LIN-39 and the EGFR/RAS/MAPK pathway regulate *C. elegans* vulval morphogenesis via the VAB-23 zinc finger protein

Mark W. Pellegrino1,2,*; Sarfaraz Hussain Farooqui1,3; Erika Fröhlili2; Hubert Rehrauer4; Stéphanie Kaeser-Pebernard5; Fritz Müller5; Robin B. Gasser1,† and Alex Hajnal2,†

**SUMMARY**

Morphogenesis represents a phase of development during which cell fates are executed. The conserved hox genes are key cell fate determinants during metazoan development, but their role in controlling organ morphogenesis is less understood. Here, we show that the *C. elegans* hox gene *lin-39* regulates epidermal morphogenesis via its novel target, the essential zinc finger protein VAB-23. During the development of the vulva, the egg-laying organ of the hermaphrodite, the EGFR/RAS/MAPK signaling pathway activates, together with LIN-39 HOX, the expression of VAB-23 in the primary cell lineage to control the formation of the seven vulval toroids. VAB-23 regulates the formation of homotypic contacts between contralateral pairs of cells with the same sub-fates at the vulval midline by inducing *smp-1* (*semaforin*) transcription. In addition, VAB-23 prevents ectopic vulval cell fusions by negatively regulating expression of the fusogen *eff-1*. Thus, LIN-39 and the EGFR/RAS/MAPK signaling pathway, which specify cell fates earlier during vulval induction, continue to act during the subsequent phase of cell fate execution by regulating various aspects of epidermal morphogenesis. Vulval cell fate specification and execution are, therefore, tightly coupled processes.

**KEY WORDS:** Morphogenesis, hox, ras, *C. elegans*, Vulva

**INTRODUCTION**

Morphogenesis is the phase in development that occurs after cells have adopted their cell fates. A pivotal condition for proper morphogenesis is the appropriate specification of cell fates by the hox genes, which encode homeodomain-containing transcription factors that typically control anterior-posterior cell fate decisions during embryogenesis and the axial patterning of the limbs (Hombria and Lovegrove, 2003; McGinnis and Krumlauf, 1992).

The nematode *Caenorhabditis elegans*, which contains a single non-redundant hox gene cluster, has been used extensively to characterize hox gene functions. Six *C. elegans* hox genes have been identified: the *labial*-like gene *celh-13* (Brunschwig et al., 1999); the *Antp* class genes *lin-39* (Clark et al., 1993) and *mab-5* (Kenyon, 1986); and the posterior *Abd-B*-like genes *egf-5*, *nob-1* and *php-3* (Chisholm, 1991; Van Auker et al., 2000). A central role for *lin-39* and its co-factors has been observed during vulval development in the hermaphrodite larva (Clark et al., 1993; Yang et al., 2005). The vulva forms a connection between the uterus and the outside, through which the fertilized eggs are laid. Composed of only 22 cells generated in a relatively simple and invariant cell lineage, the vulva serves as an excellent model to study cell fate determination (Sternberg, 2005). During the second larval stage, three out of six equipotent vulcal precursor cells (VPCs) are induced by an epidermal growth factor (EGF) signal from the uterine anchor cell (AC) to adopt a vulcal cell fate (Hill and Sternberg, 1992). The EGF receptor, encoded by *let-23*, transduces the inductive signal in the VPCs via the RAS/MAPK pathway (Aroian and Sternberg, 1991; Beitel et al., 1990; Lackner and Kim, 1998). In response to the inductive signal, a lateral signal from P6.p activates LIN-12 NOTCH in P5.p and P7.p, which represses EGFR/RAS/MAPK signaling in these cells (Berset et al., 2001; Greenwald, 2005). Thus, high MAPK activity in P6.p results in the primary (1°) fate, whereas strong LIN-12 signaling in P5.p and P7.p specifies the secondary (2°) fate (Greenwald, 2005). Once the cell fates have been determined, P5.p-P7.p undergo three rounds of divisions to generate 22 vulval cells that are further divided into seven sub-fates. VulA, VulB1, VulB2, VulC and VulD are generated by the seven 2° descendants each of P5.p and P7.p, and VulE and VulF are formed by the eight 1° descendants of P6.p (Fig. 1A) (Shemer et al., 2000). During the subsequent phase of morphogenesis, cells migrate and form extensions towards the vulval midline, where they make homotypic contacts with their contralateral partner cells of the same sub-fates, thereby forming seven symmetric rings called toroids. With the exception of VulB1 and VulB2, cells within a toroid fuse, thus generating a stack of syncytial toroids arranged in a pyramidal manner.

LIN-39, which is activated through the combined actions of the EGFR/RAS/MAPK and WNT signaling pathways, performs at least two distinct functions during vulval development (Eisenmann et al., 1998; Maloof and Kenyon, 1998). First, LIN-39 prevents the fusion of the VPCs with hyp7 at the early L2 stage (Clark et al., 1993). Second, during cell fate execution and morphogenesis, LIN-39 is required for cell proliferation and toroid formation (Shemer and Podbielweicz, 2002). The HOX co-factors CEH-20 PXB and UNC-62 MEIS play similar roles during vulval morphogenesis by regulating cell fusion and migration (Yang et al., 2005). Previous work has indicated that *let-60 ras* is also involved in controlling vulval morphogenesis (Shemer et al., 2000), though the exact role of EGFR/RAS/MAPK signaling during vulval cell fate execution is not well understood.
We have previously identified the \textit{vab-23} gene as an essential regulator of morphogenesis that controls the movements of ventral epidermal cells in the embryo (Pellegrino et al., 2009). \textit{VAB-23} is the first characterized member of a family of nuclear proteins containing a strongly conserved C4H2-type zinc finger domain at the C-terminus. All metazoans genomes encode a single \textit{VAB-23} homolog, and the human protein HCA127 (ZC4H2 – Human Gene Nomenclature Database) was found to be an auto-antigen in hepatocellular carcinomas (Wang et al., 2002). Here, we describe the functions of \textit{VAB-23} during vulval morphogenesis. The LIN-39 and RAS/MAPK target \textit{VAB-23} guides cell interactions during toroid formation. \textit{VAB-23} positively regulates, among others, the transcription of \textit{smp-1} to allow proper interactions to occur between cells of the same sub-fates. In addition, \textit{VAB-23} inhibits EFF-1-dependent cell fusion during toroid formation. Thus, \textit{vab-23} is an essential factor linking LIN-39 and the EGFR/RAS/MAPK pathway, which are first used during vulval fate specification, to the subsequent phase of morphogenesis.

