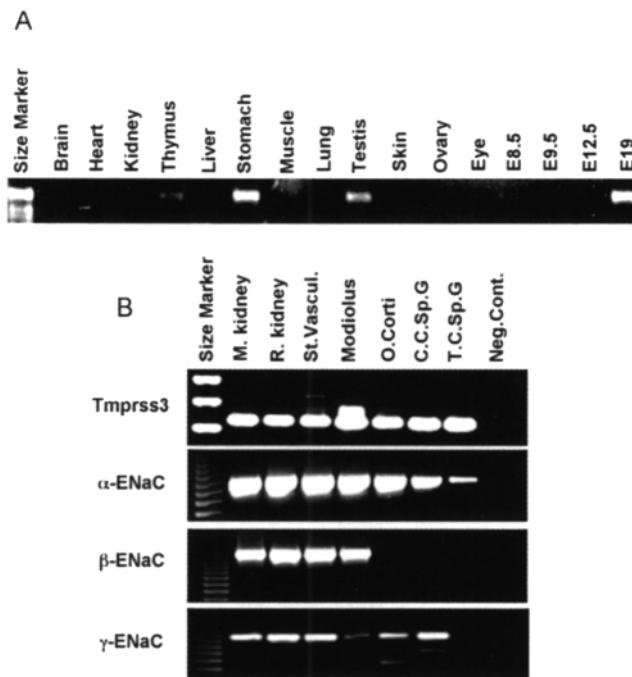


Figure 2. (A) RT-PCR analysis of mouse *Tmprss3* expression. (B) RT-PCR analysis of the expression of *Tmprss3*, ENaC α -, β - and γ -subunits in mouse (M.Kid) and rat kidney (R.Kid), stria vascularis (St.Vascul.), modiolus, organ of Corti (O.Corti), control cultured spiral ganglion (C.C.Sp.G) and BDNF-treated cultured spiral ganglion (T.C.Sp.G). Nested PCR amplification is presented for *Tmprss3* and α -ENaC subunit. First PCR amplification is presented for β - and γ -ENaC subunits.

changes a highly conserved residue and is totally inactive; and (iii) two subgroups of serine protease domain mutants, the first containing totally inactive mutants in which highly conserved residues are affected, W251C and P404L, while the second contains a partially active mutant, C407R, in which a conserved amino acid is substituted (Fig. 5A–D).

In order to investigate whether ENaC could be a substrate of TMPRSS3 in the inner ear, their respective expression profiles were determined and compared. In 2001, Grunder *et al.* (11) reported expression of the three ENaC subunits in the cochlea of 12-day-old rats by RNA *in situ* hybridization. α -, β - and γ -ENaC subunits were identified in the supporting cells of the organ of Corti, more specifically Deiters' and Claudius' cells. Only a faint staining in the Claudius' cells was observed for the β subunit. Expression of the three subunits was also detected in the spiral ganglion. However, none of the subunit was observed in the sensory hair cells and no significant expression of any of the three subunits was detected in the stria vascularis. In agreement with these data, our RT-PCR study on inner ear tissues of 5-day-old rats showed expression of the α - and γ -ENaC subunits in the organ of Corti and in the spiral ganglion and α , β and γ subunits in the stria vascularis. While β subunit expression was observed in the organ of Corti and in the spiral ganglion by *in situ* hybridization (11), our RT-PCR experiment did show expression of the β subunit in these tissues. This apparent discrepancy could be explained by the two different stages of development studied, 12-day-old versus 5-day-old rats, and/or by the fact that *in situ* hybridization is more

efficient at detecting expression in very few cells. Indeed, Grunder *et al.* (11) found consistently that the β subunit was only present in Claudius' cells and not in the Deiters' cells. In any case, from our data and those of Grunder *et al.* (11), one can propose that ENaC and TMPRSS3 co-localize in the spiral ganglion, in the stria vascularis and in the supporting cells of the organ of Corti. Interestingly, the stria vascularis and the organ of Corti are two cochlear tissues that border the endolymph and are implicated in sodium reabsorption (17–21).