**MATERIALS AND METHODS**

**Strains**

\textit{C. elegans} strains were maintained at 20°C as described (Brenner, 1974) unless noted otherwise. Wild type refers to the Bristol N2 strain. Transgenic lines were generated by microinjection of the indicated constructs at 10-20 ng/\textmu l together with transformation markers at 50-75 ng/\textmu l and blueprints carrier to a total DNA concentration of 150 ng/\textmu l. Strains used are as follows: LGI: \textit{vab-23}(tm1945)/\textit{mIs1}[\textit{mdp-14::gfp},\textit{unc-119(+)}] (this study), \textit{lin-7(e1413)}, \textit{eff-1(ok1021)}. LGII: \textit{lin-39(n1760)}, \textit{LGIV: lin-3(e1417)}, \textit{let-60(e1046)}, \textit{LGV: sos-1(cce41)}. Integrated arrays: LGII: \textit{kds36[P\textit{gfp-226::gfp},\textit{unc-119(+)}]/H11033} (Hanna-Rose and Han, 2002). LGIII: \textit{syIs90[P\textit{gfp-27::yfp},\textit{unc-119}]} (Mohler et al., 1998), \textit{zhIs1[lin-39::gfp]} (Szabo et al., 2009), \textit{swIs79[ajm-1::gfp,\textit{unc-119}], mels46[\textit{dig-1::dsred,unc-119(+)}], LGIV: \textit{syb51[P\textit{gfp-22::gfp},\textit{unc-119}]} (Inoue et al., 2002), \textit{gals36[\textit{ls-mpk-1(+),D-meck[\textit{gfp}]}]} (Lackner and Kim, 1998), \textit{zzIs16[P\textit{gfp-1::gfp,rol-6(qf)}]} (Fernandes and Sternberg, 2007). Extra-chromosomal arrays: \textit{zhEx229[vab-23::gfp,\textit{lin-48::gfp}]} (Pellegrino et al., 2009), \textit{zhEx271[P\textit{gfp-22::gfp},\textit{lin-48::gfp}]} (Pellegrino et al., 2009), \textit{zhEx275[vab-23::gfp,\textit{rol-6(d)}]} (Lackner and Kim, 1998), \textit{zhEx277[vab-23::gfp,\textit{lin-48::gfp}]} (Pellegrino et al., 2009), \textit{zhEx287[vab-23::gfp,\textit{lin-48::gfp}]} (Lackner and Kim, 1998), \textit{zhEx294[vab-23::gfp,\textit{lin-48::gfp}]} (Pellegrino et al., 2009), \textit{zhEx295[vab-23::gfp,\textit{lin-48::gfp}]} (Pellegrino et al., 2009), \textit{zhEx317[P\textit{unc-119::vab-23a::gfp}]} (Pellegrino et al., 2009). Integrated arrays: LGI: \textit{mIs346[plin-3::\textit{gfp},\textit{unc-119}]} (Pellegrino et al., 2009), \textit{zhEx267[\textit{psmp-1-2004-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (this study), \textit{Psmp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]} (Pellegrino et al., 2009), \textit{zhEx372[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx373[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx374[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx375[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx376[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx377[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx378[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx388[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx389[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx390[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx391[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx392[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx393[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx394[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx395[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx396[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx397[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx398[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx399[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009), \textit{zhEx400[P\textit{omp-1-2256-2987::nls::gfp::lacZ,\textit{myo-2::mcherry}]}]} (Pellegrino et al., 2009).
RNA interference

RNA interference (RNAi) was performed using the feeding method (Kamath et al., 2001). Worms were synchronized at the L1 stage, transferred to nematode growth media plates containing 3 mM IPTG, 50 µg/ml ampicillin and RNAi bacteria, and allowed to grow for 3-5 days at 25°C. The F1 progeny were analyzed.

Microscopy and 3D reconstructions of vulval toroids

Images were recorded of anesthetized larvae using a Leica DMRA wide-field microscope equipped with a cooled CCD camera (Hamamatsu ORCA-ER) as described (Pellegrino et al., 2009) and analyzed using the Openlab 5.0 software package (Improvement/Perkin Elmer). To quantify signal intensities (Figs 5, 6), images were recorded under identical conditions and the mean fluorescence intensity subtracted by the background fluorescence was determined using the Openlab measurement tool. To create 3D reconstructions of vulval toroids (Figs 1, 7), confocal sections through the larvae were recorded with an Olympus FV1000 confocal microscope and processed with the Velocity 2.0 software package (Improvement/Perkin Elmer).

vab-23 and smp-1 promoter and chromatin immunoprecipitation (ChIP) analysis

vab-23 promoter deletion analysis was performed by PCR using the vab-23::gfp plasmid as template. The upstream region of the vab-23 promoter for each PCR product is as follows: 3372 bp for vab-23Hox(+), 1363 bp for vab-23HoxΔ1, 1203 bp for vab-23HoxΔ2. Point mutations in both conserved PBX sites (vab-23 Hox mut) were introduced using a combination of the QuickChange Multi Site-Directed Mutagenesis Kit (Stratagene) and overlap PCR (Hobert, 2002). smp-1 promoter analysis was performed by PCR amplification of the fragments indicated in Fig. 4E subcloned into the Psil and Ssil sites of the Fire vector pPD96.04.

For ChIP analysis, chromatin prepared from animals carrying a functional LIN-39::GFP reporter was precipitated with GFP antibodies (Roche) as described by Mukhopadhyay et al. (Mukhopadhyay et al., 2008). As negative control, a mock precipitation omitting the primary antibody was performed in parallel. Binding was quantified by Q-PCR with the probes shown in Fig. 6B. For each sample, the signal was first normalized to the input DNA (Δact) and then to the signal obtained with a probe in the 3’UTR of vab-23 (ΔΔact) as internal reference to calculate relative binding. Primers used for Q-PCR analysis were: CTCAAATGTTGCTCCAAAAT and AGAGAGAGAAGAGAGAGAG for region I; TACTCTCCTCTTGGCCT and CGGACCATGATCCAA-ACAC for region II; CCCCATAGACATTCTTAC and GACAAGATGT-ATTCCGGC for region III; and CAATTTCTTGTGCATTTCC and TCCCTATACATCCGCA for the 3’ region. For ChIPseq analysis, 20 ng of immunoprecipitated DNA was used as template for the SOLID 4 whole-genome sequencing platform yielding around 4×10^6 reads of 50 bp each. DNAs isolated from the input chromatin fraction and from a ChIP experiment using an unrelated GFP fusion protein were subjected to the same sequence analysis and used as background controls. Sequence alignment to the C. elegans reference sequence (WS192) was performed with the CLC genomics workbench software using default parameters. Peak detection was performed with a custom R script described in the legend to Table S1 in the supplementary material.

RESULTS VAB-23 is essential for toroid formation during vulval morphogenesis

The function of vab-23 was analyzed using the vab-23(tm1945) deletion allele (kindly provided by S. Mitani, Tokyo Women’s University School of Medicine, Japan) that is likely to represent a null mutation (Pellegrino et al., 2009). The vab-23(tm1945) deletion results in a completely penetrant embryonic or early larval lethal phenotype due to defects in epidermal morphogenesis. The lethality could be rescued by expression of bova-23a cDNA with gfp fused at the C-terminus under control of the pan-neuronal unc-119 promoter (Punc-119::vab-23::gfp) (Pellegrino et al., 2009). This neuron-specific rescue allowed us to study the function of vab-23 during post-embryonic development. Homozygous vab-23(tm1945) mutants carrying the Punc-119::vab-23::gfp transgene developed to fertile adults with a completely penetrant protruding vulva (Pvl) phenotype. At low penetrance (<15%), Punc-119::vab-23::gfp was weakly expressed in the 1° vulval lineage, which we attribute to the leaky nature of the unc-119 promoter. However, the low level of epidermal vab-23 expression was insufficient to rescue the Pvl phenotype. For the remainder of this study, we refer to vab-23(tm1945) larvae rescued by Punc-119::vab-23::gfp as vab-23(tm1945) mutants.

We first investigated whether the vulval defects of vab-23(tm1945) were caused by an abnormal pattern of cell division. For this purpose, we directly followed the vulval lineage of vab-23(tm1945) animals, but observed no change in the cell division pattern or the orientations of the cleavage axes (n=4). In addition, we counted the number of vulval cells at the early L4 stage and found the normal number of 22 cells in all cases (n=20; data not shown). However, all vab-23(tm1945) L4 larvae displayed penetrant defects in vulval morphogenesis. The wild-type vulva is made of a stack of seven concentric toroid rings that are formed by the circumferential migration and subsequent fusion of contralateral pairs of cells of the same sub-fates (Fig. 1A,B). By contrast, the vulvae of vab-23(tm1945) larvae were asymmetric in configuration and abnormal in shape (Fig. 1C). These defects could be phenocopied by vab-23 RNAi (see Fig. S1 in the supplementary material) and rescued by introducing the entire genomic vab-23 locus into vab-23(tm1945) mutants (Fig. 8A, in this particular experiment in the absence of Punc-119::vab-23::gfp). Thus, the observed vulval morphogenesis defects are caused by a loss of vab-23 function.

Next, we used the AJM-1::GFP adherens junction marker to examine the shape of the vulval toroids in L4 larvae (Mohler et al., 1998). In the wild type, AJM-1::GFP appeared as a stack of eight concentric rings delineating the seven toroids (Fig. 1D). A mid-sagittal cross-section revealed eight dots representing the cell junctions of the toroids (Fig. 1D'). vab-23(tm1945) animals displayed two characteristic defects. First, fewer abnormally shaped toroids were formed with an average of 4.3±0.2 vulval toroid junctions per animal (n=43) (Fig. 2B). We then investigated whether the vulval defects of vab-23(tm1945) mutants might be due to ectopic cell fusions resulting from inter-toroid fusion between cells of different sub-fates or from fusion with the surrounding hyp? The fusogen EFF-1 is necessary for most somatic cell fusions, including those between vulval cells and hyp7 and between vulval cells within toroids (Mohler et al., 2002). We used a transcriptional Peff-1::gfp reporter to examine whether ectopic eff-1 expression might be responsible for the loss of toroids. In control RNAi animals, Peff-1::gfp was expressed at the L4 stage in VulA, VulC and less frequently in VulF (21%, n=43) (Fig. 2A). Reduction of vab-23 function by RNAi caused an increased expression of Peff-1::gfp in VulF (65%, n=34) and additional expression in VulE (38%, n=34), where no expression was observed in control animals (n=43) (Fig. 2B). We then investigated whether loss of eff-1 function suppresses the reduced toroid number in vab-
23(tm1945) mutants. Vulval morphogenesis was still abnormal and toroids were malformed in eff-1(ok1021); vab-23(tm1945) double mutants, although the number of cell junctions was increased from 4.3±0.2 to an average of 6.4±0.2 per animal (n=24; Fig. 1F). Taken together, loss of VAB-23 function results in the ectopic expression of EFF-1, which is partially responsible for the reduced toroid number. However, as the toroids that formed in eff-1(ok1021); vab-23(tm1945) double mutants were still abnormally shaped, VAB-23 is not only required to repress EFF-1-mediated vulval cell fusions but also to mediate proper toroid formation.