ENaC is a heterotetrameric channel, $\alpha 2\beta \gamma$ but channel activity can also be observed in the presence of heterodimers $\alpha 2\beta 2$ or $\alpha 2\gamma 2$ (22,23). As far as the response to trypsin in the *Xenopus* oocyte expression system is concerned, we have found that trypsin can activate $\alpha 2\beta \gamma$ channels in a similar manner to the CAP proteins. Interestingly, $\alpha 2\beta 2$ complexes are not sensitive to trypsin or CAPs, whereas $\alpha 2\gamma 2$ complexes are highly sensitive to trypsin or CAPs. If the oocyte experiments are valid models for what happens in intact cells *in vivo*, one could speculate that in the cochlea, a relatively small amount of $\alpha 2\beta \gamma$ channels can be made. If the β subunit mRNA is limiting, the predominant form would be an $\alpha 2\gamma 2$ channel which keeps its trypsin/CAP sensitivity. Since both $\alpha 2,\beta,\gamma$ and $\alpha 2,\gamma 2$ can be activated by CAPs (B.C. Rossier, unpublished observations), it is reasonable to conclude that inner ear cells may express ENaC with distinct subunit composition and thus different sodium transport capacity but yet controlled by CAPs.

Little is known about the role of ENaC in the hearing process. In mice, the concentrations of all the ions in endolymph reach adult levels at day 7 after birth. In the earliest postnatal days, the endolymphatic sodium concentration is significantly higher than that in adult mice. α -ENaC ($-/-$), β -ENaC ($-/-$) and γ -ENaC ($-/-$) mice die within 40 h of birth (24–26). Therefore, they are not convenient models to study the role of ENaC in the development of the inner ear sodium homeostasis which occurs during the first week of life. To rescue cochlear hair cells from perinatal death of α -ENaC ($-/-$), organotypic cultures were performed. Stimulation of the outer hair cells in culture produced transducer currents that were not different from those observed in controls, suggesting a normal vestibular function and mechano-electrical transduction (27). The human ENaC genes have been found to be mutated in two distinct human diseases, Liddle's syndrome (OMIM#177200) and autosomal recessive pseudohypoaldosteronism type 1 (PHA-1) (OMIM#264350). In both conditions, the ENaC activity is dysregulated leading to disorders of systemic blood pressure (28). Until now, no severe hearing impairment has been reported in those patients but evaluation of less severe or selective impairment is currently underway in PHA-1 patients. ENaC may be only one of several sodium transporters involved in maintaining such an unusually low sodium concentration and some redundancy would be highly desirable in this context. In addition it is unlikely that ENaC is the only substrate for TMPRSS3 *in vivo*.

Besides the ion transporters discussed above, other substrates of TMPRSS3 in the inner ear could be neurotrophins. Studies over the past decade have provided evidence that brain-derived neurotrophic factor (BDNF) and neurotrophin 3 (NT-3) and their associated high-affinity tyrosine kinase receptors (TrkB and TrkC, respectively) are necessary for the normal development of the afferent innervation of the inner ear (29). The biological