**VAB-23 represses the formation of ectopic cell contacts**

VAB-23 regulates the formation of contacts between contralateral pairs of epidermal cells during morphogenesis of the embryo (Pellegrino et al., 2009). To assess whether VAB-23 plays a similar function during vulval morphogenesis, we followed cell junctions of EFF-1, which is partially responsible for the reduced toroid expression at the early L3 stage (Pn.p to Pn.px stage, n=33). However, beginning at the Pn.pxx stage, abnormal cell extensions between adjacent vulval cells formed in 59% of vab-23(tm1945) animals (n=41; Fig. 3D), and at the onset of vulval invagination (Pn.pxxx stage) ectopic contacts between 1° and 2° vulval cells were observed in 88% of the cases (n=33; Fig. 3F). Finally, in eff-1(ok1021); vab-23(tm1945) double mutants at the L4 stage, 1° and 2° cells failed to reach their contralateral partner cells across the midline (Fig. 1F), suggesting that the vulval morphogenesis defects are similar to the defects observed during ventral epidermal enclosure of the embryo (Pellegrino et al., 2009). Thus, besides repressing cell fusions, VAB-23 is also required for the proper interaction between vulval cells of the same sub-fates.

**VAB-23 regulates smp-1 transcription during vulval morphogenesis**

The abnormal vulval cell migrations and cell contacts in vab-23(tm1945) mutants are reminiscent of mutations in smp-1, which encodes a transmembrane Semaphorin IA homolog, or in plx-1, which encodes the putative SMP-1 receptor Plexin-A4 (Dalpe et al., 2005). We therefore examined whether smp-1 and/or plx-1 expression might be regulated by VAB-23, as we previously found that the zinc finger domain of VAB-23 is essential for its nuclear function (Pellegrino et al., 2009). Using a translational SMP-1::GFP reporter (kind gift of Joseph Culotti, The Samuel Lunenfeld Research Institute, Ontario, Canada), for which expression and localization at the vulval midline has been previously described (Dalpe et al., 2005), we found that 59% of vab-23(tm1945) larvae (n=29) lacked SMP-1::GFP expression at the onset of vulval invagination, whereas SMP-1::GFP expression was absent in only 9% of wild-type animals at this stage (n=34) (Fig. 4A, B). We verified that the loss of SMP-1 expression was not due to excess cell fusions by analyzing the adherens junction marker DLG-1::dsRED (Fig. 4A, B) (Bossinger et al., 2001). Despite the abnormal shape of the vulval invagination, no reduction in DLG-1::dsRED expression was detected in vab-23(tm1945) mutants at this stage, indicating that the ectopic cell fusions occur at a later stage and are therefore unlikely to be the cause of the loss SMP-1::GFP expression.

To determine whether VAB-23 controls smp-1 expression at the transcriptional or post-transcriptional level and to identify the cis-regulatory elements, we generated a series of transcriptional smp-1 reporters (Fig. 4E). A construct in which an nls::gfp::lacZ reporter cassette was fused in frame within exon II to include 2256 bb of the 5′ smp-1 promoter/enhancer region and the entire intron I yielded specific expression in the VulF, VulE and VulD cells (P_smp-1-2256-1/H11032 and P_smp-1-2256-2987/H11033) (Bossinger et al., 2001). A construct consisting only of the 2256 bb of the 5′ promoter/enhancer region upstream of exon I did not show any detectable expression in the vulval cells but showed strong ectopic expression in body wall muscles and hypodermal cells (P_smp-1-2256-1/H11032 in Fig. 4E and data not shown). However, a construct consisting only of the 2256 bb of the 5′ promoter/enhancer region upstream of exon I did not show any detectable expression in the vulval cells but showed strong ectopic expression in body wall muscles and hypodermal cells (P_smp-1-2256-1/H11032 in Fig. 4E and data not shown). Thus, intron I must contain the regulatory elements required for vulva-specific transcription of smp-1 mRNA. We further narrowed down the region that is sufficient for vulval expression to a 493 bp fragment at the end of intron I (Fig. 4C, C′ and P_smp-1-2494-2987/H11033 in 4E). When the P_smp-1-2494-2987 reporter was introduced into the vab-23(tm1945) mutant background, expression was strongly reduced but not eliminated (Fig. 4D). Specifically, 30% (n=30) of vab-23(tm1945) L4 larvae showed detectable NLS::GFP expression in the vulval cells, whereas strong expression was observed in 85% (n=20) of wild-type larvae. Further deletion analysis identified a 102 bp region (between positions 2701 and

---

Fig. 2. VAB-23 represses the fusogen eff-1 in VulE and VulF. (A-B) Nomarski (A,B), GFP (A’,B’) and merged (A’’,B’’) images of control (A-A’’) and vab-23 RNAi-treated (B-B’’) larvae carrying an eff-1::gfp reporter. The positions of the VulE and VulF nuclei are indicated by arrows. Anterior is left. Scale bar: 10 μm.

---

Fig. 3. Cell contacts in adjacent vulval cells formed in 59% of wild-type animals at the Pn.pxx stage, abnormal cell extensions between 1° and 2° vulval cells were observed in 88% of the cases (n=33; Fig. 3F). Finally, in eff-1(ok1021); vab-23(tm1945) double mutants at the L4 stage, 1° and 2° cells failed to reach their contralateral partner cells across the midline (Fig. 1F), suggesting that the vulval morphogenesis defects are similar to the defects observed during ventral epidermal enclosure of the embryo (Pellegrino et al., 2009). Thus, besides repressing cell fusions, VAB-23 is also required for the proper interaction between vulval cells of the same sub-fates.

Fig. 4. VAB-23 regulates smp-1 transcription during vulval morphogenesis. The abnormal vulval cell migrations and cell contacts in vab-23(tm1945) mutants are reminiscent of mutations in smp-1, which encodes a transmembrane Semaphorin IA homolog, or in plx-1, which encodes the putative SMP-1 receptor Plexin-A4 (Dalpe et al., 2005). We therefore examined whether smp-1 and/or plx-1 expression might be regulated by VAB-23, as we previously found that the zinc finger domain of VAB-23 is essential for its nuclear function (Pellegrino et al., 2009). Using a translational SMP-1::GFP reporter (kind gift of Joseph Culotti, The Samuel Lunenfeld Research Institute, Ontario, Canada), for which expression and localization at the vulval midline has been previously described (Dalpe et al., 2005), we found that 59% of vab-23(tm1945) larvae (n=29) lacked SMP-1::GFP expression at the onset of vulval invagination, whereas SMP-1::GFP expression was absent in only 9% of wild-type animals at this stage (n=34) (Fig. 4A, B). We verified that the loss of SMP-1 expression was not due to excess cell fusions by analyzing the adherens junction marker DLG-1::dsRED (Fig. 4A, B) (Bossinger et al., 2001). Despite the abnormal shape of the vulval invagination, no reduction in DLG-1::dsRED expression was detected in vab-23(tm1945) mutants at this stage, indicating that the ectopic cell fusions occur at a later stage and are therefore unlikely to be the cause of the lost SMP-1::GFP expression.

To determine whether VAB-23 controls smp-1 expression at the transcriptional or post-transcriptional level and to identify the cis-regulatory elements, we generated a series of transcriptional smp-1 reporters (Fig. 4E). A construct in which an nls::gfp::lacZ reporter cassette was fused in frame within exon II to include 2256 bb of the 5′ smp-1 promoter/enhancer region and the entire intron I yielded specific expression in the VulF, VulE and VulD cells (P_smp-1-2256-1/H11032 and P_smp-1-2256-2987/H11033) (Bossinger et al., 2001). A construct consisting only of the 2256 bb of the 5′ promoter/enhancer region upstream of exon I did not show any detectable expression in the vulval cells but showed strong ectopic expression in body wall muscles and hypodermal cells (P_smp-1-2256-1/H11032 in Fig. 4E and data not shown). However, a construct consisting only of the 2256 bb of the 5′ promoter/enhancer region upstream of exon I did not show any detectable expression in the vulval cells but showed strong ectopic expression in body wall muscles and hypodermal cells (P_smp-1-2256-1/H11032 in Fig. 4E and data not shown). Thus, intron I must contain the regulatory elements required for vulva-specific transcription of smp-1 mRNA. We further narrowed down the region that is sufficient for vulval expression to a 493 bp fragment at the end of intron I (Fig. 4C, C′ and P_smp-1-2494-2987/H11033 in 4E). When the P_smp-1-2494-2987 reporter was introduced into the vab-23(tm1945) mutant background, expression was strongly reduced but not eliminated (Fig. 4D). Specifically, 30% (n=30) of vab-23(tm1945) L4 larvae showed detectable NLS::GFP expression in the vulval cells, whereas strong expression was observed in 85% (n=20) of wild-type larvae. Further deletion analysis identified a 102 bp region (between positions 2701 and
VulE and VulD cells. In addition to that V AB-23 directly activates transcription of S2A in the supplementary material). By highest read coverage) in the same region that had been identified locus over a relatively large region with a peak (i.e. region of the analysis detected strong binding of V AB-23::GFP to the defects of C. brenneri related nematode species 2809) that are strongly conserved between the four closely related species C. elegans, C. briggsae, C. remanei and C. brenneri (Fig. 4F). When this conserved 20 bp element was deleted in the minimal reporter (P_{smp-1-2604-2987}Δ_{2790-2809} in Fig. 4E), vulval expression was strongly reduced but not eliminated, similar to the reduction observed in vab-23(tm1945) mutants. Specifically, 22% of P_{smp-1-2604-2987}Δ_{2790-2809} L4 larvae showed NLS::GFP expression, whereas no expression was detected in 52% and asymmetric expression on one side of the vulval invagination in the remaining 25% of the cases (n=36).

To test whether the VAB-23 protein interacts directly with the smp-1 locus, we performed chromatin immunoprecipitation of VAB-23 followed by deep sequencing of the bound DNA fragments (ChIPseq). For this purpose, we used the translational vab-23::gfp fusion construct described previously (Pellegrino et al., 2009) and isolated the VAB-23::GFP protein-chromatin complexes with anti-GFP antibodies. Importantly, the vab-23::gfp transgene efficiently rescued the embryonic and vulval morphogenesis defects of vab-23(tm1945) mutants (Fig. 8A). The ChIPseq analysis detected strong binding of VAB-23::GFP to the smp-1 locus over a relatively large region with a peak (i.e. region of the highest read coverage) in the same region that had been identified by smp-1 promoter analysis (left-right arrow in Fig. 4E; see Fig. S2A in the supplementary material).

Taken together, our promoter and ChIPseq analysis indicated that VAB-23 directly activates transcription of smp-1 in the VulF, VulE and VulD cells. In addition to smp-1, we identified another 455 specific binding VAB-23 binding sites throughout the C. elegans genome, suggesting that VAB-23 controls the transcription of a large number of target genes (see Table S1 in the supplementary material and Materials and methods).

**VAB-23 regulates the expression of late VulE and VulF sub-fate-specific genes**

Next, we used various markers to determine whether early or late steps during cell fate specification are affected in vab-23(tm1945) mutants (Fig. S5). We examined the 1° fate specification using a P_{egl-17::yfp} reporter, which is predominantly expressed in the 1° cell lineage until the 4-cell stage (Pn.pxx). This early, 1°-specific expression of P_{egl-17::yfp} was unchanged or even slightly elevated in vab-23(tm1945) larvae, indicating proper 1° fate specification (Fig. 5A,B). To examine the VulE and VulF sub-fates, we used transcriptional P_{egl-26::gfp} and P_{lin-3::gfp} reporters, respectively (Chang et al., 1999; Hanna-Rose and Han, 2002). In vab-23(tm1945) mutants, 75% of L4 larvae (n=61) lacked P_{lin-3::gfp} expression in VulF cells, compared with 9% of wild-type larvae showing no expression at this stage (n=23; Fig. 5C,D). Similarly, a reduction of vab-23 function by RNAi caused a loss of P_{egl-26::gfp} expression in VulE in 61% of the animals (n=54; Fig. 5E,F). Moreover, VAB-23::GFP ChIPseq analysis detected a small but significant peak in the egl-26 5° regulatory region contained within the P_{egl-26::gfp} reporter (see Fig. S2B in the supplementary material), whereas no binding to the lin-3 locus was detected (data not shown).