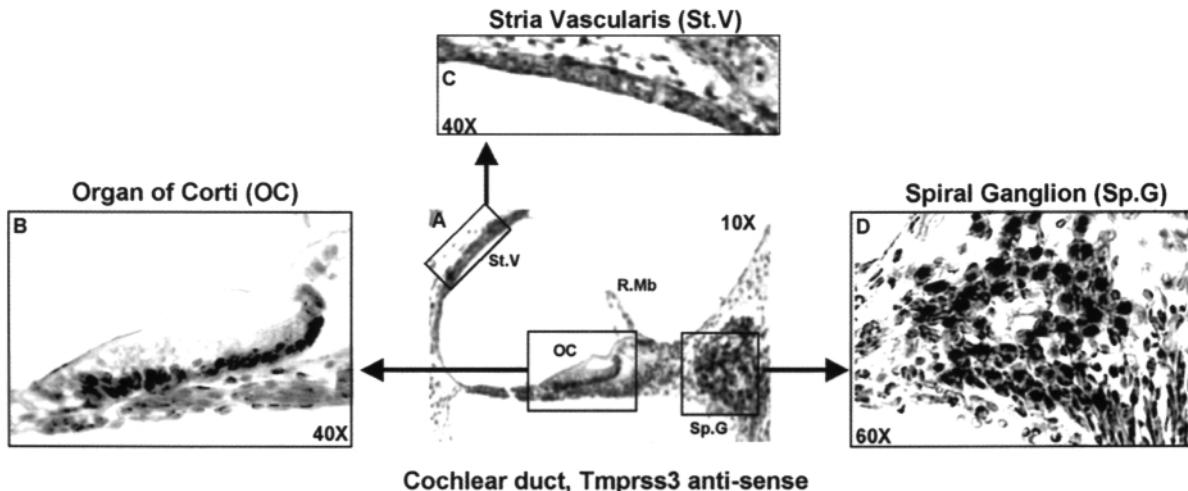


Figure 3. RNA *in situ* hybridization of mouse *Tmprss3* in cochlea of postnatal day-5-old mice. *Tmprss3* mRNA was detected in the supporting cells of the organ of Corti (panel A, magnification 10 \times and panel B, magnification 40 \times), in the stria vascularis (panel A, magnification 10 \times and panel C, magnification 40 \times) and in the spiral ganglion (panel A, magnification 10 \times and panel D, magnification 60 \times). St.V, stria vascularis; OC, organ of Corti; Sp.G, spiral ganglion; R.Mb, Reissner's membrane.

action of neurotrophins is regulated by proteolytic cleavage of a proform (proneurotrophins) to release the mature active neurotrophins. The serine protease plasmin, which belongs to the same protease family as TMPRSS3, has been shown to cleave the proform of BDNF (30). Neurotrophins, BDNF, NT-3 and their associated receptors, TrkB and TrkC, are all expressed in the organ of Corti and/or spiral ganglion (31–33). These data suggest that TMPRSS3 might also be involved in the processing of proneurotrophins and thus in the development and survival of inner ear innervation.

The results reported in this study contribute to our understanding of the role of TMPRSS3 in the inner ear and the possible etiology of DFNB8/10. We investigated the ENaC sodium channel as a potential substrate of TMPRSS3 in the inner ear focusing on the sodium homeostasis of the endolymph. Another hypothesis, not tested here, is that TMPRSS3 is involved in processing proneurotrophins and thus in the development and survival of the cochlear neurons. The generation of a mouse with targeted disruption of *Tmprss3* will provide a suitable model to investigate these and other hypotheses concerning the molecular mechanisms causing deafness in DFNB8/10.

MATERIALS AND METHODS

Isolation of the mouse *Tmprss3* cDNA and gene

A 220 kb mouse BAC clone, 556K2 (CITB-CJ7-B BAC library; Research Genetics, Paisley, UK), that spans the region of mouse chromosome 17q syntenic to the human *TMPRSS3* locus on human chromosome 21q22.3 has previously been reported (34). It contains the *Trefoil 3* gene, which in the human genome, is ~60 kb centromeric to *TMPRSS3*. A 1 kb fragment of the mouse *Tmprss3* was PCR-amplified from BAC 556K2 using degenerate primers (5'-TGYTGGAYNWSNGGNTGG-3', exon 9, codons 338–343; and 5'-CATYTGTCRTGDATCCAATC-3',

exon 11, codons 442–448 of *TMPRSS3*). The full-length *Tmprss3* cDNA was isolated by 5' and 3' RACE. *Tmprss3* introns were amplified from BAC 556K2 using primers based on predicted *Tmprss3* exons. Additional sequences were identified by BLASTN analyses of the Celera Mouse database (<http://www.celera.com/>). Sequences were assembled using the GAP4 Staden software (35). Sequences of the primers used are listed in http://medgen.unige.ch/public_1.html

Expression pattern of *Tmprss3*

RT-PCR. RT-PCR with primers 5'-CCTGGAGTCTATACTCGAATCAC-3' and 5'-ATTGGGTGGAGATCTGAGATCTG-3' located within exons 11 and 12 of *Tmprss3*, respectively, was performed on a normalized cDNA panel as described (36). Intron 11 of *Tmprss3* (3198 bp) is sufficiently large such that amplification from genomic DNA should yield either no PCR product or an easily distinguishable product.