LIN-3 EGF expression in VulF cells is necessary to specify the uv1 cell fate in the ventral uterine cells (Chang et al., 1999). Accordingly, vab-23(tm1945) mutants or RNAi-treated animals often had thick tissue blocking the connection between the vulval lumen and the uterus, suggesting a defect in uterine development (see Fig. S1B in the supplementary material). Finally, we examined the P_{egl-17::yfp} and P_{lin-3::gfp} reporters in L4 larvae to assay the specification of the 2° VulC and VulD sub-fates (Burdine et al., 1998; Inoue et al., 2002), but observed no obvious changes in their expression patterns (Fig. 5G-J). Thus, VAB-23 is not necessary for

---

**Fig. 3. VAB-23 regulates the formation of proper vulval cell contacts.** (A-F) Ventral views of AJM-1::GFP in wild-type (A,C,E) and vab-23(tm1945) (B,D,F) larvae at the Pn.p, Pn.pxx to Pn.pxxx stages. Arrows in D and F indicate abnormal cell contacts, and the arrowhead in F shows punctate AJM-1::GFP expression. Scale bar: 10 μm.
the specification of the 1° or 2° cell fates during vulval induction, but rather controls the patterning of the 1° VulF and VulE cells. Regulation of \textit{egl-26} transcription by VAB-23 might be direct, whereas regulation of \textit{lin-3} expression appears to be indirect.

**VAB-23 expression is regulated by EGFR/RAS/MAPK signaling during vulval induction and morphogenesis**

We analyzed the post-embryonic VAB-23 expression pattern using the translational VAB-23::GFP reporter described previously (Pellegrino et al., 2009). VAB-23::GFP was expressed in the AC, the vulval cells, in the ventral and dorsal uterine cells, the seam cells, the vulval muscle cells, a small cluster of unidentified tail cells, and some ventral cord neurons (Fig. 6 and data not shown). Vulval expression of VAB-23::GFP was observed predominantly in the 1° lineage beginning at the time of induction and persisting until adulthood (Fig. 6A-D and data not shown). Even though VAB-23::GFP was initially expressed at low levels in all VPCs, expression was downregulated in the 2° lineage during induction and persisted at low levels in the tertiary (3°) cells. VAB-23::GFP continued to be strongly expressed in the VulE and VulF cells of L4 larvae during vulval morphogenesis, although relatively weaker expression was also observed in VulC and VulD at this later stage (Fig. 6D, VulC and VulD nuclei are out of focus).

The upregulation of VAB-23 in the 1° cell and simultaneous downregulation in the 2° lineage suggested that \textit{vab-23} might be a target of the EGFR/RAS/MAPK signaling pathway. We therefore analyzed VAB-23::GFP expression in various mutants in which the activity of the EGFR/RAS/MAPK signaling pathway is changed. Diminished activation of LET-23 EGFR by a mutation in \textit{lin-3} (Hill and Sternberg, 1992) or through mislocalization via a \textit{lin-7} mutation (Simske et al., 1996) had two effects on VAB-23::GFP expression. First, expression in P6.p and its descendants was strongly reduced, and second, equally weak expression was observed in P5.p to P7.p.
and their descendants, probably owing to the loss of lateral inhibition from P6.p (Fig. 6E,F). By contrast, ectopic expression of VAB-23::GFP was induced in the pseudovulvae formed by P3.p, P4.p or P8.p descendants in animals carrying a constitutively active let-60 allele or by overexpressing the MAP kinase MPK-1 along with MEK-2 (Fig. 6G,H) (Beitel et al., 1990; Lackner and Kim, 1998). Lastly, we examined whether EGFR/RAS/MAPK signaling continues to regulate VAB-23::GFP expression after vulval induction. For this purpose, we used a temperature-sensitive allele of sos-1, which encodes the major guanine nucleotide exchange factor for LET-60 (Chang et al., 2000). We observed a reduced or absent expression of VAB-23::GFP in ~80% of sos-1(ts) worms that had been shifted to the restrictive temperature after vulval induction had occurred (Fig. 6J; see Fig. S2 in the supplementary material). We conclude that EGFR/RAS/MAPK signaling is first required to upregulate vab-23 expression during induction and later to maintain expression in the 1° lineage during morphogenesis.

vab-23 is a direct target of LIN-39 and CEH-20
LIN-39 performs at least two distinct functions during vulval development (Clark et al., 1993; Maloof and Kenyon, 1998; Shemer and Podbilewicz, 2002). First, LIN-39 represses the expression of the fusogen eff-1 to prevent the fusion of VPCs with the surrounding hypodermis. Second, lin-39 is required for the execution of the vulval cell lineage after the fates have been specified, because in eff-1(0); lin-39(0) double mutants the vulval cells fail to divide (Shemer and Podbilewicz, 2002). Because lin-39 expression is positively regulated by EGFR/RAS/MAPK signaling and vab-23 prevents eff-1-mediated cell fusions, we investigated whether vab-23 might function downstream of LIN-39 during vulval morphogenesis (Guerry et al., 2007; Maloof and Kenyon, 1998). For this purpose, the loss-of-function allele lin-39(n760) was examined together with the eff-1(ok1021) mutation, which suppresses the early VPC fusions. VAB-23::GFP expression was strongly reduced in the eff-1(ok1021); lin-39(n760) double mutants, but unchanged in eff-1(ok1021) single mutants (Fig. 6K,L and data not shown). In early L3 larvae at the Pn.px stage, VAB-23::GFP was absent or barely detectable in 89% of eff-1(ok1021); lin-39(n760) double mutants and strongly reduced in the remaining cases (n=26; Fig. 6K). Also, at the time of toroid formation, eff-1(ok1021); lin-39(n760) animals showed, on average, a fourfold decrease in the intensity of VAB-23::GFP expression with 63% of the animals showing no or strongly reduced expression (n=27; Fig. 6L). In the remaining 37% of cases, VAB-23::GFP expression persisted albeit at a reduced level, indicating that late VAB-23 expression is partially regulated by a LIN-39-independent pathway. Similarly, a reduction-of-function mutation in ceh-20, which encodes a LIN-39 co-factor of the Extradenticle/PBX family (Yang et al., 2005), resulted in reduced VAB-23::GFP expression during the early (Pn.p to Pn.ppx) stages of vulval development, whereas the late (Pn.pxxx stage) expression was not affected by ceh-20 (Fig. 6M,N).

Next, we determined whether vab-23 is a transcriptional target of LIN-39. We aligned the vab-23 5’ regulatory regions from C. elegans and the closely related nematodes C. briggsae and C. remanei and searched for conserved HOX/PBX consensus binding motifs, such as TGA(T/N)NAT (Cui and Han, 2003; Koh et al., 2002). Two conserved putative HOX/PBX binding sites were present in a 72 bp region located 1.3 kb upstream of the vab-23 translation initiation site (Fig. 7A, region II in 7B). Using a functional LIN-39::GFP reporter (Szabo et al., 2009), we detected strongest binding to region II, which contains the HOX/PBX sites, whereas regions I and III showed weaker binding relative a probe containing the HOX/PBX sites in ChIP experiments. Using two conserved HOX/PBX sites (Hox-mut) eliminated expression in L4 larvae (Hox-mut, n=27; Fig. 6L). In the remaining 37% of cases, VAB-23::GFP expression persisted albeit at a reduced level, indicating that late VAB-23 expression is partially regulated by a LIN-39-independent pathway. Similarly, a reduction-of-function mutation in ceh-20, which encodes a LIN-39 co-factor of the Extradenticle/PBX family (Yang et al., 2005), resulted in reduced VAB-23::GFP expression during the early (Pn.p to Pn.ppx) stages of vulval development, whereas the late (Pn.pxxx stage) expression was not affected by ceh-20 (Fig. 6M,N).

Finally, we observed a direct interaction between LIN-39 and the region containing the HOX/PBX sites in ChiP experiments. Using a functional LIN-39::GFP reporter (Szabo et al., 2009), we detected strongest binding to region II, which contains the HOX/PBX sites, whereas regions I and III showed weaker binding relative a probe in the 3’UTR used as internal reference (Fig. 7B,E) (see Materials and methods). It should be noted that the ChiPseq experiments performed by the modENCODE consortium (Celniker et al., 2009)
detected binding of LIN-39::GFP to the same region in the \textit{vab-23} gene that we identified in our analysis. Taken together, our data indicate that LIN-39 directly regulates \textit{vab-23} expression by interacting with the conserved HOX/PBX sites in the 5' regulatory region.

**LIN-39-induced expression of VAB-23 is required for toroid formation**

Finally, we examined whether the early \textit{vab-23} expression induced by LIN-39 and CEH-20 is necessary for toroid formation during the subsequent morphogenesis. For this purpose, we investigated...
toroid formation in vab-23(tm1945) mutants carrying the vab-23 Hox-mut transgene to eliminate the early but not the late VAB-23::GFP expression (Fig. 7C). The vab-23 Hox-mut transgene rescued the embryonic lethality of vab-23, allowing 36% of tm1945 homozygotes to survive to adulthood (n=359), which is comparable to the rescue obtained with wild-type vab-23 transgenes (Pellegrino et al., 2009).

VAB-23 is essential for multiple aspects of vulval morphogenesis
We propose the following model for the regulation of toroid formation by VAB-23 (Fig. 9). At the time of vulval induction, inductive signaling induces LIN-39 expression via inactivation of the LIN-1 ETS transcriptional repressor (Guerry et al., 2007; Tan et al., 1998; Wagmaister et al., 2006). LIN-39 and its co-factor CEH-20 then activate vab-23 transcription in the 1° lineage, and VAB-23 upregulates SMP-1 expression to guide the interactions between contralateral pairs of vulval cells (Dalpe et al., 2005). Later during morphogenesis, CEH-20 is replaced by another, unknown LIN-39 co-factor (Takacs-Vellai et al., 2007). LIN-39 and this new co-factor CEH-20 then activate vab-23 transcription in the 1° lineage, and VAB-23 upregulates SMP-1 expression to guide the interactions between contralateral pairs of vulval cells (Dalpe et al., 2005). Later during morphogenesis, CEH-20 is replaced by another, unknown LIN-39 co-factor (Takacs-Vellai et al., 2007). LIN-39 and this new co-factor maintain the expression of VAB-23 in cooperation with the EGFR/RAS/MAPK pathway. The late VAB-23 expression is required both for the repression of EFF-1-mediated vulval cell fusion as well as for the expression of the VulE and VulF specific genes lin-3 and egl-26.
Similar functions of VAB-23 during embryo and vulval morphogenesis
Loss of VAB-23 function in the embryo causes the formation of ectopic cell contacts between ipsilateral ventral epidermal cells, resulting in defective ventral closure (Pellegrino et al., 2009). Interestingly, VAB-23 plays an analogous role during toroid formation by guiding cells towards the vulval midline, which forms an axis of symmetry for the developing organ (Fig. 1A). Similar to the embryonic phenotype, cells of the same sub-fate cannot form contacts across the vulval midline, resulting in the malformation or absence of toroids. The vab-23 phenotype is in part due to reduced SMP-1 expression. To our knowledge, VAB-23 is the first example of a Semaphorin regulator, which links vulval morphogenesis via LIN-39 to the EGFR/RAS/MAPK pathway. However, both the vulval and embryonic morphogenesis defects of smp-1 mutants are less penetrant and generally milder than the defects observed in vab-23 mutants, suggesting that VAB-23 controls the expression of additional effectors of morphogenesis acting in parallel with SMP-1 (Dalpe et al., 2005). For example, the Rac GTPase MIG-2 and the guanine-nucleotide exchange factor UNC-73 are both required for vulval cell migration in parallel with the SMP-1/PLX-1 pathway (Dalpe et al., 2005; Kishore and Sundaram, 2002; Lundquist et al., 2001; Zipkin et al., 1997). Thus, VAB-23 might regulate multiple target genes performing diverse functions during vulval morphogenesis.

We have previously shown that during embryogenesis VAB-23 acts from the underlying neuroblasts in a cell non-autonomous manner to guide the ventral epidermal cells (Pellegrino et al., 2009). Although VAB-23 expression was detected predominantly in the 1° cell lineage (except for the weaker, later expression in VulC and VulD) and VAB-23 only controls the patterning of VulE and VulF cells, vab-23 mutants display defects in the formation of all toroids. Even the distal-most VulA cells often failed to migrate towards the midline and form a toroid. It thus appears that during vulval morphogenesis, VAB-23 guides vulval cells in a cell non-autonomous manner, possibly by regulating the production of multiple secreted cues.