RT-PCR was performed on RNA extracted from 5-day-old rat modiolus, stria vascularis, organ of Corti and cultured spiral ganglia cells, as described (37). PCRs for rat *Tmprss3*, were performed using primers based on mouse nucleotide sequence. PCRs for rat ENaC α -, β -, and γ -subunits were performed using primers based on rat nucleotide sequences (GenBank Nos. NM_031548.1, X77932.1 and U37540.1, respectively). Sequences of the primers used are listed in http://medgen.unige.ch/public_1.html

RNA *in situ* hybridization. A 1.4 kb fragment from the 3'UTR of mouse *Tmprss3* was amplified using primers 5'-CTGAAGTGAACAAGCCTGGAGTC-3' and 5'-TAGTCATCTCTTACTGTGACTAG-3' and cloned in TOPO TA PCRII vector (Invitrogen, Paisley, UK). Digoxigenin-labelled sense and anti-sense riboprobes were synthesized using T7 and SP6 RNA polymerases, respectively. Heads of 5 day-old mice were fixed

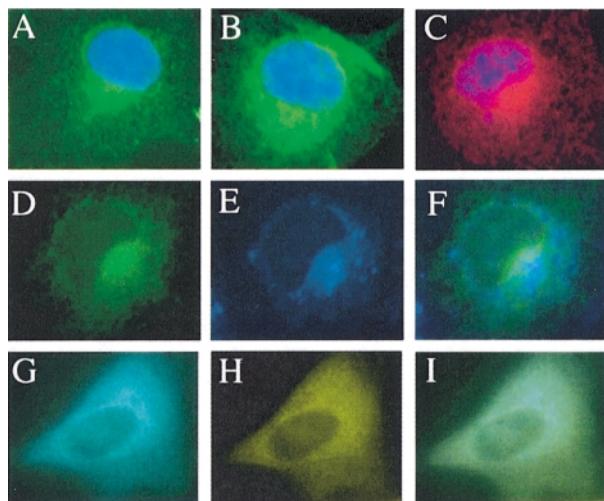


Figure 4. Intracellular localization of TMPRSS3 protein. Fluorescence microscopic analysis of fixed Cos-7 cells transiently transfected with pWT-TMPRSS3 (panel A, green signal), p5'EGFP-TMPRSS3 (panel B, green signal) and p5'HA-TMPRSS3 (panel C, red signal) and counterstained with 4,6-diamidino-2-phenylindole (DAPI) (panels A, B and C, blue signal). Fixed Cos-7 cells transiently transfected with pEGFP-TMPRSS3 (panel D, green signal) and counterstained with endoplasmic reticulum marker (ER-Tracker, panel E, blue signal). Co-localisation of EGFP-TMPRSS3 with ER marker is shown in panel F. Fixed HeLa cells transiently co-transfected with pTMPRSS3-3'/ECFP and pYFP-ER (panels G–I). TMPRSS3-ECFP signal is shown in panel G. pYFP signal is shown in panel H. Co-localisation of TMPRSS3-ECFP with the ER marker is shown in panel I.

in 4% paraformaldehyde, washed in PBS, dehydrated and paraffin embedded. Sections, 8 µm thick, were mounted onto super-frost slides. Slides were deparaffinized, rehydrated, acetylated (0.25% acetic anhydride, 45 mM NaCl, 100 mM Tris, pH 8) and washed in PBS. *In situ* hybridization was performed overnight at 58°C in 40% formamide, 5× SSC, 1× Denhardt's, 100 µg/ml tRNA and the digoxigenin-labelled probe. After washing once with 2× SSC at room temperature, twice with 2× SSC and twice with 0.1× SSC at 58°C, sections were equilibrated with Triton washing buffer (TWB), (0.05% Triton X-100, 150 mM NaCl, 100 mM Tris, pH 7.5). Non specific binding sites were blocked with TWB containing 0.5% blocking reagent (Roche, Rotkreuz, Switzerland) and sections were incubated with anti-digoxigenin antibody for 2 h. After two washing steps with TWB and equilibration with alkaline phosphatase buffer NTMT (100 mM NaCl, 50 mM MgCl₂, 100 mM Tris, pH 9.5, 0.1% Tween 20), the slides were incubated with the substrate (NBT/BCIP) overnight at room temperature. The reaction was stopped in TE buffer (pH 8.5). Sections were dehydrated and mounted. Images were recorded with a Hamamatsu digital camera.