VAB-23 regulates various aspects of vulval and uterine morphogenesis
In addition to controlling the formation of homotypic cell contacts, VAB-23 inhibits cell fusions at a later stage of vulval morphogenesis. Ectopic expression of the fusogen EFF-1 in vab-23 mutants is likely to be responsible for these abnormal fusions, as loss of eff-1 function almost completely restored the normal number of cell junctions. LIN-39 acts upstream of eff-1 to negatively regulate VPC cell fusions prior to vulval induction by inducing the GATA factors egl-18 and elt-6 (Koh et al., 2002; Shemer and Podbilewicz, 2002). It is possible that LIN-39 represses late vulval cell fusions indirectly by inducing vab-23 expression. Interestingly, the cell lineage is unchanged in vab-23 mutants, whereas the VPCs fail to proliferate in lin-39(0) single or egl-18(0); elt-6(0) double mutants. Thus, cell proliferation is controlled by LIN-39 through a VAB-23-independent branch of the cell fate execution pathway (Fig. 9).
Besides eff-1 and smp-1, vab-23 also regulates the later expression of egl-26 and lin-3 in VulE and VulF cells, respectively. Proper patterning of the VulE and VulF cells is necessary for vulval morphogenesis and uterine development. egl-26 is required to maintain the shape of the VulF toroid after the AC has retracted (Estes and Hanna-Rose, 2009; Estes et al., 2007; Hanna-Rose and Han, 2002), and LIN-3 is required for the specification of the uv1 fate by activating LET-23 signaling in the ventral uterine cells. Therefore, the reduced LIN-3 expression in VulF cells probably accounts for the uterine-seam cell defects in vab-23 mutants (Chang et al., 1999).

Based on our previous data it was not possible to distinguish whether VAB-23 acts as a transcriptional or post-transcriptional regulator of gene expression (Pellegrino et al., 2009). Here, we show that VAB-23 binds to regulatory regions in two of its target genes: to the first intron of smp-1 and, more weakly, to the 5′ regulatory region of egl-26. Finally, we have detected specific binding of VAB-23 to a relatively large number of sites throughout the genome. Although we could not extract an obvious consensus sequence from the VAB-23 binding sites identified by ChiPseq, the palindromic TTCNGAA motif in the center of the conserved regulatory element in smp-1 is around sevenfold over-represented in the VAB-23 binding sites. The TTCNGAA motif was found 286 times versus the 40 times that would be expected in a random distribution.) These results indicate that VAB-23 acts as a transcription factor that controls the expression of multiple target genes regulating various aspects of epidermal morphogenesis.

**Vulval cell fate specification and morphogenesis are linked processes**

During the specification of segment-specific structures and individual organs, the hox genes are thought to regulate a large number of target genes in a context-specific manner (Hueber and Lohmann, 2008). However, only a relatively small number of hox targets that are directly involved in morphogenesis, the so-called ‘realisators’, have been identified. Furthermore, the conserved EGFR/RAS/MAPK, Notch and WNT signaling pathways control various aspects of epithelial morphogenesis in *Drosophila* and *C. elegans*, but their transcriptional targets during morphogenesis are poorly characterized (Galindo et al., 2005; O’Keefe et al., 2007; Rasmussen et al., 2008).

vab-23 reveals how EGFR/RAS/MAPK signaling coordinates via LIN-39 different aspects of organogenesis after the cell fate decisions have been made. *let-60* has previously been reported to regulate cell migration and fusions (Shemer et al., 2000), and *smp-1* is expressed in the pseudovulvae of *let-60(ge)* worms (Dalpe et al., 2005), suggesting that SMP-1 expression is indeed linked to EGFR/RAS/MAPK signaling. Moreover, conditional inactivation of *sos-1* indicates that continuous EGFR/RAS/MAPK signaling after induction is necessary to maintain VAB-23 expression in the 1° cell lineage. VAB-23 thus acts as a link between the cell fate specification and execution pathways. Although LIN-39 mediates most of the regulation of VAB-23 by the EGFR/RAS/MAPK pathway, LIN-39 clearly performs additional, VAB-23-independent functions. LIN-39 prevents VPC fusion before and promotes cell proliferation after vulval induction (Shemer and Podbilewicz, 2002). Owing to the lack of cell proliferation, it is difficult to determine whether any vulval sub-fates are specified and, thus, whether proper cell contacts are formed in *lin-39(0)* mutants. However, the few (usually one to four) toroids that are formed in *lin-39(0); eff-1(0)* double mutants appear disorganized (Shemer and Podbilewicz, 2002).

In summary, this study shows that vulval fate specification and morphogenesis are more tightly linked than previously thought, as there exists a considerable temporal overlap between these two phases of organogenesis. Continuous signaling by the EGFR/RAS/MAPK pathway after vulval fate specification might be necessary to induce and maintain the expression of key regulators of epidermal morphogenesis. The identification of additional late EGFR/RAS/MAPK targets might shed light on the regulatory network controlling vulval morphogenesis.

**Acknowledgements**

We thank the members of our group for critical discussion and J.M. Escobar and M. Walser for comments on the manuscript. We are also grateful to the Foundation for the Functional Genomics Center Zurich for SOLiD sequencing, and to Andrew Fire for vectors.

**Funding**

This work was supported by a postgraduate scholarship from the University of Melbourne, the Dr Sue Newton Research Award, and a travel grant from the National Health and Medical Research Council Parasitology Research Network to M.W.P.; the Australian Academy of Science; a grant from the Forschungskredit of the University of Zürich to S.F.; a grant from the Australian Research Council to R.B.G.; and a grant from the Swiss National Science Foundation to A.H.

**Competing interests statement**

The authors declare no competing financial interests.

**Supplementary material**

Supplementary material for this article is available at http://dev.biologists.org/lookup/suppl/doi:10.1242/dev.071951/-/DC1

**References**