cDNA constructs. To visualize the subcellular localization of TMPRSS3, four plasmids that express either tagged or wild-type TMPRSS3 were constructed: WT-TMPRSS3 was PCR-amplified and subcloned into vectors pECFP-N1 (Clontech, Palo Alto, California), pcDNA3X(+)-MycEGFP, pcDNA3X(+)-HA and pcDNA3.1 (Invitrogen). For expression studies, WT-TMPRSS3 was subcloned into pSD5 expression

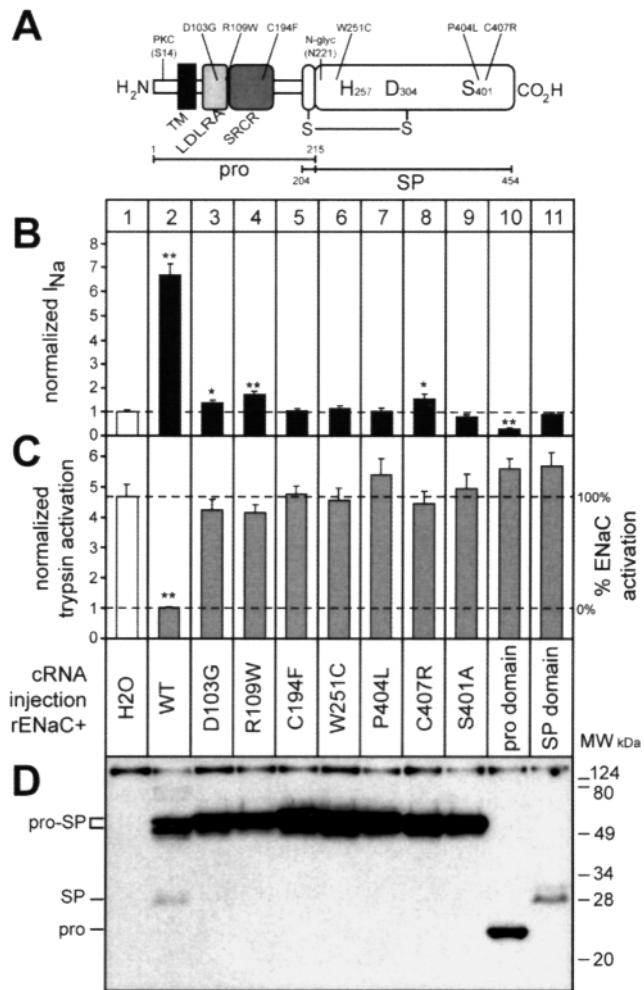


Figure 5. Functional expression of TMPRSS3 in *Xenopus* oocytes. (A) Predicted structure of TMPRSS3. Numbers indicate the amino acid positions. The serine protease catalytic triad residues (H257, D304 and S401), the predicted disulfide bridge which links the pro- and catalytic domains of TMPRSS3 and the tested human deafness associated TMPRSS3 missense mutations D103G, R109W, C194F, W251C, P404L and C407R are shown. TM, transmembrane domain; LDLRA, low-density lipoprotein receptor class A domain; SRCR, scavenger receptor cysteine-rich domain; pro, pro-domain; SP, serine protease domain. (B) Comparison of the effect of WT-TMPRSS3 and mutants on I_{Na} in *Xenopus* oocytes. Oocytes were injected with rat ENaC subunits in the presence of either water (lane 1, white column), WT-TMPRSS3 (lane 2), TMPRSS3 missense deafness associated mutants (lanes 3–8), serine catalytic mutant (lane 9), pro domain construct (lane 10) or serine protease domain construct (SP) of TMPRSS3 (black columns). ** = $P < 0.01$; * = $P < 0.05$ versus 1. Lanes 1–9 and 11, ($n \geq 25$); lane 10, ($n = 16$). (C) Measure of the fold increase of ENaC-mediated I_{Na} in injected oocytes after 3 min of perfusion of trypsin. Same conditions as described under B. ** = $P < 0.01$. Lanes 1–9 and 11, ($n \geq 16$); lane 10, ($n = 10$). (D) Biochemical analysis of WT and mutants TMPRSS3 by western blot on protein extracts from pools of 10 injected oocytes. Pro-SP, pro-serine protease; pro, pro-domain; SP, serine protease domain. Same conditions as described under B.

vector (9). Mutants (D103G, R109W, C194F, W251C, P404L, C407R, S401A) were generated using the QuickChange mutagenesis kit (Stratagene, La Jolla, CA, USA). Two truncated TMPRSS3 constructs (pro domain M1–S215: TMPRSS3 without the catalytic domain and SP

domain M-C204-T454: TMPRSS3 catalytic domain alone) were generated by PCR and subcloned into PSD5 vector (Fig. 5A). All constructs were verified by DNA sequencing. Sequences of the primers and restriction enzymes used are listed in http://medgen.unige.ch/public_1.html

Human TMPRSS3 polyclonal antibodies. Two polyclonal antibodies were raised in rabbits against a mixture of two synthetic peptides NH₂-MGENDPPAVEAPFSFC-COOH (amino acids 1–16 of human TMPRSS3) and NH₂-CFLDWI-HEQMERDLKT-COOH (amino acids 439–454 of human TMPRSS3) (CovalAb, Lyon, France, <http://www.covalab.com>).

Subcellular localization of TMPRSS3 protein. Cos-7, U2OS and MDCK cells were transfected with 0.5 µg of 5' MycEGFP-TMPRSS3 or 5' HA-TMPRSS3 or WT-TMPRSS3 using LipofectAMINE PLUS reagent (Life Technologies, Paisley, UK, #10964-013) and fixed in 4% paraformaldehyde. 5' MycEGFP-TMPRSS3 fluorescence was detected using epifluorescence microscopy (Zeiss Axiophot I, color Axiocam). 5' HA-TMPRSS3 was detected using anti-HA antibody (Santa Cruz Biotechnology, Santa Cruz, CA, USA, #sc7392) followed by a rhodamine-conjugated secondary antibodies (Santa Cruz Biotechnology, #sc-2093). WT-TMPRSS3 was detected using rabbit anti-TMPRSS3 serum (CovalAb) followed by fluorescein isothiocyanate-conjugated secondary antibodies (Santa Cruz Biotechnology, #sc-2090). Images were processed with the Adobe Photoshop software.

To perform co-localization experiments, transfected cells were incubated with markers for the Golgi apparatus (mouse anti-Giantin antibody, Calbiochem, San Diego, CA, USA, #324450) followed by rhodamine-conjugated secondary antibody (Santa Cruz #sc-2093) or the endoplasmic reticulum (ER-Tracker Blue-White DPX, Molecular Probes, Eugene, OR, USA, #E-2353). Alternatively, HeLa and Cos-7 cells were cotransfected with 1 µg of pTMPRSS3-ECFP-N1 and marker vectors including pEYFP-Peroxi, pEYFP-Mito, pEYFP-Golgi, pEYFP-ER, or pEYFP-Mem (Clontech) using Fugene 6 (Roche). Empty expression vectors were used as negative controls.

Electrophysiological measurements in *Xenopus* oocytes. Expression studies were performed in stage V/VI oocytes isolated from *Xenopus laevis* (Noerdhoek, South Africa). Oocytes were injected with 0.25 ng of each cRNA coding for the rat α-, β- and γ-ENaC subunits in the presence or absence of 2.5 ng of wild-type or mutant TMPRSS3 cRNA in a total volume of 100 nL. Oocytes were incubated in modified Barth saline (MBS) solution and 48 h after injection, electrophysiological measurements were performed. Using the two-electrode voltage clamp technique, the amiloride-sensitive current (I_{Na}) was measured in the presence of 120 mM of Na⁺ in Frog Ringer with 5 µM amiloride at a holding potential of -100 mV. The oocytes were perfused with 2 µg/ml of trypsin during 2–3 min and I_{Na} was re-measured. All results are reported as means ± SEM. Comparing independent sets of data, unpaired *t*-tests were used

to determine significance. In experiments where oocytes were perfused with trypsin, paired *t*-tests were used. (n) Represents the number of experiments performed.

Western blot analysis. Injected oocytes were incubated in MBS for 48 h. Microsomal membrane protein extracts were obtained from pools of 10 injected oocytes lysed as described (38). Proteins were separated on a 10% SDS-polyacrylamide gel under reducing and denaturing conditions and transferred to nitrocellulose membrane. The membrane was processed using rabbit anti-TMPRSS3 serum (Covalab) according to standard procedures.

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